

Australian Government

Department of Industry, Science and Resources National Measurement Institute

# Proficiency Test Final Report AQA 22-14 PFAS in Biota and Food

January 2023

AQA 22-14 PFAS in Biota and Food

# ACKNOWLEDGMENTS

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I would like to thank the management and staff of the participating laboratories for supporting the study. It is only through widespread participation that we can provide an effective service to laboratories.

The assistance of the following NMI staff members in the planning, conduct and reporting of the study is acknowledged.

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## SUMMARY

AQA 22-14 PFAS in Biota and Food commenced in August 2022. Twenty laboratories registered to participate, with one laboratory requesting two sets of samples to be analysed independently. Twenty participants submitted results by the due date.

The sample set consisted of one spiked prawn sample (Sample S1) and one spiked carrot sample (Sample S2). The per- and polyfluoroalkyl substances (PFAS) analytes assessed in this study were: PFBS, PFPeS, PFHxS, PFHpS, PFOS, PFNS, PFDS, PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFNA, PFDA, PFUdA, PFTrDA, PFOSA, MeFOSA, EtFOSA, 6:2 FTS, GenX, ADONA, 9C1-PF3ONS and 11C1-PF3OUdS.

Of 811 possible results, 597 (74%) were numeric results. One hundred and eight results were a 'less than' value (< x) or Not Reported (NR), and 106 results were Not Tested (NT).

The assigned values for all scored analytes were the robust averages of participants' results, and associated uncertainties were estimated from the robust standard deviations. The results from Laboratories **3**, **8**, **10**, **16** and **20** in Sample S1, and from Laboratory **16** in Sample S2, were consistently lower than the spiked value by around the same factor for all analytes, an indication of laboratory bias. To avoid unfair scoring, these results were excluded from robust average calculations as it will bias low the assigned value; they were also excluded from the calculation of all summary statistics.

**Traceability**: The consensus of participants' results is not traceable to any external reference, so although expressed in SI units, metrological traceability has not been established.

The outcomes of the study were assessed against the aims as follows:

• Assess the ability of participants to correctly identify PFAS in biota and food matrices.

Of the participants who returned results, 16 participants were sent both matrices, three participants were sent prawn only, and one participant was sent carrot only. Laboratories 7, 12, 13 and 17 reported numeric results for all 39 scored analytes across both matrices.

Ten participants did not report results for analytes that they tested for and were spiked into the samples (total of 53 results), while seven participants reported analytes that were not spiked into the samples (total of 11 results).

• Compare the performances of participants and assess their accuracy in the measurement of PFAS in biota and food matrices.

Of 528 *z*-scores, 453 (86%) returned  $|z| \le 2.0$ , indicating a satisfactory performance.

Of 528  $E_n$ -scores, 374 (71%) returned  $|E_n| \le 1.0$ , indicating agreement of the participant's result with the assigned value within their respective expanded uncertainties.

Laboratories 7 and 12 returned satisfactory *z*-scores for all scored analytes (39). No participant returned satisfactory  $E_n$ -scores for all analytes of interest in this study.

• Evaluate participants' test methods for PFAS in biota and food analysis.

Participants used a variety of methods for extraction and analysis. The most popular for this study was homogenisation as pretreatment, followed by alkaline digestion with basified methanol and solid-phase extraction clean-up, and then analysis using LC-MS/MS or LC-QQQ.

In this study, extraction with basified methanol, and long extraction times (> 8 h) were more effective, giving results with higher recoveries with respect to the spiked values.

Participants should take care to avoid any potential dilution, standard preparation, or similar errors in their analyses.

• Develop the practical application of traceability and measurement uncertainty.

Of 597 numeric results for spiked analytes, 580 (97%) were reported with an associated expanded measurement uncertainty.

Although it is a requirement of ISO/IEC 17025 that laboratories have procedures to estimate the uncertainty since 2017, a large number of laboratories are still reporting potentially unrealistically small or large relative uncertainties for routine PFAS. The magnitude of the reported measurement uncertainties for spiked analytes in this study was within the range 1.8% to 100% of the reported value. Additionally, some laboratories are still reporting numeric estimates of uncertainties for non-numeric results.

• Compare the performance of participants with their past performance.

NMI has been conducting PFAS in biota and food PT studies since 2016.

The proportion of total possible results being reported by participants as numeric results has remained fairly consistent, even with the significantly increased number of PFAS analytes over the last few studies, indicating that participants have the capacity to analyse a wide range of PFAS at relevant mass fractions.

Proportions of satisfactory z-scores and  $E_n$ -scores have also remained relatively consistent over this period, though for this study there was a slight decrease as compared to the previous year's study.

• Produce materials that can be used in method validation and as control samples.

The test samples of this proficiency study are homogeneous and are well characterised. Surplus samples are available for purchase from NMI and can be used for quality control and method validation purposes.

# 1 INTRODUCTION

## 1.1 NMI Proficiency Testing Program

The National Measurement Institute (NMI) is responsible for Australia's national measurement infrastructure, providing a range of services including a chemical proficiency testing program.

Proficiency testing (PT) is the 'evaluation of participant performance against pre-established criteria by means of interlaboratory comparison'.<sup>1</sup> NMI PT studies target chemical testing in areas of high public significance such as trade, environment, law enforcement and food safety. NMI offers studies in:

- pesticide residues in soil and water, fruit, vegetables and herbs;
- petroleum hydrocarbons in soil and water;
- per- and polyfluoroalkyl substances (PFAS) in soil, water, biota and food;
- inorganic analytes in soil, water, filters, food and pharmaceuticals;
- controlled drug assay, drugs in wipes and clandestine laboratory; and
- allergens in food.

#### 1.2 Study Aims

The aims of the study were to:

- assess the ability of participants to correctly identify PFAS in biota and food matrices.
- compare the performances of participants and assess their accuracy in the measurement of PFAS in biota and food matrices;
- evaluate participants' test methods for PFAS in biota and food analysis;
- develop the practical application of traceability and measurement uncertainty;
- compare the performance of participants with their past performance; and
- produce materials that can be used in method validation and as control samples.

The choice of test method was left to the participating laboratories.

## 1.3 Study Conduct

The conduct of NMI proficiency tests is described in the NMI Study Protocol for Proficiency Testing.<sup>2</sup> The statistical methods used are described in the NMI Chemical Proficiency Testing Statistical Manual.<sup>3</sup> These documents have been prepared with reference to ISO/IEC 17043:2010,<sup>1</sup> and The International Harmonized Protocol for the Proficiency Testing of Analytical Chemistry Laboratories.<sup>4</sup>

NMI is accredited by the National Association of Testing Authorities, Australia (NATA) to ISO/IEC 17043 as a provider of proficiency testing schemes. This study falls within the scope of NMI's accreditation.

# 2 STUDY INFORMATION

# 2.1 Study Timetable

The timetable of the study was:

Invitations sent	8/08/2022
Samples sent	30/08/2022
Results due	26/10/2022
Interim report	1/11/2022

The results due date was extended to accommodate sample delivery delays to some international participants.

## 2.2 Participation and Laboratory Code

Twenty laboratories registered to participant, with one laboratory requesting two sets of samples to be analysed independently. All participants were assigned a confidential laboratory code number for this study. Twenty participants submitted results by the due date.

## 2.3 Selection of PFAS Analytes and Test Material Preparation

Participants were provided with a list of potential PFAS analytes that were spiked into the study's samples, as presented in Table 1.

PFBS	PFTrDS	PFDoA	N-EtFOSE
PFPeS	PFBA	PFTrDA	4:2 FTS
PFHxS	PFPeA	PFTeDA	6:2 FTS
PFHpS	PFHxA	PFOSA	8:2 FTS
PFOS	PFHpA	MeFOSA	10:2 FTS
PFNS	PFOA	EtFOSA	GenX
PFDS	PFNA	MeFOSAA	ADONA
PFUdS	PFDA	EtFOSAA	9C1-PF3ONS
PFDoS	PFUdA	MeFOSE	11Cl-PF3OUdS

Table 1 Potential Spiked PFAS Analytes

The two samples were prepared in August 2022. Care was taken to avoid any PFAS contamination during sample preparation. The prepared samples were:

- Sample S1: Prawn (5 g portions) spiked with 21 different PFAS analytes.
- Sample S2: Carrot (30 g portions) spiked with 20 different PFAS analytes.

Details of spiked analytes and values are presented in Table 2.

Table 2 Spiked Values of Test Samples

Analyte	Sample S1 Prawn (µg/kg)	Sample S2 Carrot (µg/kg)
PFBS	0.399	0.891
PFPeS	4.65	7.47
PFHxS*	1.89	6.61
PFHpS	2.00	3.00
PFOS*	4.77	2.12

Analyte	Sample S1 Prawn (µg/kg)	Sample S2 Carrot (µg/kg)
PFNS	11.5	1.72
PFDS	Not Spiked	6.80
PFBA	2.96	1.19
PFPeA	1.13	2.20
PFHxA	5.31	7.45
PFHpA	7.54	1.50
PFOA	7.92	1.20
PFNA	0.503	2.31
PFDA	0.902	9.47
PFUdA	1.21	Not Spiked
PFTrDA	8.17	Not Spiked
PFOSA	4.46	4.95
MeFOSA	4.99	4.99
EtFOSA	3.99	3.99
6:2 FTS	Not Spiked	1.89
GenX	Not Spiked	11.1
ADONA	5.64	14.0
9C1-PF3ONS	14.4	Not Spiked
11Cl-PF3OUdS	4.70	Not Spiked

\* Participants were requested to report both linear isomers and total value. The samples were spiked with linear standards only for these analytes.

Further sample preparation details can be found in Appendix 1.

## 2.4 Homogeneity and Stability of Test Materials

The process used to prepare, store and dispatch the test samples has been demonstrated to produce sufficiently homogeneous and stable samples for previous NMI PFAS in biota and food PT studies, for timeframes similar to that of this study.

To consider possible instability of the samples, the results returned by participants were compared to the spiked values. Robust averages for scored analytes were within 69% to 93% and 76% to 111% of the spiked values for Samples S1 and S2 respectively, which were similar to values observed in previous NMI PFAS in biota and food PT studies and provides support for the stability of these analytes.

Additionally, homogeneity and stability testing was conducted on Sample S2 carrot. The samples were demonstrated to be sufficiently homogeneous and stable for the evaluation of participants' performance in this study.

Further details on the homogeneity and stability assessment of the samples are given in Appendix 2.

## 2.5 Test Material Storage and Dispatch

After preparation, the test material were dispensed into sample tubes, labelled and shrink-wrapped. Prior to sample dispatch, all samples were stored frozen at -80 °C.

Samples were packed into insulated polystyrene foam boxes with cooler bricks and sent by courier on 30 August 2022.

The following items were packaged with the samples:

- a covering letter which included a description of the test samples and instructions for participants; and
- a form for participants to confirm the receipt and condition of the samples.

An Excel spreadsheet for the electronic reporting of results was emailed to all participants.

#### 2.6 Instructions to Participants

Participants were instructed as follows:

- Quantitatively analyse the samples for PFAS, using your routine test method and report results in units of  $\mu g/kg$  on an as received basis.
  - For PFAS analytes that contain linear and branched isomers, report total (the sum of linear and branched isomers).
  - For PFOS and PFHxS you are asked to report total (the sum of linear and branched isomers) and linear (the linear isomers only).
- Report results using the electronic results sheet emailed to you.
- For each analyte report a single result expressed as if reporting to a client (i.e. corrected for recovery or not, according to your standard procedure, but state if results are corrected on the result sheet). This figure will be used in all statistical analysis in the study report.
- For each analyte report the associated expanded measurement uncertainty as  $\mu g/kg$  (e.g.  $0.50 \pm 0.02 \mu g/kg$ ), if determined.
- No limit of reporting has been set for this study. Report results as you would to a client, applying the limit of reporting of the method used for analysis.
- Report any listed analyte not tested as NT.
- Please complete the method details and report the basis of your uncertainty estimates as required by the results sheet.
- If determined, report your internal standard percentage recovery. This will be presented in the report for information only.
- Return the completed results sheet by email (proficiency@measurement.gov.au) by 7 October 2022.

Due to sample delivery delays to some international participants caused by customs clearance issues, the results due date was extended to 26 October 2022 for all participants.

#### 2.7 Interim Report

An interim report was emailed to all participants on 1 November 2022.

# **3 PARTICIPANT LABORATORY INFORMATION**

## 3.1 Participants' Test Methods

Participants were requested to provide information about their methodology. Responses are presented in Appendix 4.

# 3.2 Basis of Participants' Measurement Uncertainty Estimates

Participants were requested to provide information about their basis of measurement uncertainty (MU). Responses are presented in Tables 3 and 4. Responses may be modified so that the participant cannot be identified.

Lab.	Approach to Estimating	Information Sources	Guide Document	
Code	MU	Precision	Method Bias	for Estimating MU
1	Standard deviation of replicate analyses multiplied by 2 or 3Control samples - SS Duplicate analysisCRM Instrument calibration 		NATA GAG Estimating and Reporting MU	
2	standard deviation of triplicate measurements	Standard deviation f	rom PT studies only	
3	Professional judgment	Standard deviation f	from PT studies only	
4	Top Down - precision and estimates of the method and laboratory bias	Control samples - CRM Duplicate analysis Instrument calibration	CRM Laboratory bias from PT studies Recoveries of SS	NMI Uncertainty Course
5	Standard deviation of replicate analyses multiplied by 2 or 3Control samples - SSRecoveries of SS		ISO/GUM	
6	Bottom Up (ISO/GUM, fish bone/cause and effect diagram) Duplicate analysis CRM		ISO/GUM	
7	Top Down - precision and estimates of the method and laboratory biasControl samples Duplicate analysis Instrument calibrationCRM Instrument calibration PT studies Recoveries of SS Standard purity		NATA GAG Estimating and Reporting MU (replaced Technical Note 33)	
8	Top Down - precision and estimates of the method and laboratory bias	Control samples - SS Duplicate analysis Instrument calibration	CRM Instrument calibration Recoveries of SS	ISO/GUM
9	Top Down - precision and estimates of the method and laboratory biasControl samples - SS Duplicate analysisInstrument calibration Recoveries of SS Standard purity		Eurachem/CITAC Guide	
10	Standard deviation of replicate analyses multiplied by 2 or 3	replicate analyses Control samples - SS CRM		NMI Uncertainty Course
11	Top Down - precision and estimates of the method and laboratory biasControl samples - RM Duplicate analysisCRM Laboratory bias from PT studies Recoveries of SS		Nordtest Report TR537	

Table 3 Basis of Participants' Uncertainty Estimate

Lab.	Approach to Estimating	Information Sources	for MU Estimation*	Guide Document
Code	MU	Precision	Method Bias	for Estimating MU
12	Standard deviation of replicate analyses multiplied by 2 or 3	Control samples		ISO/GUM
	Standard deviation of	Standard deviation f	from PT studies only	
13	replicate analyses multiplied by 2 or 3	Control samples - SS Duplicate analysis Instrument calibration	Instrument calibration Recoveries of SS	Eurachem/CITAC Guide
15	Top Down - precision and estimates of the method and laboratory bias	Control samples - SS	Recoveries of SS	NATA - Estimating and reporting MU of chemical test results.
16	Top Down - precision and estimates of the method and laboratory bias	Control samples - SS Duplicate analysis Instrument calibration	Instrument calibration Laboratory bias from PT studies Recoveries of SS	AQS Baden- Württemberg
17	Top Down - precision and estimates of the method and laboratory bias	Control samples - SS Duplicate analysis Instrument calibration	Laboratory bias from PT studies Recoveries of SS	ISO/GUM
18	Top Down - precision and estimates of the method and laboratory bias	Control samples - RM Duplicate analysis		Eurachem/CITAC Guide
19	Standard deviation of replicate analyses multiplied by 2 or 3	Control samples - SS		Statistics and Chemometrics for Analytical Chemistry, Miller and Miller, 5th Edition
20	Professional judgment	Duplicate analysis Instrument calibration	CRM Instrument calibration Laboratory bias from PT studies Recoveries of SS	Internal document
21	Bottom Up (ISO/GUM, fish bone/cause and effect diagram)	Duplicate analysis	CRM	

\* SS = Spiked Samples, RM = Reference Material, CRM = Certified Reference Material

# Table 4 Uncertainty Estimate Additional Comments

Lab. Code	Uncertainty Estimate Comments	
1	Recovery and uncertainty data given for analytes at method limit of reporting.	
12	The expanded measurement uncertainty values were calculated using a coverage factor (K) value of 2.00 and at the 95% confidence limit.	
13	Uncertainty calculated as 3xSD of replicate analysis.	
19	Measurement Uncertainty (U) estimated from the standard deviation (u) of replicate recovery samples using the expression $U = 2 x u$ . Procedure as set out in Statistics and Chemometrics for Analytical Chemistry, Miller and Miller, 5th Edition	

## 3.3 Participants' Comments

Participants were invited to make comments on the samples, this PT study, or suggestions for future studies. Such feedback may be useful in improving future studies. Participants' comments are presented in Table 5, along with the study coordinator's response where applicable. Responses may be modified so that the participant cannot be identified.

Lab. Code	Sample	Participant's Comments	Study Coordinator's Response
	S1	Extra Compounds Detected < LOR : PFBA, PFPeA, PFOA, PFUdA, N-EtFOSA, N-MeFOSA	
1	S2	Extra Compounds Detected < LOR : PFBA Compounds not in the method scope that were detected: PFDS ca. 6 ug/kg	
	All	Methodology: In this method the linear standards are used to quantify both the linear as well as the branched isomers.	
2	S1	shrimp had gone through significant decomposition due to being in thawed state for prolonged period of time	Stability testing conducted in previous PT studies for PFAS in prawn showed that analytes were sufficiently stable for at least two months at room temperature. <sup>5</sup>
All		We use a technical mixture for PFOS as an analytical standard. It appears there is only linear PFOS in this sample, this may result in some bias compared to using just a linear isomer standard	
3	S1	Linear isomers reported only.	
12	All	The sample was received at a temperature of 20.4°C; which was above the laboratory method recommended sample storage temperature (less than or equal to 6°C). Methodology: Isotopically labelled surrogate standards were spiked into the sample prior to extraction	Please see response to Laboratory 2. Additionally, in this study, stability testing was conducted for Sample S2 carrot, and analytes were found to be sufficiently stable (see Appendix 2).
13	All	Please send more of the biota sample next time if possible. 5g does not leave much room for re- doing the experiment if you use 1 g for each replicate	The amount per sample has been selected to balance the preparation method requirements while allowing participants to perform some replicates, with most participants using around 1 g per analysis. Participants can also order additional samples if required for their analyses.
15	All	All linear and branched present have been reported although some branched peaks are not confirmed by traceable standards.	
20	All	A recovery standard was used to calculate the recovery of the internal standard. The samples were at customs for 4 weeks. There was no information about the storage conditions of the sample. The samples smelled slightly spoiled and also showed visual conspicuities.	Please see responses to Laboratories 2 and 12.

Table 5 Participants' Comments

# 4 PRESENTATION OF RESULTS AND STATISTICAL ANALYSIS

## 4.1 Results Summary

Participant results are presented in Tables 6 to 50 with summary statistics: robust average, median, mean, number of numeric results (N), maximum (Max), minimum (Min), robust standard deviation (Robust SD) and robust coefficient of variation (Robust CV), as well as other estimates of analyte mass fraction. Bar charts of results and performance scores are presented in Figures 2 to 46. An example chart with interpretation guide is shown in Figure 1.

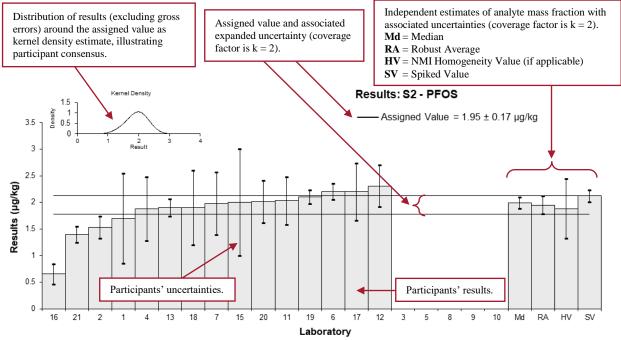


Figure 1 Guide to Presentation of Results

# 4.2 Outliers, Gross Errors and Results Excluded from Robust Average Calculations

Outliers were results less than 50% and greater than 150% of the robust average, and these were removed before the calculation of the assigned value.<sup>3,4</sup> Gross errors were extreme outliers or obvious blunders e.g. results reported with incorrect units or basis, or for a different analyte or sample, and such results were removed for the calculation of all summary statistics.<sup>3,4</sup>

The results from Laboratories **3**, **8**, **10**, **16** and **20** in Sample S1, and from Laboratory **16** in Sample S2, were consistently lower than the spiked value by around the same factor for all analytes. This is an indication of laboratory or method bias. To avoid unfair scoring, these results were excluded from robust average calculations as it will bias low the assigned value; they were also excluded from the calculation of all summary statistics.

## 4.3 Assigned Value

The assigned value is defined as the 'value attributed to a particular property of a proficiency test item'.<sup>1</sup> In this PT study, the property is the mass fraction of analytes in the samples. Assigned values in this study were the robust averages of participants' results and the expanded uncertainties were estimated from the associated robust SDs (Appendix 3).

# 4.4 Robust Average and Robust Between-Laboratory Coefficient of Variation

The robust averages and associated expanded MUs, and robust CVs (a measure of the variability of participants' results) were calculated using the procedure described in ISO 13528:2022.<sup>6</sup>

# 4.5 Performance Coefficient of Variation (PCV)

The performance coefficient of variation (PCV) is a fixed measure of the between-laboratory variation that in the judgement of the study coordinator would be expected from participants given the levels of analytes present. The PCV is not the CV of participants' results; it is set by the study coordinator and is based on the mass fraction of the analytes and experience from previous studies, and is supported by mathematical models such as the Thompson-Horwitz equation.<sup>7</sup> By setting a fixed and realistic value for the PCV, a participant's performance does not depend on the performance of other participants and can be compared from study to study.

# 4.6 Target Standard Deviation for Proficiency Assessment

The target standard deviation for proficiency assessment ( $\sigma$ ) is the product of the assigned value (*X*) and the PCV, as presented in Equation 1.

$$\sigma = X \times PCV \qquad Equation \ 1$$

## 4.7 *z*-Score

For each participant's result, a *z*-score is calculated according to Equation 2.

$$z = \frac{(\chi - X)}{\sigma} \qquad Equation 2$$

where:

- z is z-score
- $\chi$  is a participant's result
- X is the assigned value
- $\sigma$  is the target standard deviation from Equation 1

For the absolute value of a *z*-score:

- $|z| \le 2.0$  is satisfactory;
- 2.0 < |z| < 3.0 is questionable; and
- $|z| \ge 3.0$  is unsatisfactory.

To account for potential low bias in consensus values due to inefficient methodologies, scores may be adjusted for a 'maximum acceptable result'. Additional information is given in Section 6.3.

## 4.8 *E<sub>n</sub>*-Score

The  $E_n$ -score is complementary to the *z*-score in assessment of laboratory performance.  $E_n$ -score includes measurement uncertainty and is calculated according to Equation 3.

$$E_n = \frac{(\chi - X)}{\sqrt{U_{\chi}^2 + U_X^2}} \qquad Equation 3$$

where:

 $E_n$  is  $E_n$ -score

- $\chi$  is a participant's result
- X is the assigned value
- $U_{\chi}$  is the expanded uncertainty of the participant's result
- $U_X$  is the expanded uncertainty of the assigned value

For the absolute value of an  $E_n$ -score:

- $|E_n| \le 1.0$  is satisfactory;
- $|E_n| > 1.0$  is unsatisfactory.

# 4.9 Traceability and Measurement Uncertainty

Laboratories accredited to ISO/IEC 17025 must establish and demonstrate the traceability and measurement uncertainty associated with their test results.<sup>8</sup>

Guidelines for quantifying uncertainty in analytical measurement are described in the Eurachem/CITAC Guide.<sup>9</sup>

# 5 TABLES AND FIGURES

Table 6

# Sample Details

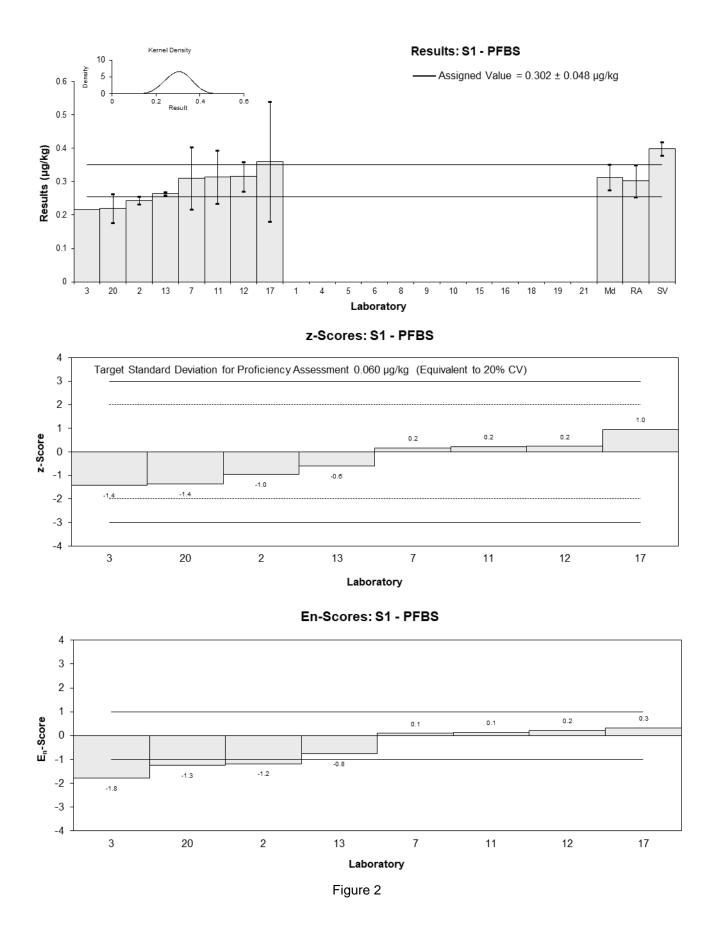
Sample No.	S1
Matrix	Prawn
Analyte	PFBS
Unit	µg/kg

## **Participant Results**

Lab. Code	Result	Uncertainty	Rec	z	En
1	< 2	1	106		
2	0.244	0.011	94	-0.96	-1.18
3**	0.216	NR	NR	-1.42	-1.79
4	<0.5	NR	76		
5	NS	NS	NS		
6	<1	NR	NT		
7	0.311	0.093	88	0.15	0.09
8	<0.5	NR	99		
9	<0.5	NR	98		
10	<1	NR	NR		
11	0.315	0.08	94.0	0.22	0.14
12	0.316	0.0442	90.3	0.23	0.21
13	0.265	0.005	105	-0.61	-0.77
15	<1	NR	82		
16	< 0.1	NR	138		
17	0.36	0.18	89	0.96	0.31
18	<1	NR	135		
19	< 1.0	NR	104		
20**	0.220	0.044	73	-1.36	-1.26
21	<1	NR	NT		

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	0.302	0.048
Spike Value	0.399	0.020
Robust Average	0.302	0.048
Median	0.313	0.038
Mean	0.302	
Ν	6	
Мах	0.36	
Min	0.244	
Robust SD	0.047	
Robust CV	16%	



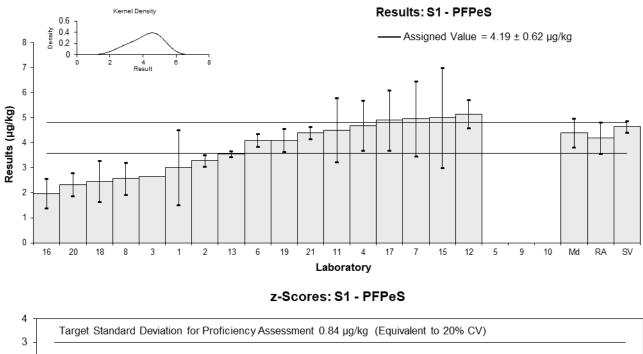
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFPeS
Unit	µg/kg

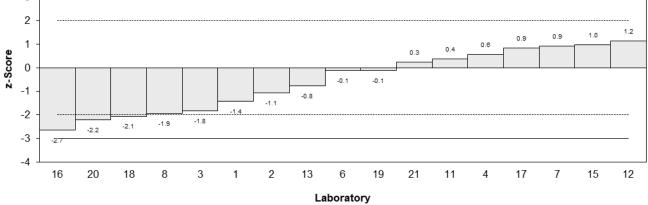
## Participant Results

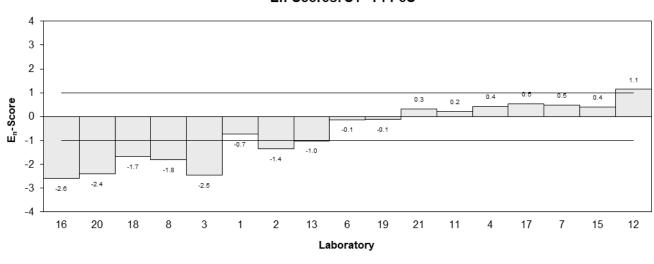
Lab. Code	Result	Uncertainty	Rec	z	En
1	3	1.5	NR	-1.42	-0.73
2	3.29	0.232	97	-1.07	-1.36
3**	2.662	NR	NR	-1.82	-2.46
4	4.68	1	NR	0.58	0.42
5	NS	NS	NS		
6	4.1	0.25	NT	-0.11	-0.13
7	4.96	1.5	79	0.92	0.47
8**	2.57	0.64	NR	-1.93	-1.82
9	NT	NT	NT		
10	NR	NR	NR		
11	4.51	1.2822	94.0	0.38	0.22
12	5.15	0.566	88.9	1.15	1.14
13	3.55	0.108	106	-0.76	-1.02
15	5	2	89	0.97	0.39
16**	1.97	0.591	138	-2.65	-2.59
17	4.9	1.2	89	0.85	0.53
18	2.46	0.83	135	-2.06	-1.67
19	4.1	0.47	113	-0.11	-0.12
20**	2.33	0.466	73	-2.22	-2.40
21	4.4	0.25	NT	0.25	0.31

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	4.19	0.62
Spike Value	4.65	0.23
Robust Average	4.19	0.62
Median	4.40	0.58
Mean	4.16	
Ν	13	
Мах	5.15	
Min	2.46	
Robust SD	0.90	
Robust CV	21%	







En-Scores: S1 - PFPeS

Figure 3

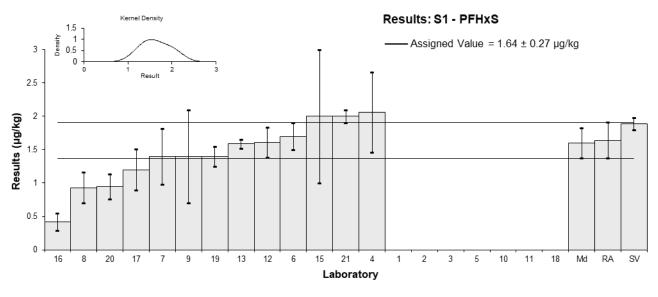
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFHxS
Unit	µg/kg

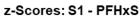
## Participant Results

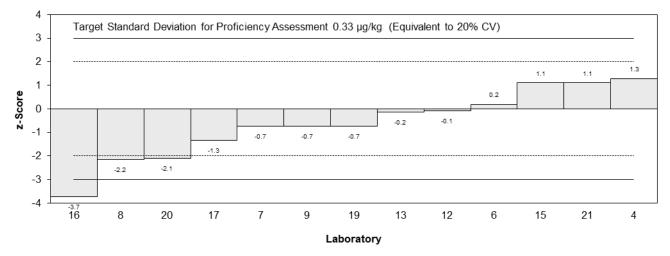
Lab. Code	Result	Uncertainty	Rec	z	En
1	< 2	1	107		
2	NR	NR	NR		
3	NT	NT	NT		
4	2.06	0.6	74	1.28	0.64
5	NS	NS	NS		
6	1.7	0.2	NT	0.18	0.18
7	1.40	0.42	83	-0.73	-0.48
8**	0.93	0.23	114	-2.16	-2.00
9	1.4	0.7	97	-0.73	-0.32
10	<1	NR	NR		
11	NT	NT	NT		
12	1.61	0.226	88.9	-0.09	-0.09
13	1.59	0.068	106	-0.15	-0.18
15	2	1	89	1.10	0.35
16**	0.418	0.1254	147	-3.73	-4.10
17	1.2	0.31	89	-1.34	-1.07
18	<1	NR	135		
19	1.4	0.15	107	-0.73	-0.78
20**	0.950	0.189	80	-2.10	-2.09
21	2.0	0.1	NT	1.10	1.25

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.64	0.27
Spike Value	1.89	0.09
Robust Average	1.64	0.27
Median	1.60	0.23
Mean	1.64	
Ν	10	
Max	2.06	
Min	1.2	
Robust SD	0.34	
Robust CV	21%	







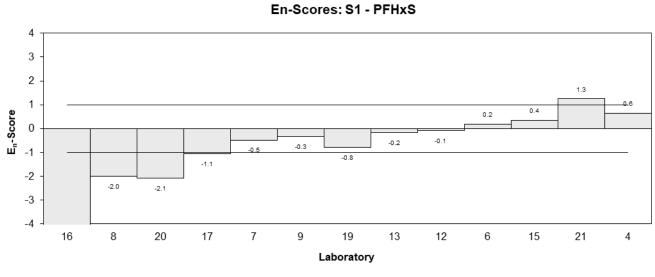


Figure 4

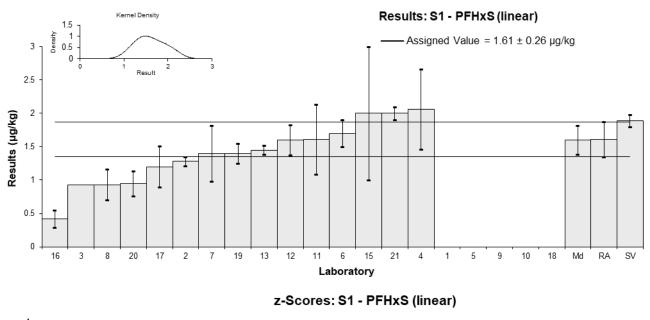
Sample No.	S1
Matrix	Prawn
Analyte	PFHxS (linear)
Unit	µg/kg

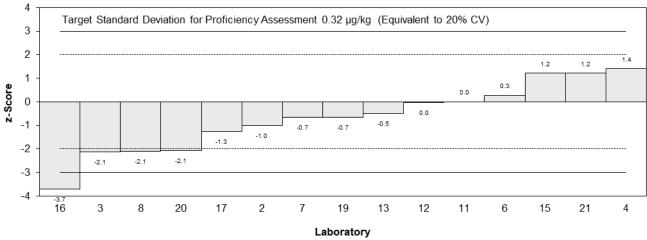
## Participant Results

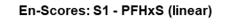
Lab. Code	Result	Uncertainty	Rec	z	En
1	< 2	1	NR		
2	1.28	0.066	97	-1.02	-1.23
3**	0.928	NR	NR	-2.12	-2.62
4	2.06	0.6	NR	1.40	0.69
5	NS	NS	NS		
6	1.7	0.2	NT	0.28	0.27
7	1.40	0.42	83	-0.65	-0.43
8**	0.93	0.23	114	-2.11	-1.96
9	NT	NT	NT		
10	NR	NR	NR		
11	1.61	0.521	94.0	0.00	0.00
12	1.6	0.226	88.9	-0.03	-0.03
13	1.45	0.067	106	-0.50	-0.60
15	2	1	89	1.21	0.38
16**	0.418	0.1254	147	-3.70	-4.13
17	1.2	0.31	89	-1.27	-1.01
18	NT	NT	NT		
19	1.4	0.15	107	-0.65	-0.70
20**	0.945	0.186	80	-2.07	-2.08
21	2.0	0.1	NT	1.21	1.40

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.61	0.26
Spike Value	1.89	0.09
Robust Average	1.61	0.26
Median	1.60	0.22
Mean	1.61	
Ν	11	
Max	2.06	
Min	1.2	
Robust SD	0.34	
Robust CV	21%	







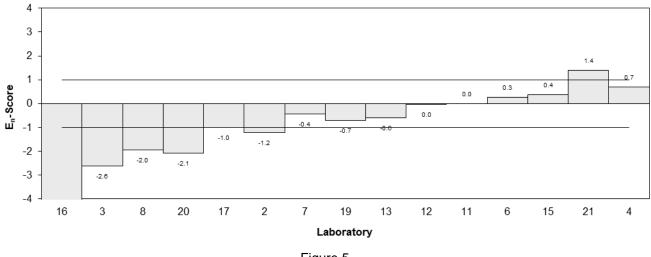


Figure 5

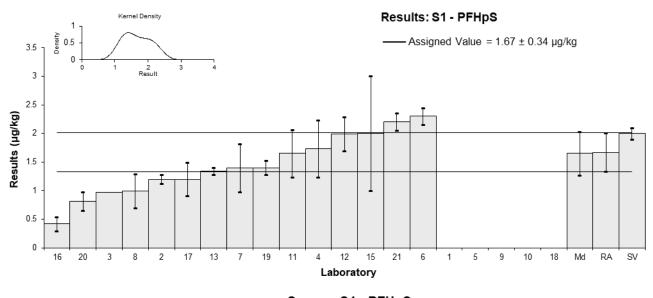
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFHpS
Unit	µg/kg

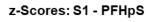
## Participant Results

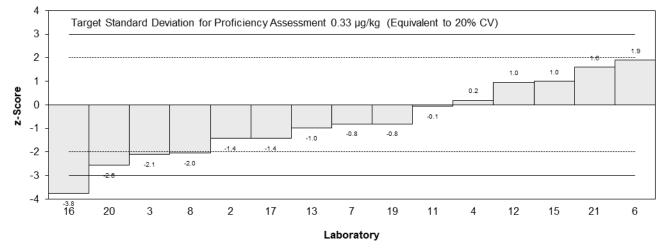
Lab. Code	Result	Uncertainty	Rec	Z	En
1	< 1	0.5	NR		
2	1.20	0.077	97	-1.41	-1.35
3**	0.967	NR	NR	-2.10	-2.07
4	1.73	0.5	NR	0.18	0.10
5	NS	NS	NS		
6	2.3	0.15	NT	1.89	1.70
7	1.40	0.42	85	-0.81	-0.50
8**	0.99	0.30	NR	-2.04	-1.50
9	NT	NT	NT		
10	<1	NR	NR		
11	1.65	0.416	94.0	-0.06	-0.04
12	1.99	0.298	83.7	0.96	0.71
13	1.34	0.067	106	-0.99	-0.95
15	2	1	89	0.99	0.31
16**	0.418	0.1254	147	-3.75	-3.45
17	1.2	0.29	89	-1.41	-1.05
18	<1	NR	151		
19	1.4	0.12	107	-0.81	-0.75
20**	0.811	0.162	80	-2.57	-2.28
21	2.2	0.15	NT	1.59	1.43

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.67	0.34
Spike Value	2.00	0.10
Robust Average	1.67	0.34
Median	1.65	0.38
Mean	1.67	
Ν	11	
Мах	2.3	
Min	1.2	
Robust SD	0.45	
Robust CV	27%	







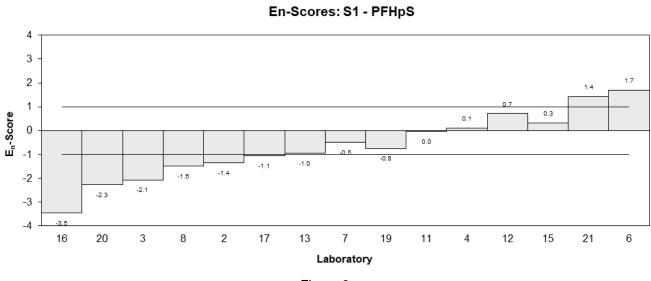


Figure 6

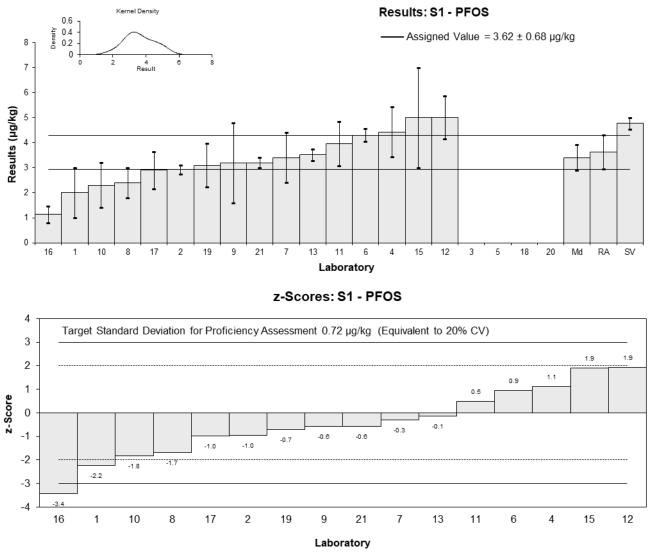
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFOS
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	2	1	111	-2.24	-1.34
2	2.93	0.183	88	-0.95	-0.98
3	NT	NT	NT		
4	4.43	1	74	1.12	0.67
5	NS	NS	NS		
6	4.3	0.25	107	0.94	0.94
7	3.4	1.0	78	-0.30	-0.18
8**	2.40	0.60	106	-1.69	-1.35
9	3.2	1.6	77	-0.58	-0.24
10**	2.3	0.9	NR	-1.82	-1.17
11	3.97	0.881	93.2	0.48	0.31
12	5.01	0.851	83.7	1.92	1.28
13	3.52	0.233	104	-0.14	-0.14
15	5	2	91	1.91	0.65
16**	1.13	0.339	136	-3.44	-3.28
17	2.9	0.74	89	-0.99	-0.72
18	<1	NR	151		
19	3.1	0.87	102	-0.72	-0.47
20	NR	NR	69		
21	3.2	0.2	107	-0.58	-0.59

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	3.62	0.68
Spike Value	4.77	0.24
Robust Average	3.62	0.68
Median	3.40	0.51
Mean	3.61	
Ν	13	
Мах	5.01	
Min	2	
Robust SD	0.98	
Robust CV	27%	



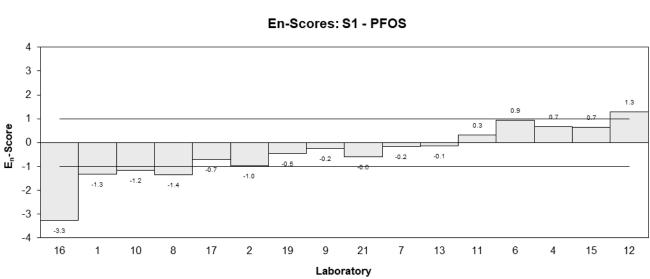




Figure 7

24

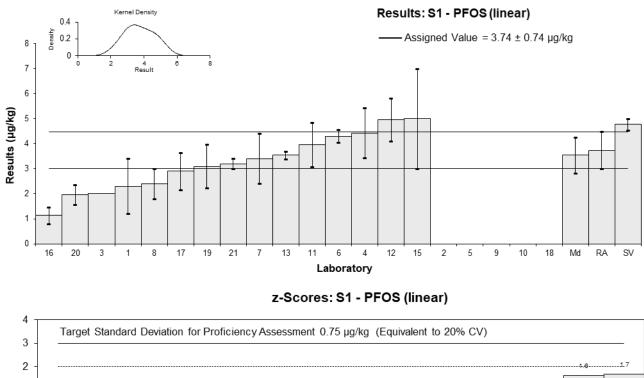
Sample No.	S1
Matrix	Prawn
Analyte	PFOS (linear)
Unit	µg/kg

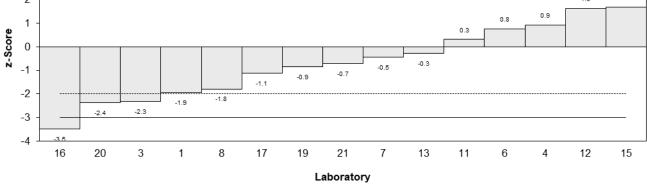
## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	2.3	1.1	NR	-1.93	-1.09
2	NR	NR	NR		
3**	2.015	NR	NR	-2.31	-2.33
4	4.43	1	NR	0.92	0.55
5	NS	NS	NS		
6	4.3	0.25	107	0.75	0.72
7	3.4	1.0	78	-0.45	-0.27
8**	2.40	0.60	106	-1.79	-1.41
9	NT	NT	NT		
10	NR	NR	NR		
11	3.97	0.881	93.2	0.31	0.20
12	4.96	0.851	83.7	1.63	1.08
13	3.54	0.162	104	-0.27	-0.26
15	5	2	91	1.68	0.59
16**	1.13	0.339	136	-3.49	-3.21
17	2.9	0.74	89	-1.12	-0.80
18	<1	NR	151		
19	3.1	0.87	102	-0.86	-0.56
20**	1.97	0.394	69	-2.37	-2.11
21	3.2	0.2	107	-0.72	-0.70

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	3.74	0.74
Spike Value	4.77	0.24
Robust Average	3.74	0.74
Median	3.54	0.72
Mean	3.74	
N	11	
Мах	5	
Min	2.3	
Robust SD	0.99	
Robust CV	26%	





En-Scores: S1 - PFOS (linear)

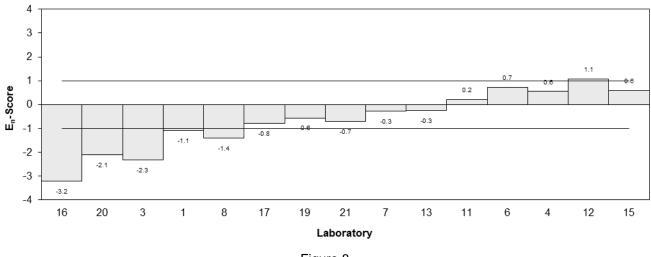


Figure 8

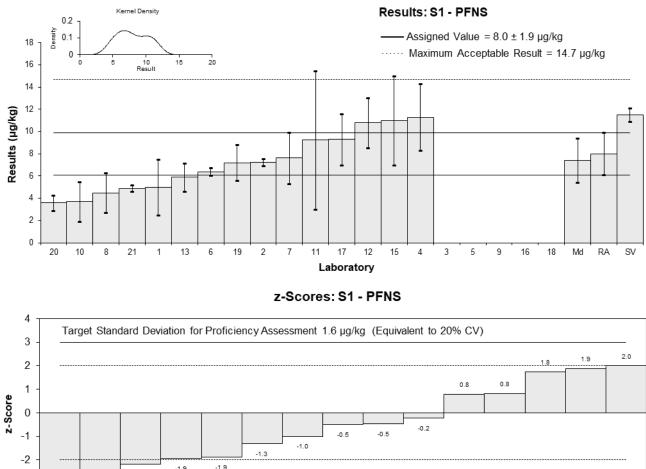
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFNS
Unit	µg/kg

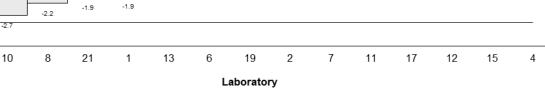
## Participant Results

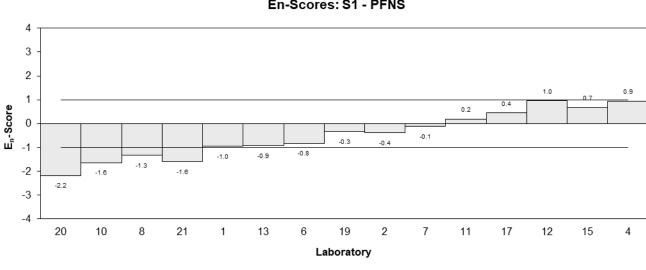
Lab. Code	Result	Uncertainty	Rec	Z	En
1	5	2.5	NR	-1.88	-0.96
2	7.25	0.314	88	-0.47	-0.39
3	NT	NT	NT		
4	11.3	3	NR	2.00▼	0.93
5	NS	NS	NS		
6	6.4	0.35	NT	-1.00	-0.83
7	7.63	2.3	82	-0.23	-0.12
8**	4.49	1.79	NR	-2.19	-1.34
9	NT	NT	NT		
10**	3.7	1.8	NR	-2.69	-1.64
11	9.24	6.23	93.2	0.78	0.19
12	10.8	2.26	83.7	1.75	0.95
13	5.89	1.25	104	-1.32	-0.93
15	11	4	91	1.88	0.68
16	NT	NT	NT		
17	9.3	2.3	89	0.81	0.44
18	NT	NT	NT		
19	7.2	1.6	102	-0.50	-0.32
20**	3.58	0.715	69	-2.76	-2.18
21	4.9	0.3	NT	-1.94	-1.61

\*\* Not included in robust average calculations, see Section 4.2; ▼ Adjusted Score, see Section 6.3

Assigned Value	8.0	1.9
Spike Value	11.5	0.6
Robust Average	8.0	1.9
Max Acceptable	14.7	
Result		
Median	7.4	2.0
Mean	8.0	
Ν	12	
Мах	11.3	
Min	4.9	
Robust SD	2.6	
Robust CV	33%	







En-Scores: S1 - PFNS

-3

-4

-2.8

20

Figure 9

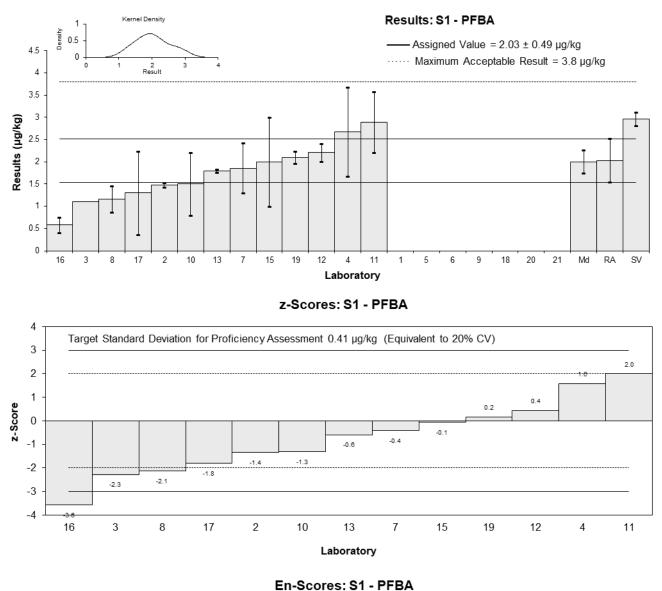
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFBA
Unit	µg/kg

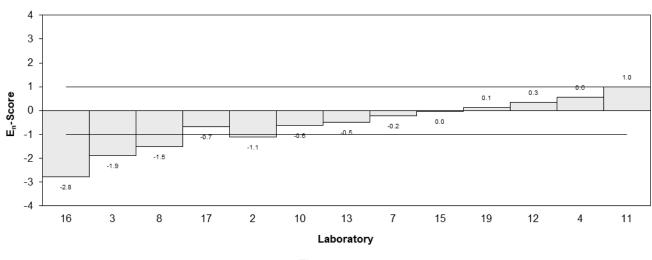
## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	< 2	1	103		
2	1.48	0.051	67	-1.35	-1.12
3**	1.104	NR	NR	-2.28	-1.89
4	2.67	1	77	1.58	0.57
5	NS	NS	NS		
6	<5	NR	105		
7	1.86	0.56	79	-0.42	-0.23
8**	1.16	0.29	88	-2.14	-1.53
9	NT	NT	NT		
10**	1.5	0.7	NR	-1.31	-0.62
11	2.89	0.680	95.7	2.00▼	1.00▼
12	2.21	0.199	85.4	0.44	0.34
13	1.79	0.035	104	-0.59	-0.49
15	2	1	64	-0.07	-0.03
16**	0.582	0.1746	102	-3.57	-2.78
17	1.3	0.94	89	-1.80	-0.69
18	<5	NR	128		
19	2.1	0.14	123	0.17	0.14
20	NT	NT	NT		
21	<5	NR	105		

\*\* Not included in robust average calculations, see Section 4.2; ▼ Adjusted Score, see Section 6.3

Assigned Value	2.03	0.49
Spike Value	2.96	0.15
Robust Average	2.03	0.49
Max Acceptable	3.8	
Result		
Median	2.00	0.26
Mean	2.03	
Ν	9	
Мах	2.89	
Min	1.3	
Robust SD	0.58	
Robust CV	29%	





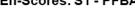


Figure 10

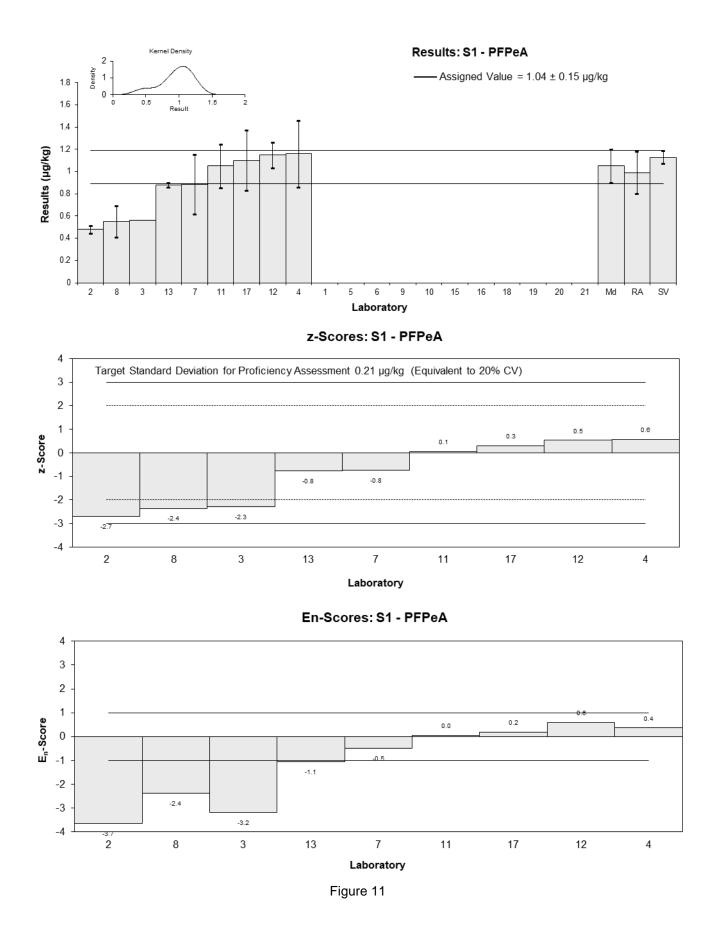
Sample No.	S1
Matrix	Prawn
Analyte	PFPeA
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	< 2	1	106		
2*	0.479	0.032	73	-2.70	-3.66
3**	0.562	NR	NR	-2.30	-3.19
4	1.16	0.3	71	0.58	0.36
5	NS	NS	NS		
6	<1	NR	NT		
7	0.885	0.27	85	-0.75	-0.50
8**	0.55	0.14	98	-2.36	-2.39
9	NT	NT	NT		
10	<1	NR	NR		
11	1.05	0.196	93.2	0.05	0.04
12	1.15	0.115	101	0.53	0.58
13	0.878	0.021	106	-0.78	-1.07
15	<2	NR	76		
16	< 0.3	NR	133		
17	1.1	0.27	89	0.29	0.19
18	<2	NR	138		
19	< 1.0	NR	118		
20	NT	NT	NT		
21	<1	NR	NT		

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.04	0.15
Spike Value	1.13	0.06
Robust Average	0.99	0.19
Median	1.05	0.15
Mean	0.96	
Ν	7	
Мах	1.16	
Min	0.479	
Robust SD	0.20	
Robust CV	20%	



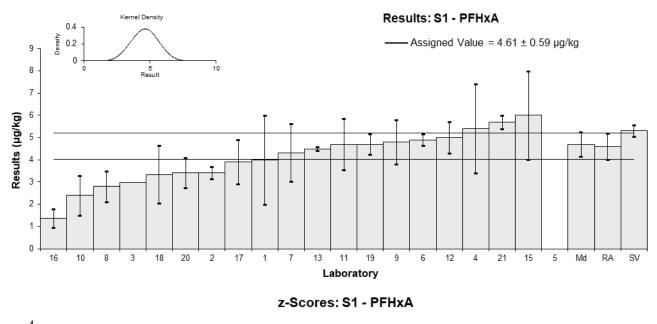
Sample No.	S1
Matrix	Prawn
Analyte	PFHxA
Unit	µg/kg

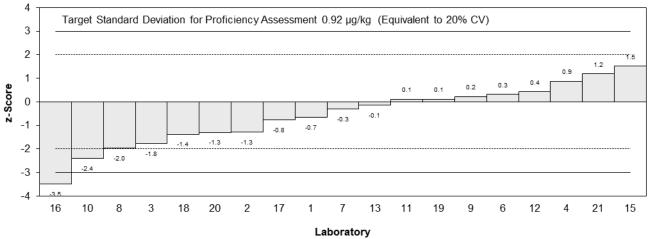
## Participant Results

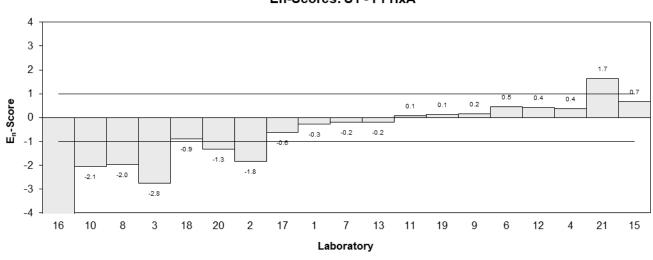
Lab. Code	Result	Uncertainty	Rec	Z	En
1	4	2	100	-0.66	-0.29
2	3.42	0.265	73	-1.29	-1.84
3**	2.979	NR	NR	-1.77	-2.76
4	5.4	2	84	0.86	0.38
5	NS	NS	NS		
6	4.9	0.25	NT	0.31	0.45
7	4.32	1.3	91	-0.31	-0.20
8**	2.80	0.70	104	-1.96	-1.98
9	4.8	1.0	95	0.21	0.16
10**	2.4	0.9	NR	-2.40	-2.05
11	4.70	1.15	96.4	0.10	0.07
12	5	0.7	83.3	0.42	0.43
13	4.49	0.092	116	-0.13	-0.20
15	6	2	85	1.51	0.67
16**	1.38	0.414	141	-3.50	-4.48
17	3.9	0.99	89	-0.77	-0.62
18	3.34	1.3	124	-1.38	-0.89
19	4.7	0.46	113	0.10	0.12
20**	3.41	0.682	68	-1.30	-1.33
21	5.7	0.3	NT	1.18	1.65

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	4.61	0.59
Spike Value	5.31	0.27
Robust Average	4.61	0.59
Median	4.70	0.54
Mean	4.62	
N	14	
Мах	6	
Min	3.34	
Robust SD	0.88	
Robust CV	19%	







En-Scores: S1 - PFHxA

Figure 12

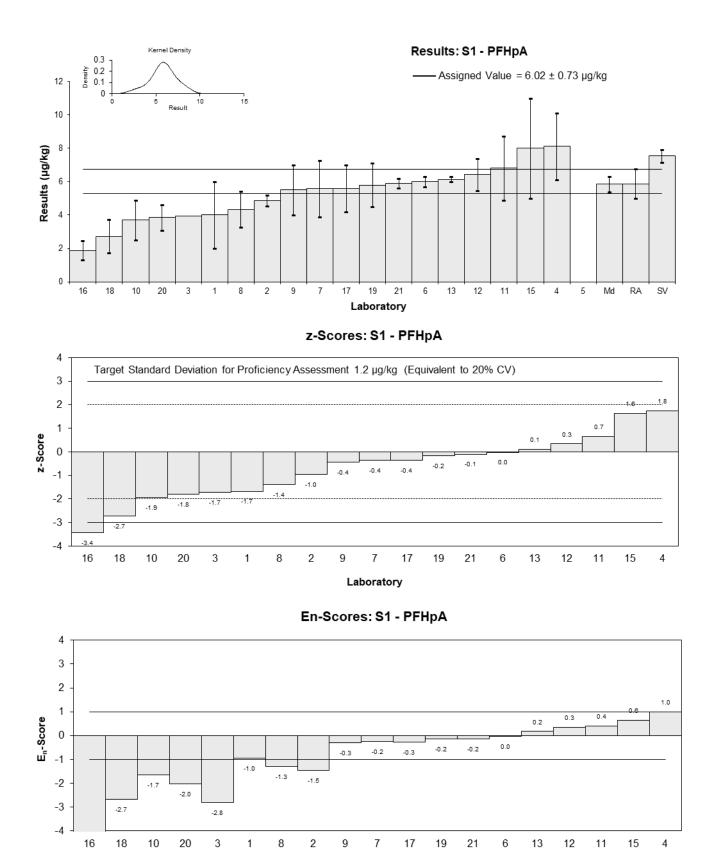
Sample No.	S1
Matrix	Prawn
Analyte	PFHpA
Unit	µg/kg

## **Participant Results**

Lab. Code	Result	Uncertainty	Rec	z	En
1	4	2	111	-1.68	-0.95
2	4.85	0.330	73	-0.97	-1.46
3**	3.958	NR	NR	-1.71	-2.82
4	8.13	2	81	1.75	0.99
5	NS	NS	NS		
6	6.0	0.3	NT	-0.02	-0.03
7	5.58	1.7	82	-0.37	-0.24
8**	4.33	1.08	103	-1.40	-1.30
9	5.5	1.5	105	-0.43	-0.31
10**	3.7	1.2	NR	-1.93	-1.65
11	6.82	1.92	94.9	0.66	0.39
12	6.43	0.965	89.3	0.34	0.34
13	6.15	0.161	104	0.11	0.17
15	8	3	87	1.64	0.64
16**	1.88	0.564	142	-3.44	-4.49
17	5.6	1.4	89	-0.35	-0.27
18*	2.72	1	122	-2.74	-2.67
19	5.8	1.3	119	-0.18	-0.15
20**	3.85	0.772	71	-1.80	-2.04
21	5.9	0.3	NT	-0.10	-0.15

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	6.02	0.73
Spike Value	7.54	0.38
Robust Average	5.88	0.90
Median	5.85	0.46
Mean	5.82	
N	14	
Мах	8.13	
Min	2.72	
Robust SD	1.3	
Robust CV	23%	



Laboratory

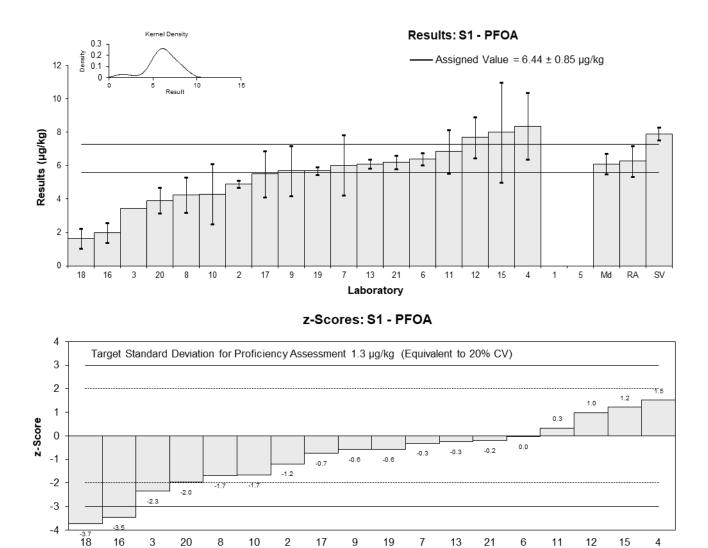
Sample No.	S1
Matrix	Prawn
Analyte	PFOA
Unit	µg/kg

## **Participant Results**

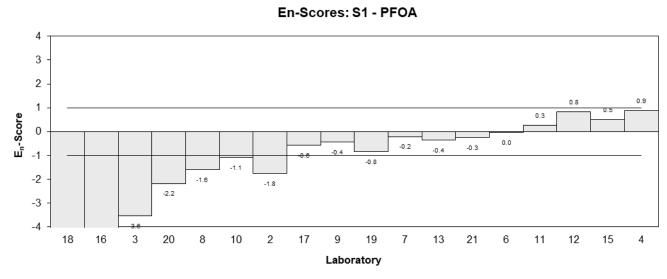
Lab. Code	Result	Uncertainty	Rec	z	En
1	< 5	2.5	103		
2	4.91	0.202	76	-1.19	-1.75
3**	3.426	NR	NR	-2.34	-3.55
4	8.38	2	73	1.51	0.89
5	NS	NS	NS		
6	6.4	0.35	108	-0.03	-0.04
7	6.03	1.8	80	-0.32	-0.21
8**	4.26	1.06	111	-1.69	-1.60
9	5.7	1.5	95	-0.57	-0.43
10**	4.3	1.8	NR	-1.66	-1.08
11	6.86	1.31	93.1	0.33	0.27
12	7.7	1.23	88.5	0.98	0.84
13	6.11	0.284	110	-0.26	-0.37
15	8	3	94	1.21	0.50
16**	1.98	0.594	152	-3.46	-4.30
17	5.5	1.4	89	-0.73	-0.57
18*	1.64	0.6	136	-3.73	-4.61
19	5.7	0.24	112	-0.57	-0.84
20**	3.92	0.784	64	-1.96	-2.18
21	6.2	0.4	108	-0.19	-0.26

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	6.44	0.85
Spike Value	7.92	0.40
Robust Average	6.28	0.92
Median	6.11	0.63
Mean	6.09	
Ν	13	
Мах	8.38	
Min	1.64	
Robust SD	1.3	
Robust CV	21%	







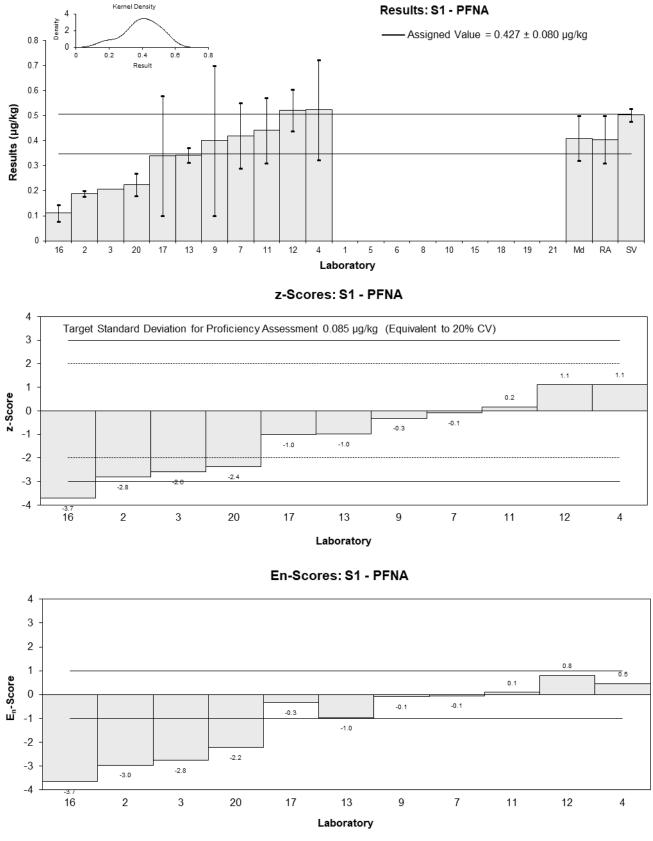
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFNA
Unit	µg/kg

## **Participant Results**

Lab. Code	Result	Uncertainty	Rec	z	En
1	< 2	1	111		
2*	0.188	0.011	76	-2.80	-2.96
3**	0.206	NR	NR	-2.59	-2.76
4	0.523	0.2	70	1.12	0.45
5	NS	NS	NS		
6	<1	NR	NT		
7	0.42	0.13	93	-0.08	-0.05
8	<0.5	0.15	95		
9	0.4	0.3	85	-0.32	-0.09
10	<1	NR	NR		
11	0.441	0.130	94.5	0.16	0.09
12	0.521	0.0834	86.7	1.10	0.81
13	0.343	0.029	102	-0.98	-0.99
15	<1	NR	93		
16**	0.111	0.0333	171	-3.70	-3.65
17	0.34	0.24	89	-1.02	-0.34
18	<1	NR	149		
19	< 1.0	NR	118		
20**	0.224	0.045	74	-2.38	-2.21
21	<1	NR	NT		

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	0.427	0.080
Spike Value	0.503	0.025
Robust Average	0.404	0.095
Median	0.410	0.090
Mean	0.397	
Ν	8	
Мах	0.523	
Min	0.188	
Robust SD	0.11	
Robust CV	27%	





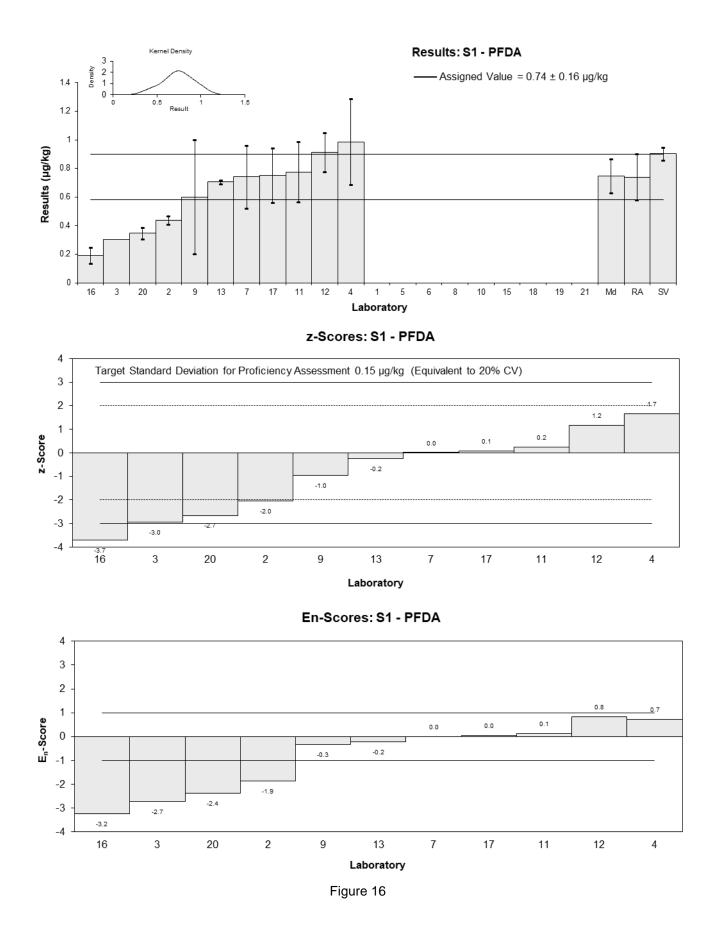
-	
Sample No.	S1
Matrix	Prawn
Analyte	PFDA
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	< 5	2.5	109		
2	0.438	0.028	76	-2.04	-1.86
3**	0.304	NR	NR	-2.95	-2.72
4	0.986	0.3	70	1.66	0.72
5	NS	NS	NS		
6	<1	NR	101		
7	0.741	0.22	95	0.01	0.00
8	<0.5	0.15	120		
9	0.6	0.4	78	-0.95	-0.32
10	NR	NR	NR		
11	0.776	0.210	94.4	0.24	0.14
12	0.913	0.137	85	1.17	0.82
13	0.705	0.014	87	-0.24	-0.22
15	<2	NR	90		
16**	0.19	0.057	154	-3.72	-3.24
17	0.75	0.19	89	0.07	0.04
18	<1	NR	168		
19	< 1.0	NR	112		
20**	0.346	0.042	75	-2.66	-2.38
21	<1	NR	101		

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	0.74	0.16
Spike Value	0.902	0.045
Robust Average	0.74	0.16
Median	0.75	0.12
Mean	0.74	
Ν	8	
Мах	0.986	
Min	0.438	
Robust SD	0.19	
Robust CV	25%	



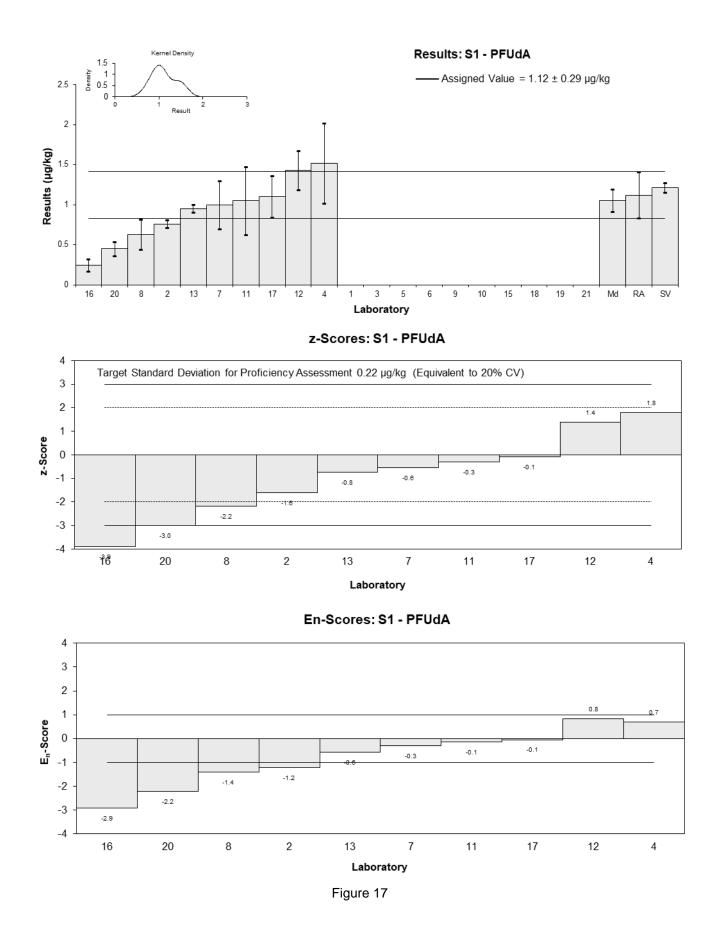
Sample No.	S1
Matrix	Prawn
Analyte	PFUdA
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	< 2	1	115		
2	0.760	0.047	66	-1.61	-1.23
3	NT	NT	NT		
4	1.52	0.5	58	1.79	0.69
5	NS	NS	NS		
6	<1	NR	NT		
7	0.994	0.30	81	-0.56	-0.30
8**	0.63	0.19	109	-2.19	-1.41
9	NT	NT	NT		
10	<1	NR	NR		
11	1.05	0.427	101.1	-0.31	-0.14
12	1.43	0.243	76	1.38	0.82
13	0.953	0.051	71	-0.75	-0.57
15	<2	NR	91		
16**	0.246	0.0738	184	-3.90	-2.92
17	1.1	0.26	89	-0.09	-0.05
18	<1	NR	159		
19	< 1.0	NR	107		
20**	0.450	0.090	76	-2.99	-2.21
21	<1	NR	NT		

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.12	0.29
Spike Value	1.21	0.06
Robust Average	1.12	0.29
Median	1.05	0.14
Mean	1.12	
N	7	
Мах	1.52	
Min	0.76	
Robust SD	0.31	
Robust CV	27%	



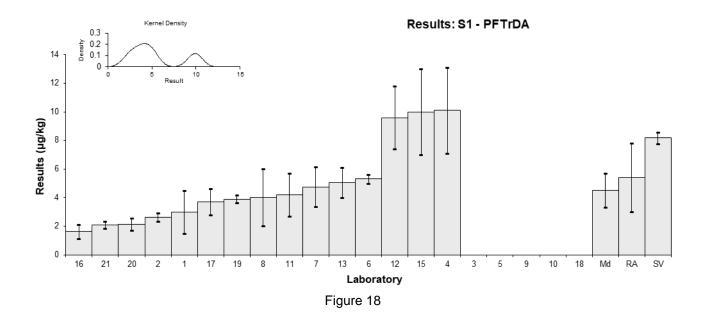
Sample No.	S1
Matrix	Prawn
Analyte	PFTrDA
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec
1	3	1.5	NR
2	2.64	0.291	56
3	NT	NT	NT
4	10.1	3	NR
5	NS	NS	NS
6	5.3	0.3	NT
7	4.75	1.4	81
8**	4.01	2.01	NR
9	NT	NT	NT
10	NR	NR	NR
11	4.20	1.49	69.1
12	9.59	2.21	62.8
13	5.03	1.05	32
15	10	3	99
16**	1.63	0.489	129
17	3.7	0.92	89
18	<2	NR	149
19	3.9	0.25	103
20**	2.141	0.428	76
21	2.1	0.25	NT

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	Not Set	
Spike Value	8.17	0.41
Robust Average	5.4	2.4
Median	4.5	1.2
Mean	5.4	
Ν	12	
Мах	10.1	
Min	2.1	
Robust SD	3.3	
Robust CV	61%	



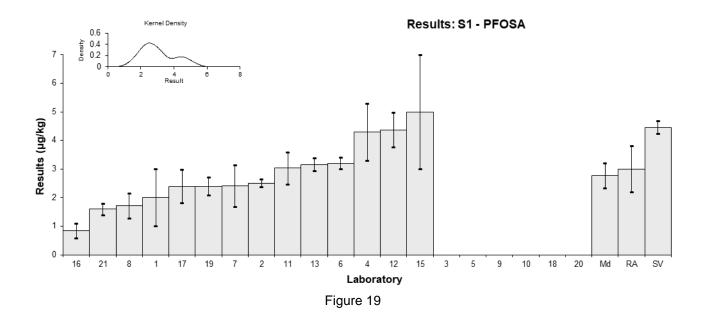
Sample No.	S1
Matrix	Prawn
Analyte	PFOSA
Unit	µg/kg

### Participant Results

Lab. Code	Result	Uncertainty	Rec
1	2	1	111
2	2.51	0.135	92
3	NT	NT	NT
4	4.29	1	76
5	NS	NS	NS
6	3.2	0.2	NT
7	2.41	0.72	83
8**	1.72	0.43	89
9	NT	NT	NT
10	NR	NR	NR
11	3.03	0.563	19.5
12	4.37	0.612	112
13	3.16	0.227	76
15	5	2	92
16**	0.844	0.2532	105
17	2.4	0.59	89
18	<5	NR	114
19	2.4	0.32	105
20	NT	NT	NT
21	1.6	0.2	NT

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	Not Set	
Spike Value	4.46	0.22
Robust Average	3.00	0.81
Median	2.77	0.44
Mean	3.03	
N	12	
Мах	5	
Min	1.6	
Robust SD	1.1	
Robust CV	37%	



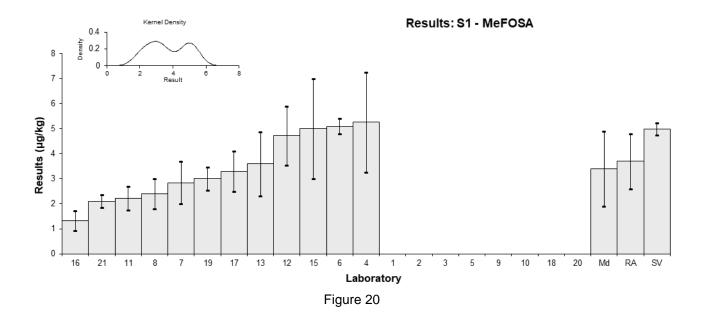
-	
Sample No.	S1
Matrix	Prawn
Analyte	MeFOSA
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec
1	< 5	2.5	110
2	NT	NT	NT
3	NT	NT	NT
4	5.26	2	6
5	NS	NS	NS
6	5.1	0.3	NT
7	2.84	0.85	75
8**	2.40	0.60	115
9	NT	NT	NT
10	NR	NR	NR
11	2.21	0.478	19.5
12	4.72	1.18	56.9
13	3.59	1.28	19
15	5	2	88
16**	1.32	0.396	144
17	3.3	0.81	89
18	<5	NR	194
19	3.0	0.47	115
20**	NT	NT	NT
21	2.1	0.25	NT

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	Not Set	
Spike Value	4.99	0.25
Robust Average	3.7	1.1
Median	3.4	1.5
Mean	3.71	
N	10	
Мах	5.26	
Min	2.1	
Robust SD	1.4	
Robust CV	37%	



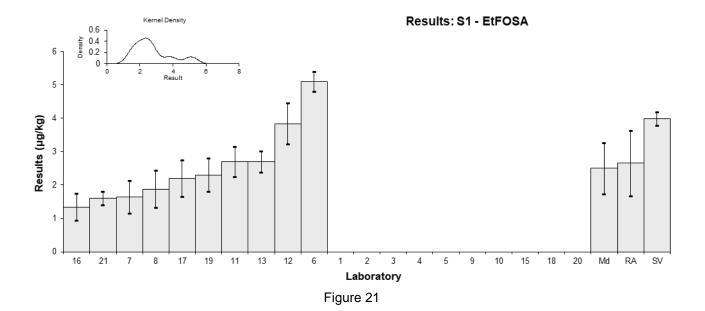
Sample No.	S1
Matrix	Prawn
Analyte	EtFOSA
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec
1	< 5	2.5	115
2	NT	NT	NT
3	NT	NT	NT
4	<5	NR	2
5	NS	NS	NS
6	5.1	0.3	NT
7	1.64	0.49	73
8**	1.88	0.56	126
9	NT	NT	NT
10	NR	NR	NR
11	2.70	0.453	19.5
12	3.84	0.614	58.8
13	2.7	0.313	11
15	<5	NR	90
16**	1.34	0.402	98
17	2.2	0.54	89
18	<2	NR	100
19	2.3	0.50	112
20	NT	NT	NT
21	1.6	0.2	NT

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	Not Set	
Spike Value	3.99	0.20
Robust Average	2.66	0.98
Median	2.50	0.76
Mean	2.76	
N	8	
Мах	5.1	
Min	1.6	
Robust SD	1.1	
Robust CV	41%	



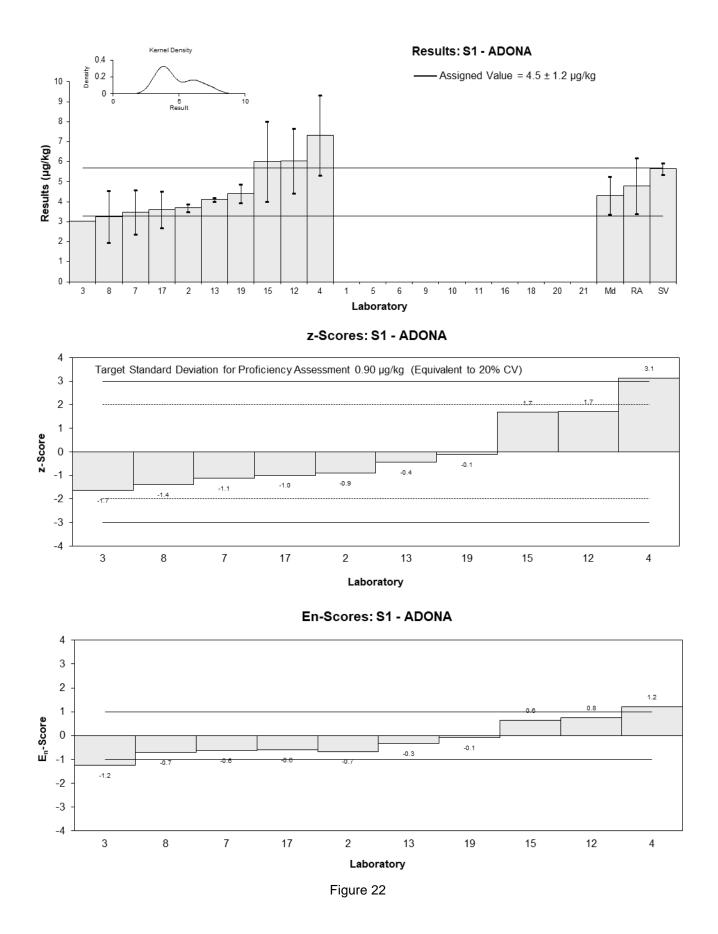
Sample No.	S1
Matrix	Prawn
Analyte	ADONA
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	NT	NT	NT		
2	3.69	0.183	76	-0.90	-0.67
3**	3.013	NR	NR	-1.65	-1.24
4*	7.31	2	NR	3.12	1.20
5	NS	NS	NS		
6	NT	NT	NT		
7	3.48	1.1	87	-1.13	-0.63
8**	3.25	1.30	NR	-1.39	-0.71
9	NT	NT	NT		
10	NR	NR	NR		
11	NT	NT	NT		
12	6.04	1.63	78.7	1.71	0.76
13	4.11	0.099	110	-0.43	-0.32
15	6	2	91	1.67	0.64
16	NR	NR	NR		
17	3.6	0.92	89	-1.00	-0.60
18	NT	NT	NT		
19	4.4	0.46	119	-0.11	-0.08
20	NT	NT	NT		
21	NT	NT	NT		

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	4.5	1.2
Spike Value	5.64	0.28
Robust Average	4.8	1.4
Median	4.26	0.94
Mean	4.8	
N	8	
Мах	7.31	
Min	3.48	
Robust SD	1.6	
Robust CV	33%	



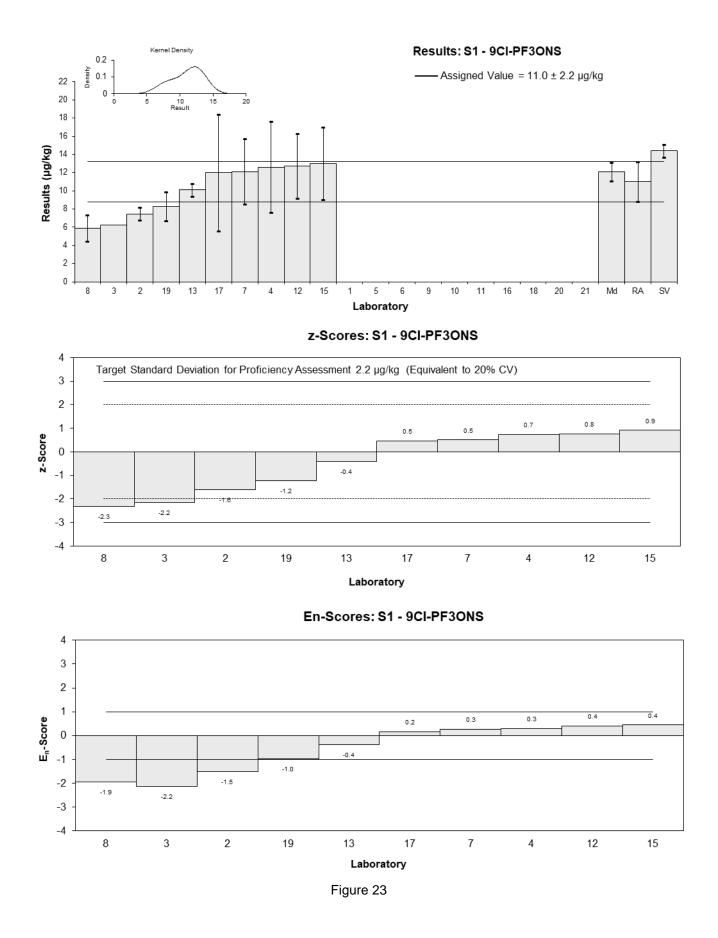
Sample No.	S1
Matrix	Prawn
Analyte	9CI-PF3ONS
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	NT	NT	NT		
2	7.47	0.673	97	-1.60	-1.53
3**	6.268	NR	NR	-2.15	-2.15
4	12.6	5	NR	0.73	0.29
5	NS	NS	NS		
6	NT	NT	NT		
7	12.1	3.6	82	0.50	0.26
8**	5.88	1.47	NR	-2.33	-1.94
9	NT	NT	NT		
10	NR	NR	NR		
11	NT	NT	NT		
12	12.7	3.55	78.7	0.77	0.41
13	10.1	0.692	104	-0.41	-0.39
15	13	4	91	0.91	0.44
16	NR	NR	NR		
17	12	6.4	89	0.45	0.15
18	NT	NT	NT		
19	8.3	1.6	102	-1.23	-0.99
20	NT	NT	NT		
21	NT	NT	NT		

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	11.0	2.2
Spike Value	14.4	0.7
Robust Average	11.0	2.2
Median	12.1	1.0
Mean	11.0	
Ν	8	
Мах	13	
Min	7.47	
Robust SD	2.4	
Robust CV	22%	



Sample No.	S1
Matrix	Prawn
Analyte	11CI-PF3OUdS
Unit	µg/kg

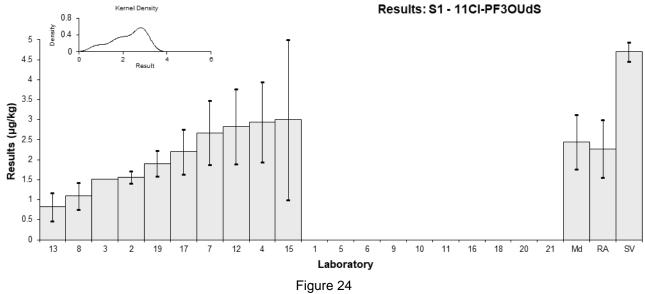
## Participant Results

Lab. Code	Result	Uncertainty	Rec
1	NT	NT	NT
2	1.56	0.150	97
3**	1.522	NR	NR
4	2.94	1	NR
5	NS	NS	NS
6	NT	NT	NT
7	2.67	0.80	82
8**	1.09	0.33	NR
9	NT	NT	NT
10	NR	NR	NR
11	NT	NT	NT
12	2.83	0.932	78.7
13	0.82	0.356	104
15	3	2	91
16	NR	NR	NR
17	2.2	0.56	89
18	NT	NT	NT
19	1.9	0.32	103
20	NT	NT	NT
21	NT	NT	NT

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	Not Set	
Spike Value	4.70	0.24
Robust Average	2.27	0.72
Median	2.44	0.68
Mean	2.24	
Ν	8	
Мах	3	
Min	0.82	
Robust SD	0.81	
Robust CV	36%	

Results: S1 - 11CI-PF3OUdS



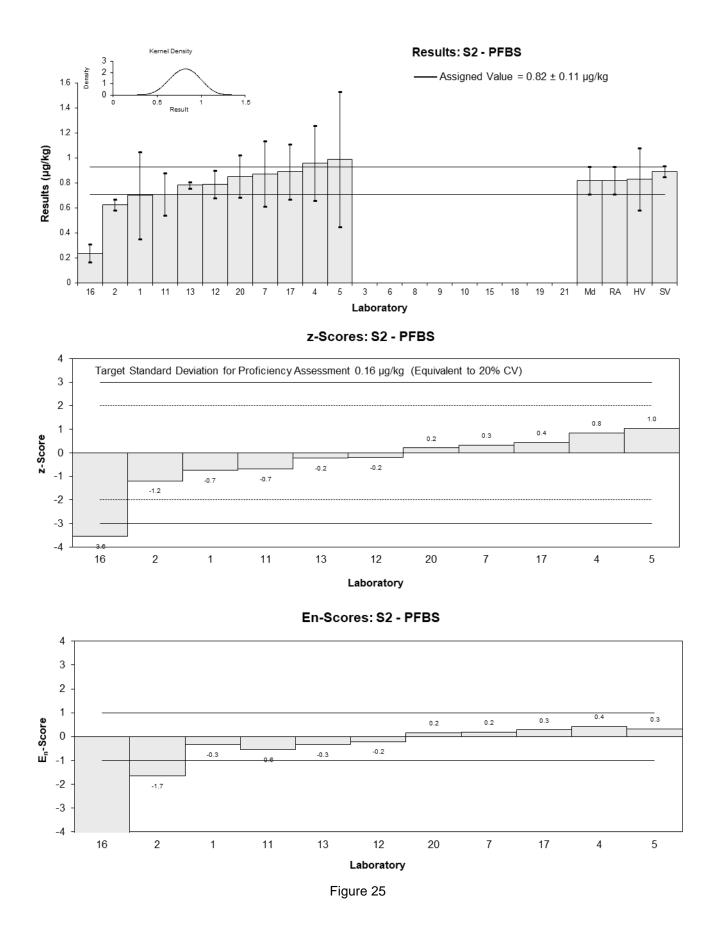
Sample No.	S2
Matrix	Carrot
Analyte	PFBS
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	0.7	0.35	95	-0.73	-0.33
2	0.625	0.043	108	-1.19	-1.65
3	NS	NS	NS		
4	0.957	0.3	77	0.84	0.43
5	0.99	0.54	NR	1.04	0.31
6	<1	NR	NT		
7	0.873	0.26	96	0.32	0.19
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	0.709	0.170	86.4	-0.68	-0.55
12	0.787	0.11	91.2	-0.20	-0.21
13	0.782	0.025	145	-0.23	-0.34
15	<1	NR	87		
16**	0.238	0.0714	151	-3.55	-4.44
17	0.89	0.22	94	0.43	0.28
18	<1	NR	128		
19	NR	NR	103		
20	0.853	0.171	61	0.20	0.16
21	<1	NR	NT		

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	0.82	0.11
Spike Value	0.891	0.045
Homogeneity Value	0.83	0.25
Robust Average	0.82	0.11
Median	0.82	0.11
Mean	0.817	
Ν	10	
Мах	0.99	
Min	0.625	
Robust SD	0.13	
Robust CV	16%	



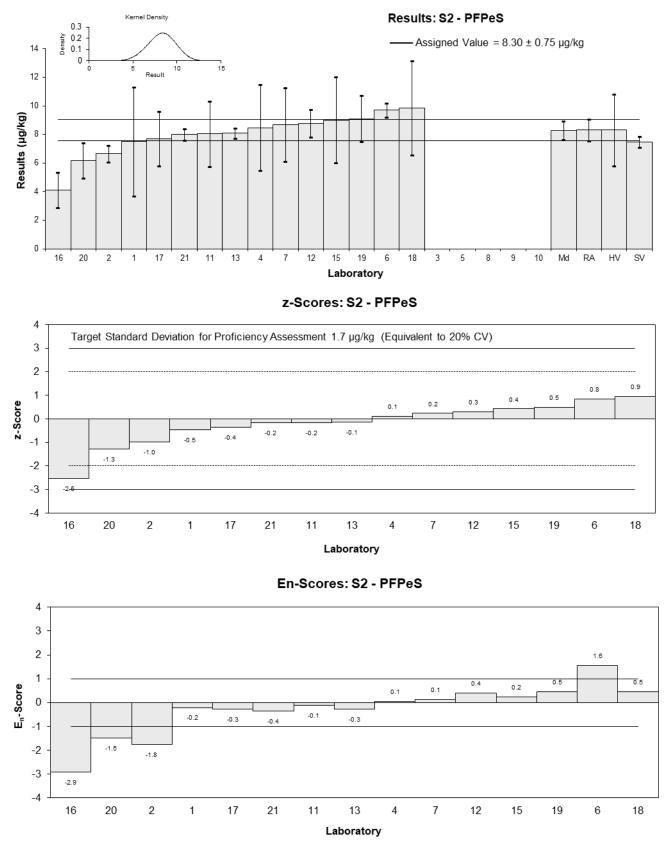
Sample No.	S2
Matrix	Carrot
Analyte	PFPeS
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	7.5	3.8	NR	-0.48	-0.21
2	6.65	0.572	108	-0.99	-1.75
3	NS	NS	NS		
4	8.47	3	NR	0.10	0.05
5	NT	NT	NT		
6	9.7	0.5	NT	0.84	1.55
7	8.68	2.6	85	0.23	0.14
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	8.04	2.28	86.4	-0.16	-0.11
12	8.78	0.966	87.6	0.29	0.39
13	8.08	0.347	117	-0.13	-0.27
15	9	3	88	0.42	0.23
16**	4.09	1.227	151	-2.54	-2.93
17	7.7	1.9	94	-0.36	-0.29
18	9.86	3.3	128	0.94	0.46
19	9.1	1.6	93	0.48	0.45
20	6.17	1.23	142	-1.28	-1.48
21	8.0	0.4	NT	-0.18	-0.35

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	8.30	0.75
Spike Value	7.47	0.37
Homogeneity Value	8.3	2.5
Robust Average	8.30	0.75
Median	8.28	0.64
Mean	8.27	
N	14	
Мах	9.86	
Min	6.17	
Robust SD	1.1	
Robust CV	13%	





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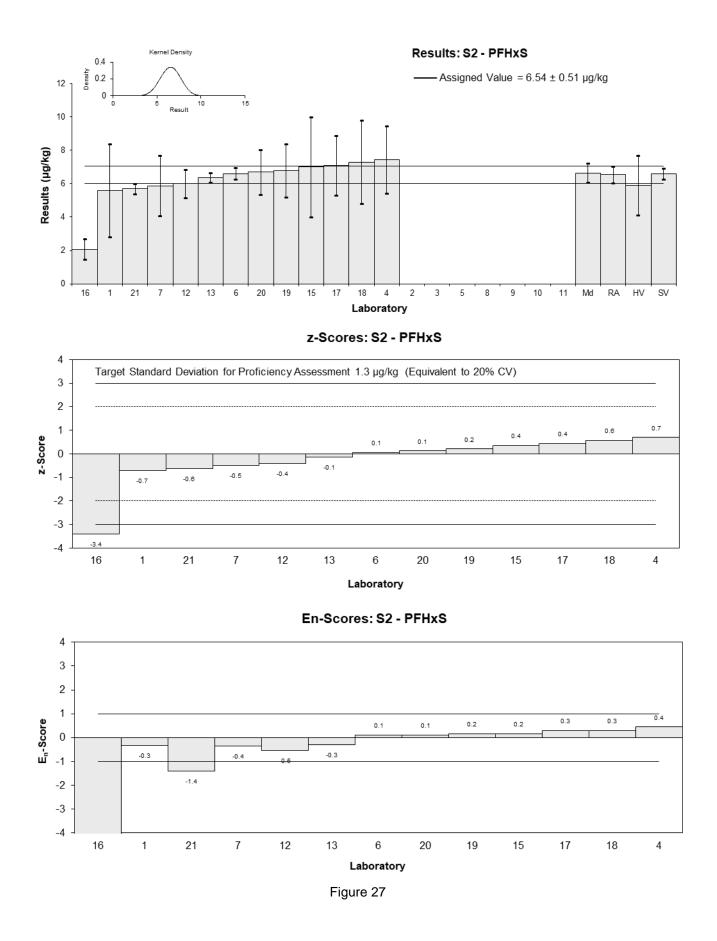
Sample No.	S2
Matrix	Carrot
Analyte	PFHxS
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	5.6	2.8	98	-0.72	-0.33
2	NR	NR	NR		
3	NS	NS	NS		
4	7.44	2	81	0.69	0.44
5	NT	NT	NT		
6	6.6	0.35	NT	0.05	0.10
7	5.88	1.8	92	-0.50	-0.35
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	NT	NT	NT		
12	6.01	0.842	87.6	-0.41	-0.54
13	6.36	0.289	117	-0.14	-0.31
15	7	3	88	0.35	0.15
16**	2.07	0.621	134	-3.42	-5.56
17	7.1	1.8	94	0.43	0.30
18	7.3	2.5	136	0.58	0.30
19	6.8	1.6	106	0.20	0.15
20	6.70	1.34	142	0.12	0.11
21	5.7	0.3	NT	-0.64	-1.42

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	6.54	0.51
Spike Value	6.61	0.33
Homogeneity Value	5.9	1.8
Robust Average	6.54	0.51
Median	6.65	0.58
Mean	6.54	
N	12	
Мах	7.44	
Min	5.6	
Robust SD	0.71	
Robust CV	11%	



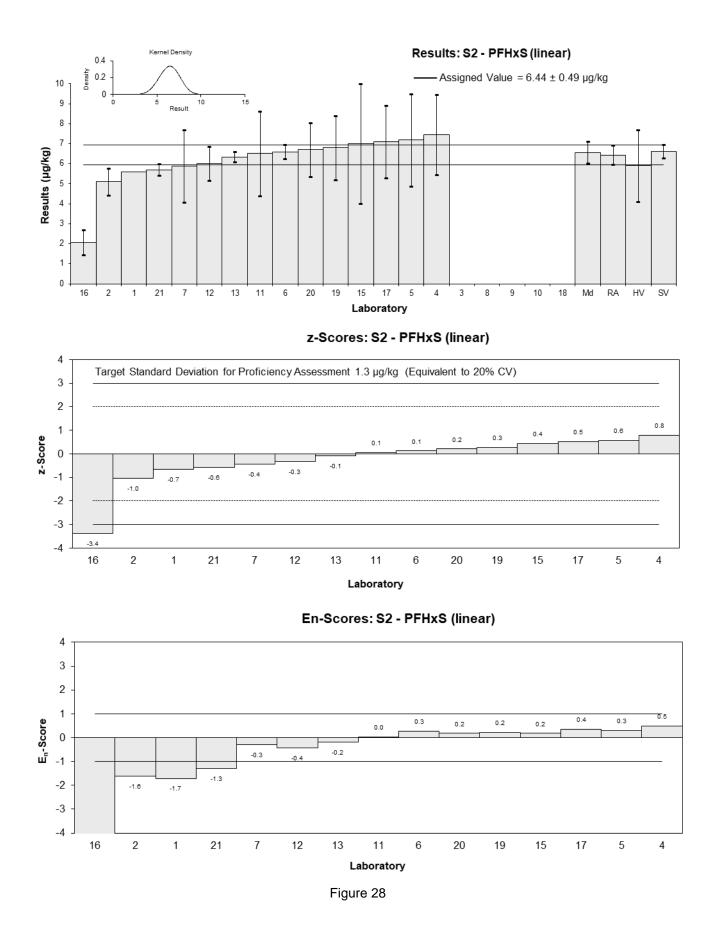
Sample No.	S2
Matrix	Carrot
Analyte	PFHxS (linear)
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	5.6	NR	NR	-0.65	-1.71
2	5.10	0.665	99	-1.04	-1.62
3	NS	NS	NS		
4	7.44	2	NR	0.78	0.49
5	7.18	2.3	NR	0.57	0.31
6	6.6	0.35	NT	0.12	0.27
7	5.88	1.8	92	-0.43	-0.30
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	6.51	2.11	86.4	0.05	0.03
12	6.01	0.842	87.6	-0.33	-0.44
13	6.34	0.248	117	-0.08	-0.18
15	7	3	88	0.43	0.18
16**	2.07	0.621	134	-3.39	-5.52
17	7.1	1.8	94	0.51	0.35
18	NT	NT	NT		
19	6.8	1.6	106	0.28	0.22
20	6.70	1.34	142	0.20	0.18
21	5.7	0.3	NT	-0.57	-1.29

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	6.44	0.49
Spike Value	6.61	0.33
Homogeneity Value	5.9	1.8
Robust Average	6.44	0.49
Median	6.56	0.54
Mean	6.43	
Ν	14	
Мах	7.44	
Min	5.1	
Robust SD	0.74	
Robust CV	11%	



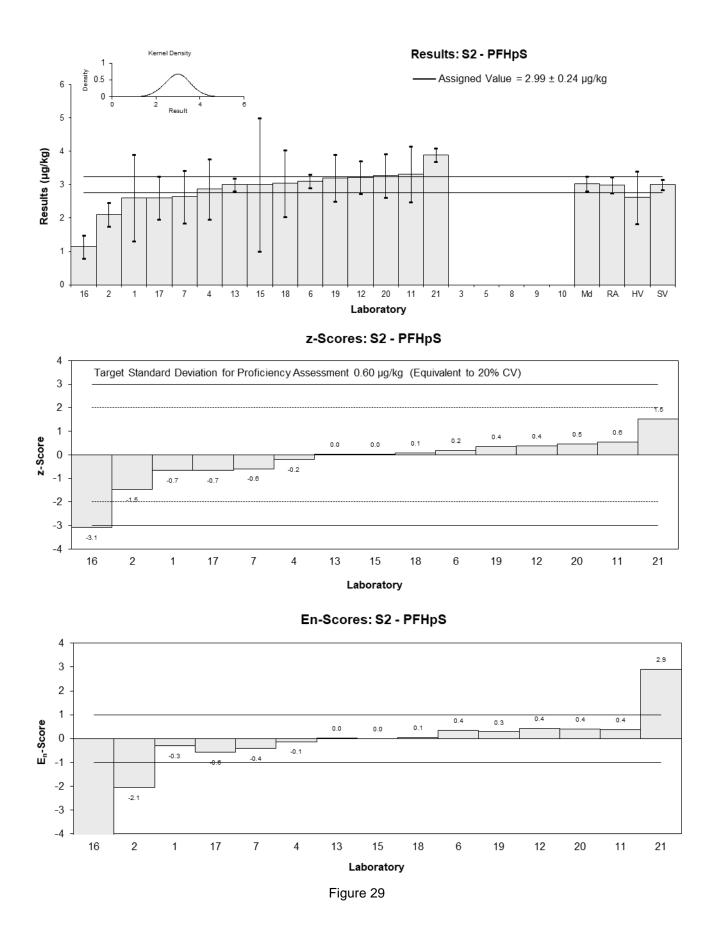
-	
Sample No.	S2
Matrix	Carrot
Analyte	PFHpS
Unit	µg/kg

## Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	2.6	1.3	NR	-0.65	-0.30
2	2.11	0.357	99	-1.47	-2.05
3	NS	NS	NS		
4	2.87	0.9	NR	-0.20	-0.13
5	NT	NT	NT		
6	3.1	0.2	NT	0.18	0.35
7	2.64	0.79	82	-0.59	-0.42
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	3.32	0.840	86.4	0.55	0.38
12	3.22	0.483	92.1	0.38	0.43
13	3	0.186	117	0.02	0.03
15	3	2	88	0.02	0.00
16**	1.14	0.342	162	-3.09	-4.43
17	2.6	0.64	94	-0.65	-0.57
18	3.04	1	137	0.08	0.05
19	3.2	0.70	106	0.35	0.28
20	3.27	0.654	142	0.47	0.40
21	3.9	0.2	NT	1.52	2.91

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	2.99	0.24
Spike Value	3.00	0.15
Homogeneity Value	2.62	0.79
Robust Average	2.99	0.24
Median	3.02	0.22
Mean	2.99	
Ν	14	
Мах	3.9	
Min	2.11	
Robust SD	0.36	
Robust CV	12%	



#### AQA 22-14 PFAS in Biota and Food

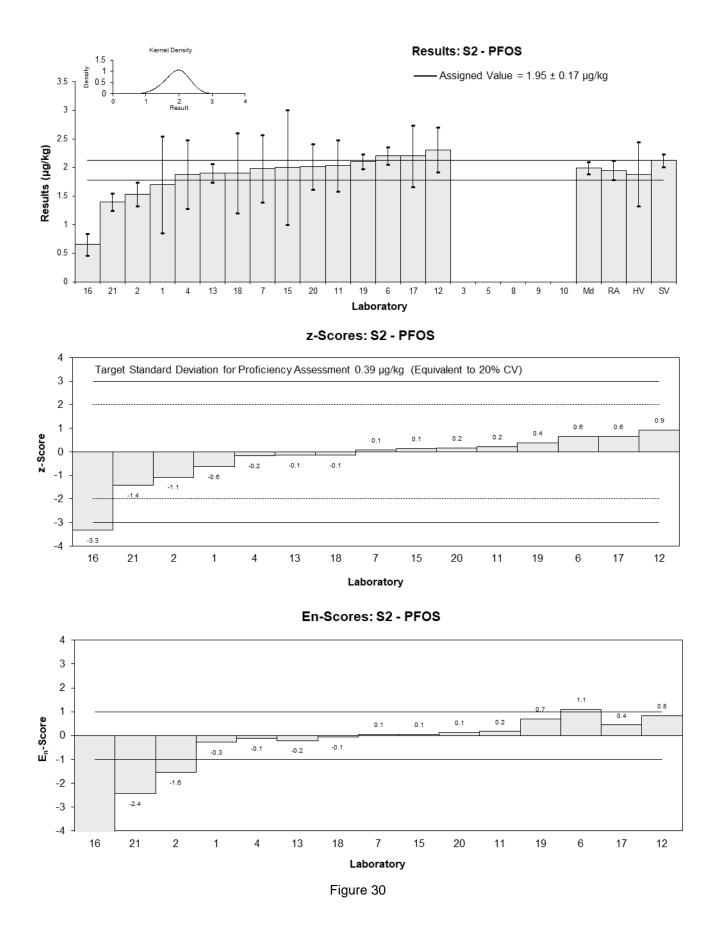
Sample No.	S2
Matrix	Carrot
Analyte	PFOS
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	1.7	0.85	100	-0.64	-0.29
2	1.53	0.211	88	-1.08	-1.55
3	NS	NS	NS		
4	1.88	0.6	89	-0.18	-0.11
5	NT	NT	NT		
6	2.2	0.15	92	0.64	1.10
7	1.98	0.59	91	0.08	0.05
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	2.03	0.450	88.7	0.21	0.17
12	2.31	0.392	92.1	0.92	0.84
13	1.9	0.159	120	-0.13	-0.21
15	2	1	89	0.13	0.05
16**	0.651	0.1953	157	-3.33	-5.02
17	2.2	0.54	94	0.64	0.44
18	1.9	0.7	137	-0.13	-0.07
19	2.1	0.13	104	0.38	0.70
20	2.01	0.40	83	0.15	0.14
21	1.4	0.15	92	-1.41	-2.43

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.95	0.17
Spike Value	2.12	0.11
Homogeneity Value	1.88	0.56
Robust Average	1.95	0.17
Median	1.99	0.11
Mean	1.94	
Ν	14	
Мах	2.31	
Min	1.4	
Robust SD	0.25	
Robust CV	13%	



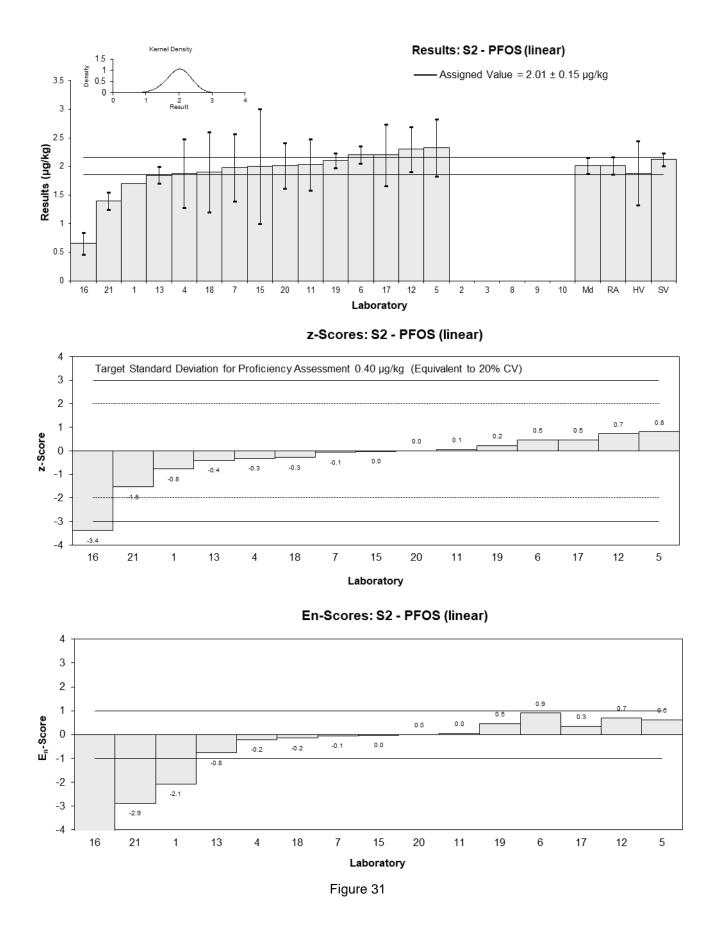
Sample No.	S2
Matrix	Carrot
Analyte	PFOS (linear)
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	1.7	NR	NR	-0.77	-2.07
2	NR	NR	NR		
3	NS	NS	NS		
4	1.88	0.6	NR	-0.32	-0.21
5	2.33	0.50	NR	0.80	0.61
6	2.2	0.15	92	0.47	0.90
7	1.98	0.59	91	-0.07	-0.05
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	2.03	0.450	88.7	0.05	0.04
12	2.3	0.392	92.1	0.72	0.69
13	1.85	0.146	120	-0.40	-0.76
15	2	1	89	-0.02	-0.01
16**	0.651	0.1953	157	-3.38	-5.52
17	2.2	0.54	94	0.47	0.34
18	1.9	0.7	137	-0.27	-0.15
19	2.1	0.13	104	0.22	0.45
20	2.01	0.40	83	0.00	0.00
21	1.4	0.15	92	-1.52	-2.88

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	2.01	0.15
Spike Value	2.12	0.11
Homogeneity Value	1.88	0.56
Robust Average	2.01	0.15
Median	2.01	0.14
Mean	1.99	
N	14	
Мах	2.33	
Min	1.4	
Robust SD	0.23	
Robust CV	11%	

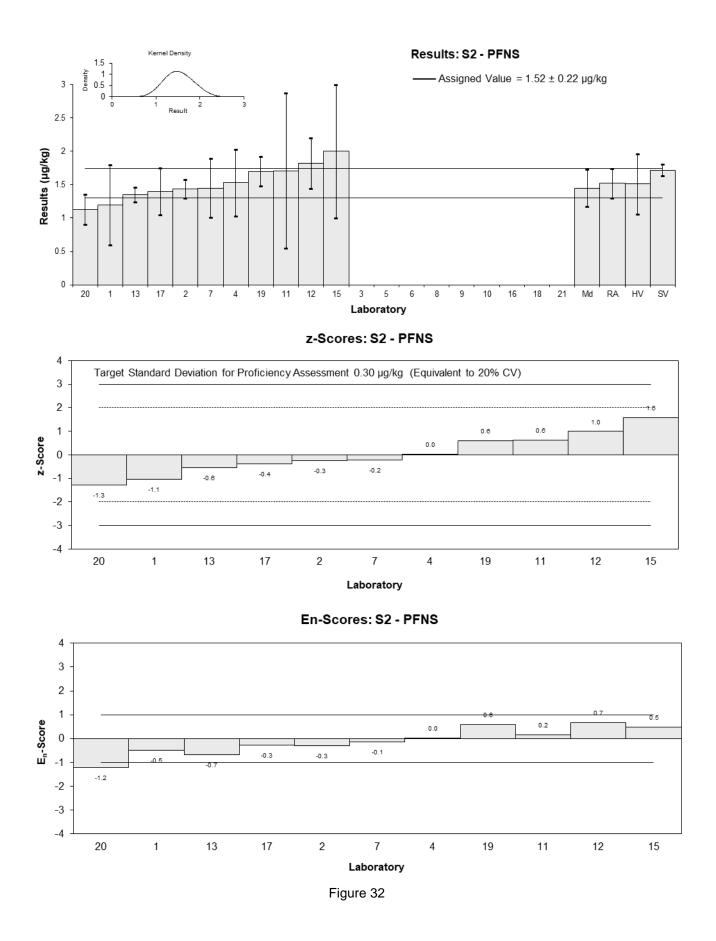


Sample No.	S2
Matrix	Carrot
Analyte	PFNS
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	1.2	0.6	NR	-1.05	-0.50
2	1.44	0.140	88	-0.26	-0.31
3	NS	NS	NS		
4	1.53	0.5	NR	0.03	0.02
5	NT	NT	NT		
6	<1	NR	NT		
7	1.45	0.44	87	-0.23	-0.14
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	1.71	1.16	88.7	0.62	0.16
12	1.82	0.381	92.1	0.99	0.68
13	1.35	0.109	120	-0.56	-0.69
15	2	1	89	1.58	0.47
16	NT	NT	NT		
17	1.4	0.35	94	-0.39	-0.29
18	NT	NT	NT		
19	1.7	0.22	104	0.59	0.58
20	1.13	0.23	83	-1.28	-1.23
21	<1	NR	NT		

Assigned Value	1.52	0.22
Spike Value	1.72	0.09
Homogeneity Value	1.51	0.45
Robust Average	1.52	0.22
Median	1.45	0.28
Mean	1.52	
Ν	11	
Max	2	
Min	1.13	
Robust SD	0.29	
Robust CV	19%	



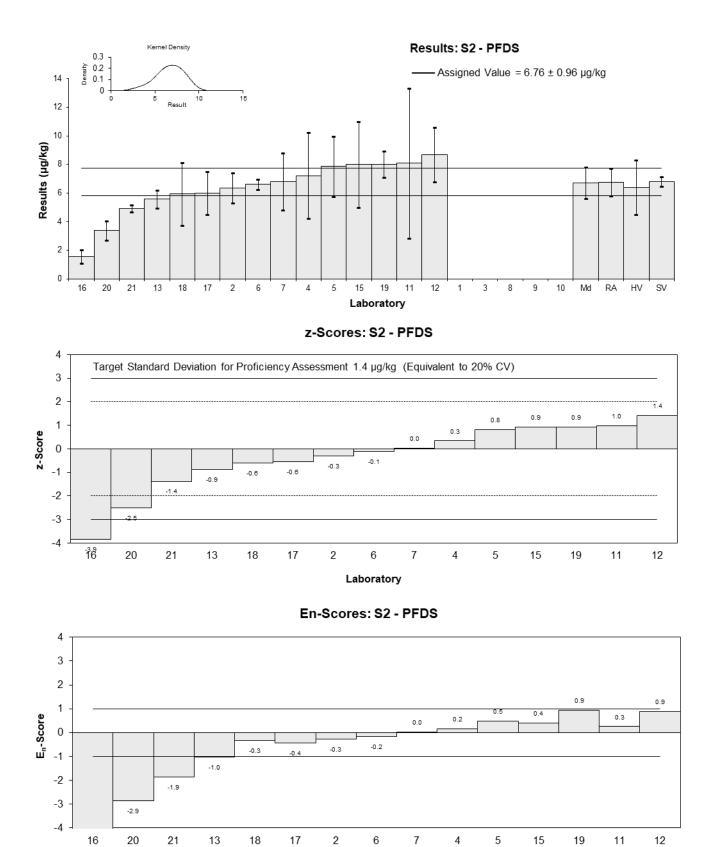
-	
Sample No.	S2
Matrix	Carrot
Analyte	PFDS
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	NT	NT	NT		
2	6.35	1.04	88	-0.30	-0.29
3	NS	NS	NS		
4	7.22	3	89	0.34	0.15
5	7.86	2.1	NR	0.81	0.48
6	6.6	0.35	NT	-0.12	-0.16
7	6.80	2.0	86	0.03	0.02
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	8.08	5.24	88.7	0.98	0.25
12	8.67	1.91	92.1	1.41	0.89
13	5.57	0.631	120	-0.88	-1.04
15	8	3	89	0.92	0.39
16**	1.55	0.465	181	-3.85	-4.88
17	6.0	1.5	94	-0.56	-0.43
18	5.94	2.2	128	-0.61	-0.34
19	8.0	0.93	104	0.92	0.93
20	3.38	0.68	83	-2.50	-2.87
21	4.9	0.25	NT	-1.38	-1.87

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	6.76	0.96
Spike Value	6.80	0.34
Homogeneity Value	6.4	1.9
Robust Average	6.76	0.96
Median	6.7	1.1
Mean	6.67	
N	14	
Мах	8.67	
Min	3.38	
Robust SD	1.4	
Robust CV	21%	



Laboratory

Figure 33

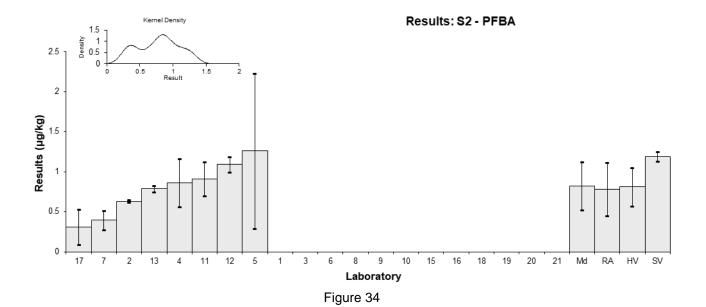
76

Sample No.	S2
Matrix	Carrot
Analyte	PFBA
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec
1	< 1	0.5	92
2	0.630	0.016	91
3	NS	NS	NS
4	0.861	0.3	85
5	1.26	0.97	NR
6	<5	NR	81
7	0.395	0.12	81
8	NS	NS	NS
9	NT	NT	NT
10	NS	NS	NS
11	0.908	0.213	74.3
12	1.09	0.0983	86.9
13	0.788	0.04	108
15	<2	NR	86
16	< 0.3	NR	97.3
17	0.31	0.22	94
18	<5	NR	91
19	NR	NR	112
20	NT	NT	NT
21	<5	NR	81

Assigned Value	Not Set	
Spike Value	1.19	0.06
Homogeneity Value	0.81	0.24
Robust Average	0.78	0.33
Median	0.82	0.30
Mean	0.78	
Ν	8	
Мах	1.26	
Min	0.31	
Robust SD	0.37	
Robust CV	47%	



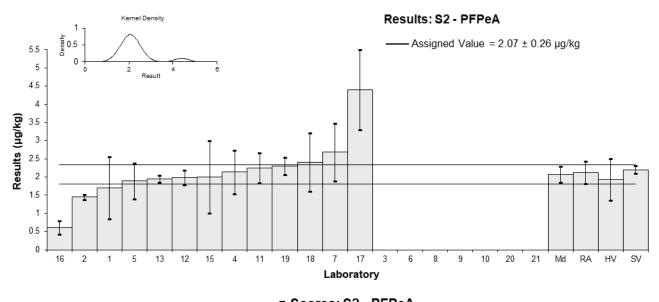
Sample No.	S2
Matrix	Carrot
Analyte	PFPeA
Unit	µg/kg

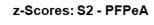
### Participant Results

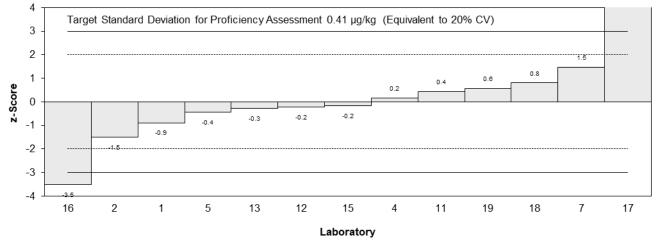
Lab. Code	Result	Uncertainty	Rec	z	En
1	1.7	0.85	126	-0.89	-0.42
2	1.45	0.071	98	-1.50	-2.30
3	NS	NS	NS		
4	2.14	0.6	70	0.17	0.11
5	1.89	0.49	NR	-0.43	-0.32
6	<1	NR	NT		
7	2.68	0.80	86	1.47	0.73
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	2.25	0.417	75.7	0.43	0.37
12	1.98	0.198	100.3	-0.22	-0.28
13	1.95	0.096	120	-0.29	-0.43
15	2	1	89	-0.17	-0.07
16**	0.612	0.1836	142	-3.52	-4.58
17*	4.4	1.1	94	5.63	2.06
18	2.4	0.8	133	0.80	0.39
19	2.3	0.24	98	0.56	0.65
20	NT	NT	NT		
21	<1	NR	NT		

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	2.07	0.26
Spike Value	2.20	0.11
Homogeneity Value	1.93	0.58
Robust Average	2.13	0.31
Median	2.07	0.22
Mean	2.26	
N	12	
Мах	4.4	
Min	1.45	
Robust SD	0.42	
Robust CV	20%	







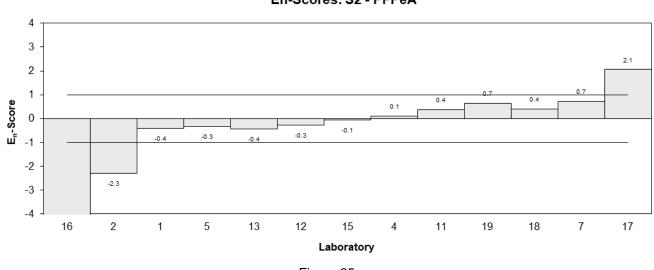


Figure 35

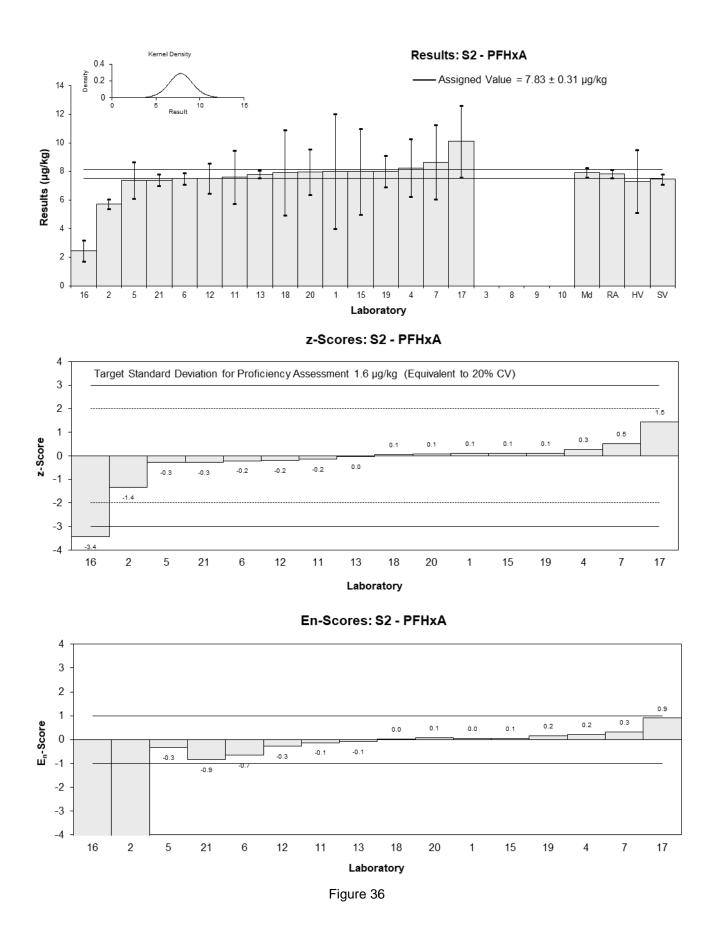
Sample No.	S2
Matrix	Carrot
Analyte	PFHxA
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	8	4	113	0.11	0.04
2	5.71	0.339	95	-1.35	-4.62
3	NS	NS	NS		
4	8.25	2	76	0.27	0.21
5	7.38	1.3	NR	-0.29	-0.34
6	7.5	0.4	NT	-0.21	-0.65
7	8.65	2.6	79	0.52	0.31
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	7.59	1.86	112.2	-0.15	-0.13
12	7.51	1.05	85.6	-0.20	-0.29
13	7.79	0.264	99	-0.03	-0.10
15	8	3	93	0.11	0.06
16**	2.44	0.732	153	-3.44	-6.78
17	10.1	2.5	94	1.45	0.90
18	7.92	3	122	0.06	0.03
19	8.0	1.1	93	0.11	0.15
20	7.95	1.59	78	0.08	0.07
21	7.4	0.4	NT	-0.27	-0.85

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	7.83	0.31
Spike Value	7.45	0.37
Homogeneity Value	7.3	2.2
Robust Average	7.83	0.31
Median	7.92	0.32
Mean	7.85	
N	15	
Мах	10.1	
Min	5.71	
Robust SD	0.48	
Robust CV	6.1%	



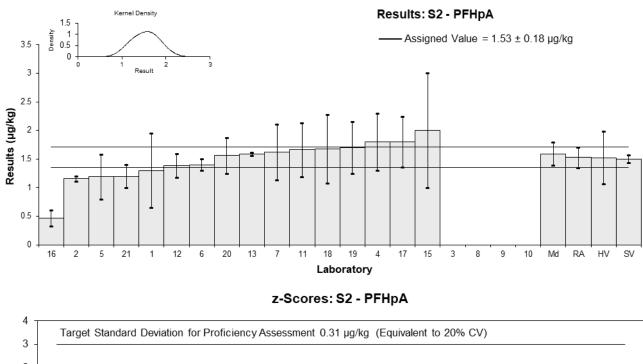
-	
Sample No.	S2
Matrix	Carrot
Analyte	PFHpA
Unit	µg/kg

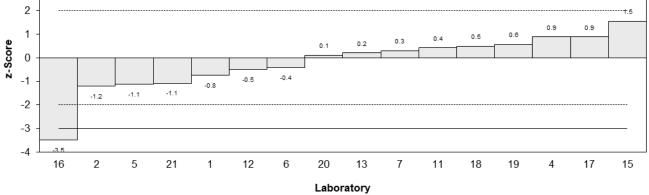
# Participant Results

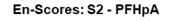
Lab. Code	Result	Uncertainty	Rec	Z	En
1	1.3	0.65	101	-0.75	-0.34
2	1.16	0.045	95	-1.21	-1.99
3	NS	NS	NS		
4	1.8	0.5	76	0.88	0.51
5	1.19	0.39	NR	-1.11	-0.79
6	1.4	0.1	NT	-0.42	-0.63
7	1.62	0.49	80	0.29	0.17
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	1.66	0.470	86.3	0.42	0.26
12	1.38	0.207	87	-0.49	-0.55
13	1.59	0.029	72	0.20	0.33
15	2	1	91	1.54	0.46
16**	0.462	0.1386	162	-3.49	-4.70
17	1.8	0.44	94	0.88	0.57
18	1.68	0.6	125	0.49	0.24
19	1.7	0.45	99	0.56	0.35
20	1.56	0.311	98	0.10	0.08
21	1.2	0.2	NT	-1.08	-1.23

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.53	0.18
Spike Value	1.50	0.07
Homogeneity Value	1.52	0.46
Robust Average	1.53	0.18
Median	1.59	0.20
Mean	1.54	
Ν	15	
Мах	2	
Min	1.16	
Robust SD	0.28	
Robust CV	18%	







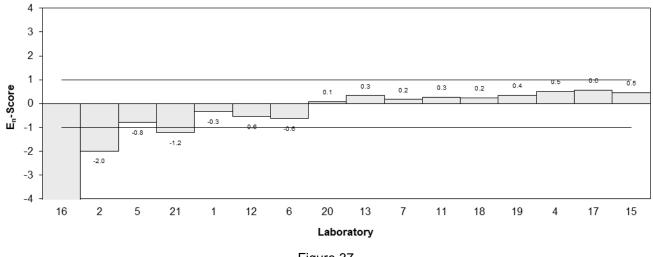


Figure 37

Sample No.	S2
Matrix	Carrot
Analyte	PFOA
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	1	0.5	111	-0.97	-0.46
2	0.935	0.068	85	-1.23	-1.96
3	NS	NS	NS		
4	1.34	0.4	78	0.40	0.24
5	1.31	0.32	NR	0.28	0.20
6	1.1	0.1	86	-0.56	-0.81
7	1.45	0.44	86	0.85	0.45
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	1.20	0.230	91.2	-0.16	-0.15
12	1.48	0.236	84.7	0.97	0.87
13	1.31	0.066	107	0.28	0.45
15	1	1	97	-0.97	-0.24
16**	0.381	0.1143	188	-3.46	-4.75
17	1.45	0.36	94	0.85	0.54
18	1.4	0.5	156	0.65	0.31
19	1.3	0.33	92	0.24	0.17
20	1.26	0.25	73	0.08	0.07
21	1.0	0.2	86	-0.97	-0.98

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.24	0.14
Spike Value	1.20	0.06
Homogeneity	1.20	0.36
Value		
Robust Average	1.24	0.14
Median	1.30	0.14
Mean	1.24	
N	15	
Мах	1.48	
Min	0.935	
Robust SD	0.21	
Robust CV	17%	

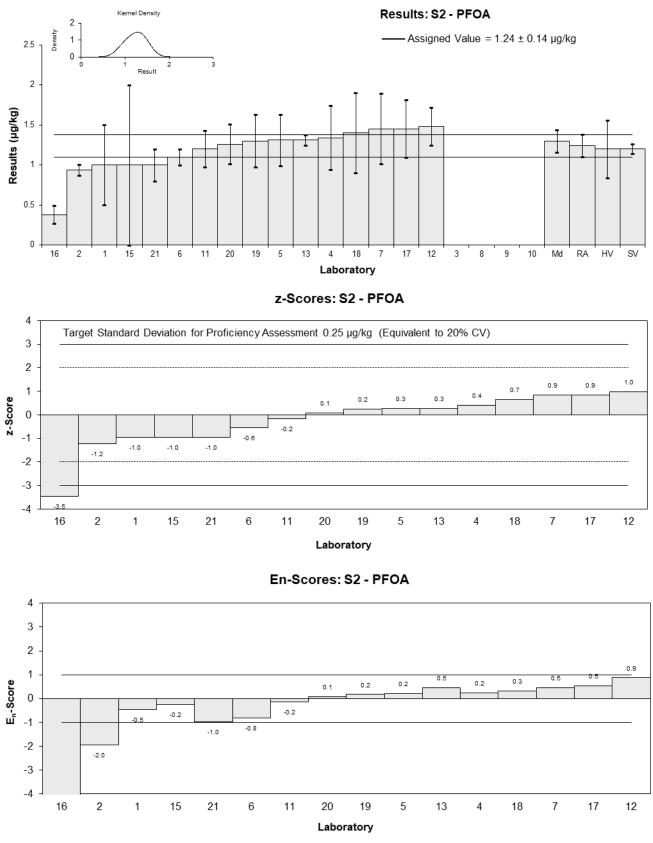


Figure 38

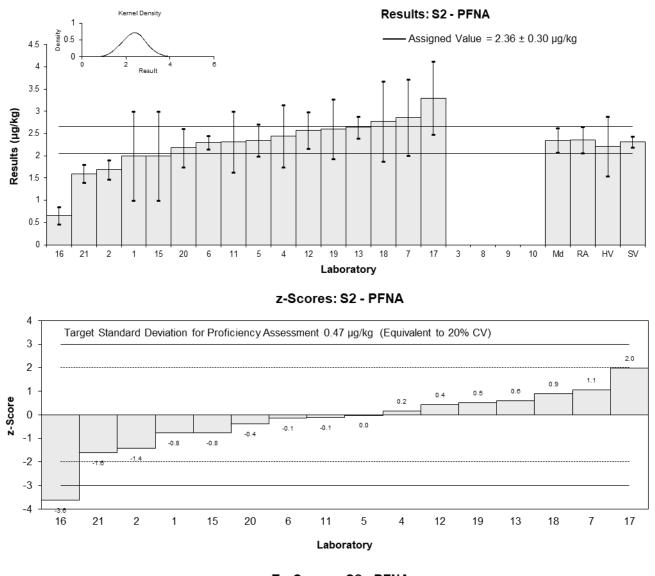
Sample No.	S2
Matrix	Carrot
Analyte	PFNA
Unit	µg/kg

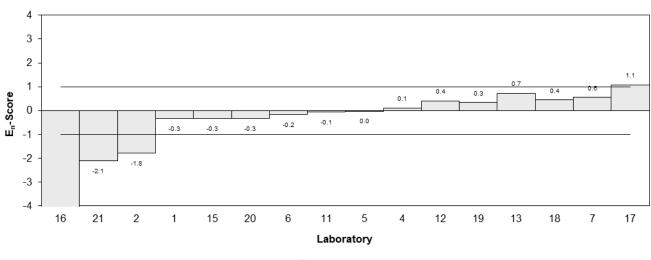
# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	2.0	1	124	-0.76	-0.34
2	1.69	0.220	85	-1.42	-1.80
3	NS	NS	NS		
4	2.44	0.7	80	0.17	0.11
5	2.35	0.36	NR	-0.02	-0.02
6	2.3	0.15	NT	-0.13	-0.18
7	2.86	0.86	84	1.06	0.55
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	2.31	0.682	90.7	-0.11	-0.07
12	2.57	0.411	92.4	0.44	0.41
13	2.64	0.25	118	0.59	0.72
15	2	1	96	-0.76	-0.34
16**	0.652	0.1956	178	-3.62	-4.77
17	3.3	0.82	94	1.99	1.08
18	2.78	0.9	145	0.89	0.44
19	2.6	0.67	99	0.51	0.33
20	2.18	0.436	83	-0.38	-0.34
21	1.6	0.2	NT	-1.61	-2.11

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	2.36	0.30
Spike Value	2.31	0.12
Homogeneity Value	2.22	0.67
Robust Average	2.36	0.30
Median	2.35	0.28
Mean	2.37	
Ν	15	
Мах	3.3	
Min	1.6	
Robust SD	0.46	
Robust CV	20%	





En-Scores: S2 - PFNA

Figure 39

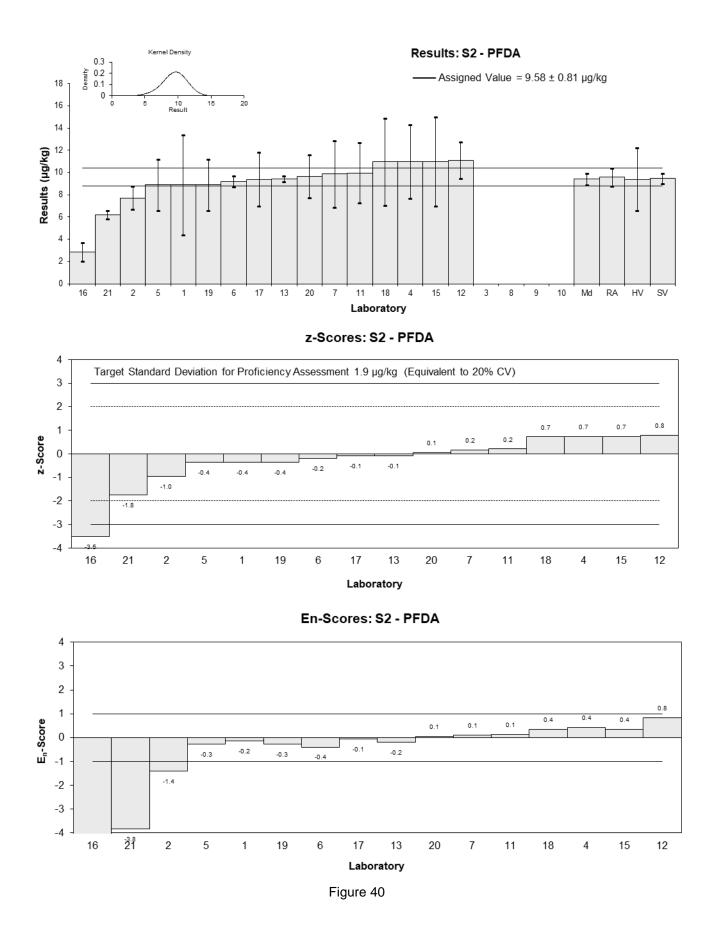
Sample No.	S2
Matrix	Carrot
Analyte	PFDA
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	8.9	4.5	102	-0.35	-0.15
2	7.72	1.04	85	-0.97	-1.41
3	NS	NS	NS		
4	11	3.3	80	0.74	0.42
5	8.89	2.3	NR	-0.36	-0.28
6	9.2	0.5	84	-0.20	-0.40
7	9.88	3.0	81	0.16	0.10
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	9.97	2.70	101.6	0.20	0.14
12	11.1	1.66	91.7	0.79	0.82
13	9.42	0.241	145	-0.08	-0.19
15	11	4	93	0.74	0.35
16**	2.85	0.855	181	-3.51	-5.71
17	9.4	2.4	94	-0.09	-0.07
18	10.96	3.9	158	0.72	0.35
19	8.9	2.3	109	-0.35	-0.28
20	9.68	1.94	44	0.05	0.05
21	6.2	0.35	84	-1.76	-3.83

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	9.58	0.81
Spike Value	9.47	0.47
Homogeneity Value	9.4	2.8
Robust Average	9.58	0.81
Median	9.42	0.51
Mean	9.48	
N	15	
Мах	11.1	
Min	6.2	
Robust SD	1.3	
Robust CV	13%	



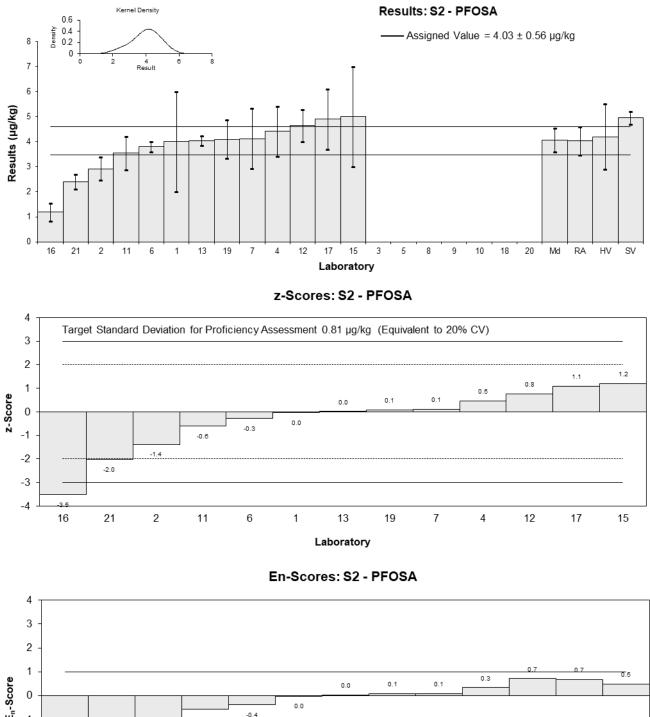
Sample No.	S2
Matrix	Carrot
Analyte	PFOSA
Unit	µg/kg

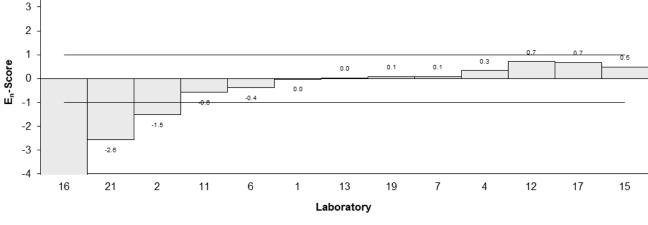
# Participant Results

Lab. Code	Result	Uncertainty	Rec	Z	En
1	4	2	103	-0.04	-0.01
2	2.92	0.458	92	-1.38	-1.53
3	NS	NS	NS		
4	4.41	1	85	0.47	0.33
5	NT	NT	NT		
6	3.8	0.2	NT	-0.29	-0.39
7	4.12	1.2	91	0.11	0.07
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	3.54	0.658	41.6	-0.61	-0.57
12	4.64	0.651	108	0.76	0.71
13	4.04	0.198	87	0.01	0.02
15	5	2	93	1.20	0.47
16**	1.19	0.357	105	-3.52	-4.28
17	4.9	1.2	94	1.08	0.66
18	<5	NR	121		
19	4.1	0.76	102	0.09	0.07
20	NT	NT	NT		
21	2.4	0.3	NT	-2.02	-2.57

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	4.03	0.56
Spike Value	4.95	0.25
Homogeneity Value	4.2	1.3
Robust Average	4.03	0.56
Median	4.07	0.47
Mean	3.99	
Ν	12	
Мах	5	
Min	2.4	
Robust SD	0.77	
Robust CV	19%	







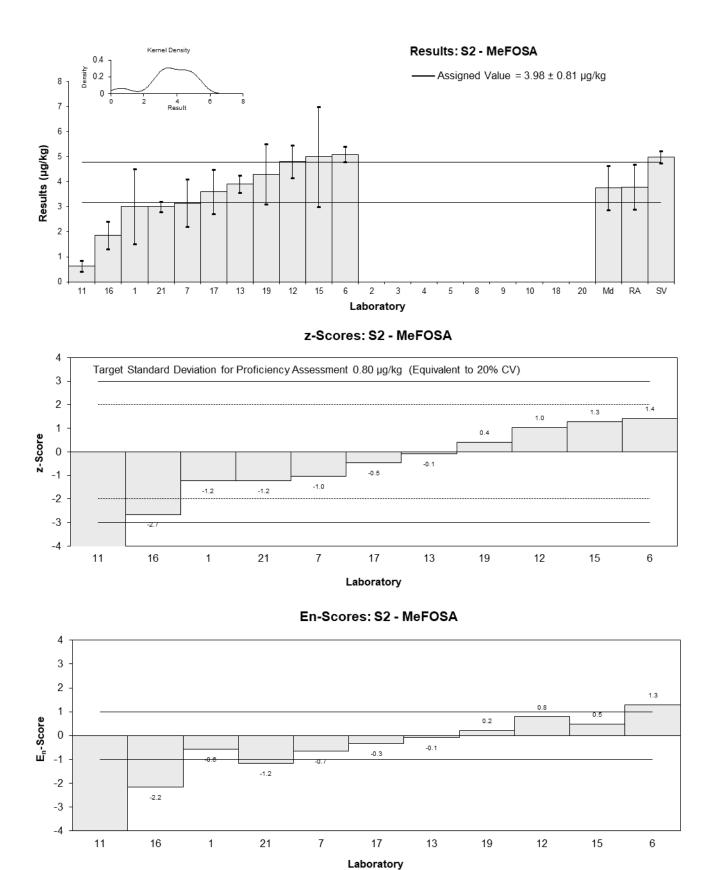
Sample No.	S2
Matrix	Carrot
Analyte	MeFOSA
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	3	1.5	78	-1.23	-0.57
2	NT	NT	NT		
3	NS	NS	NS		
4	<5	NR	36		
5	NT	NT	NT		
6	5.1	0.3	NT	1.41	1.30
7	3.15	0.95	87	-1.04	-0.66
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11*	0.620	0.215	41.6	-4.22	-4.01
12	4.8	0.651	93.5	1.03	0.79
13	3.91	0.34	45	-0.09	-0.08
15	5	2	90	1.28	0.47
16**	1.86	0.558	105	-2.66	-2.16
17	3.6	0.89	94	-0.48	-0.32
18	<5	NR	128		
19	4.3	1.2	100	0.40	0.22
20	NT	NT	NT		
21	3.0	0.2	NT	-1.23	-1.17

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	3.98	0.81
Spike Value	4.99	0.25
Robust Average	3.79	0.90
Median	3.76	0.89
Mean	3.65	
Ν	10	
Мах	5.1	
Min	0.62	
Robust SD	1.1	
Robust CV	30%	





94

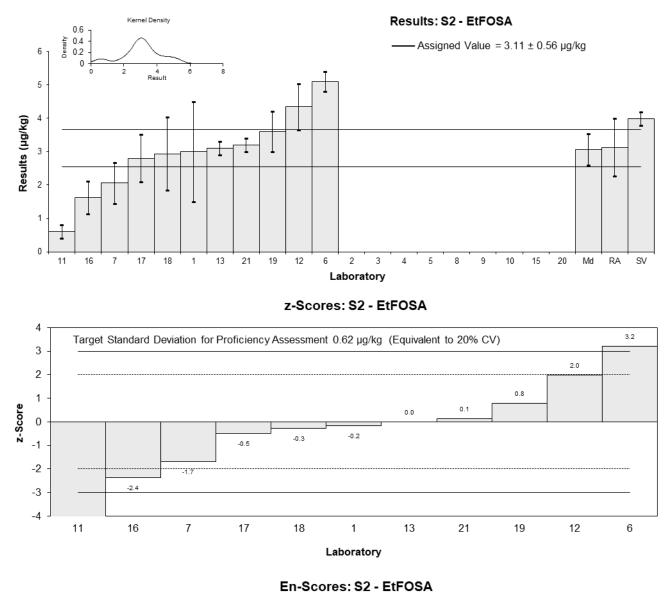
Sample No.	S2
Matrix	Carrot
Analyte	EtFOSA
Unit	µg/kg

# **Participant Results**

Lab. Code	Result	Uncertainty	Rec	Z	En
1	3	1.5	89	-0.18	-0.07
2	NT	NT	NT		
3	NS	NS	NS		
4	<5	NR	24		
5	NT	NT	NT		
6*	5.1	0.3	NT	3.20	3.13
7	2.06	0.62	89	-1.69	-1.26
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11*	0.610	0.2	41.6	-4.02	-4.20
12	4.35	0.696	73.6	1.99	1.39
13	3.11	0.201	44	0.00	0.00
15	<5	NR	92		
16**	1.63	0.489	92.4	-2.38	-1.99
17	2.8	0.71	94	-0.50	-0.34
18	2.94	1.1	105	-0.27	-0.14
19	3.6	0.61	100	0.79	0.59
20	NT	NT	NT		
21	3.2	0.2	NT	0.14	0.15

\* Outlier, \*\* Not included in robust average calculations, see Section 4.2

Assigned Value	3.11	0.56
Spike Value	3.99	0.20
Robust Average	3.13	0.86
Median	3.06	0.47
Mean	3.08	
Ν	10	
Мах	5.1	
Min	0.61	
Robust SD	1.1	
Robust CV	35%	



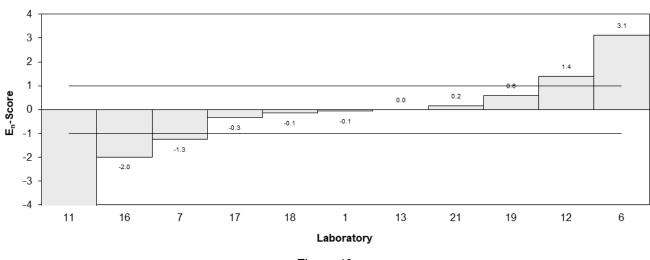




Figure 43

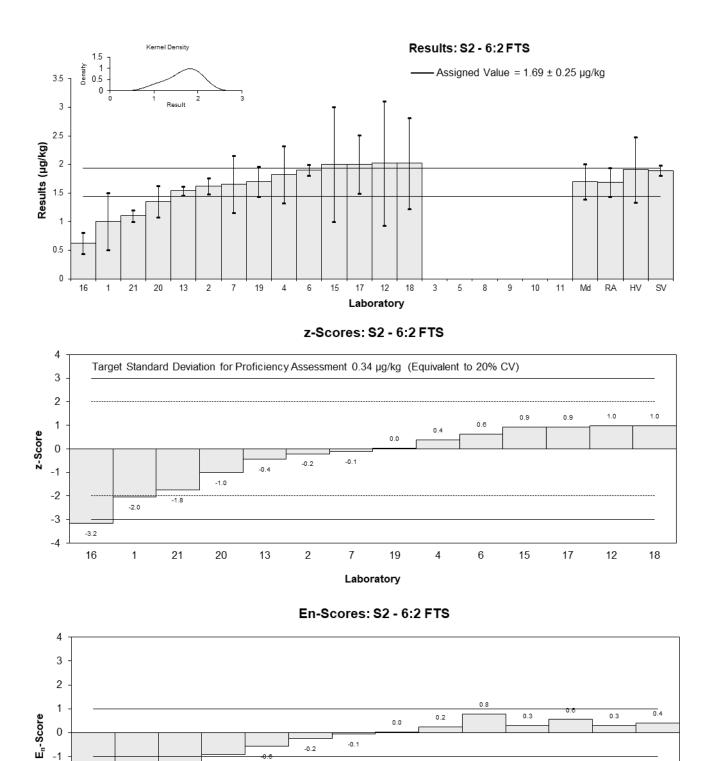
-	
Sample No.	S2
Matrix	Carrot
Analyte	6:2 FTS
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	1	0.5	313	-2.04	-1.23
2	1.62	0.142	63	-0.21	-0.24
3	NS	NS	NS		
4	1.82	0.5	59	0.38	0.23
5	NT	NT	NT		
6	1.9	0.1	NT	0.62	0.78
7	1.65	0.50	91	-0.12	-0.07
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	NT	NT	NT		
12	2.02	1.09	92.3	0.98	0.30
13	1.54	0.08	228	-0.44	-0.57
15	2	1	95	0.92	0.30
16**	0.618	0.1854	391	-3.17	-3.44
17	2.0	0.51	94	0.92	0.55
18	2.02	0.8	173	0.98	0.39
19	1.7	0.26	73	0.03	0.03
20	1.35	0.271	175	-1.01	-0.92
21	1.1	0.1	NT	-1.75	-2.19

\*\* Not included in robust average calculations, see Section 4.2

Assigned Value	1.69	0.25
Spike Value	1.89	0.09
Homogeneity Value	1.91	0.57
Robust Average	1.69	0.25
Median	1.70	0.31
Mean	1.67	
Ν	13	
Мах	2.02	
Min	1	
Robust SD	0.36	
Robust CV	21%	



-1

-2

-3

-4

-3.4

16

-1.2

1

-2.2

21

-0.9

20

13

2

7

Figure 44

Laboratory

19

4

6

15

17

12

18

98

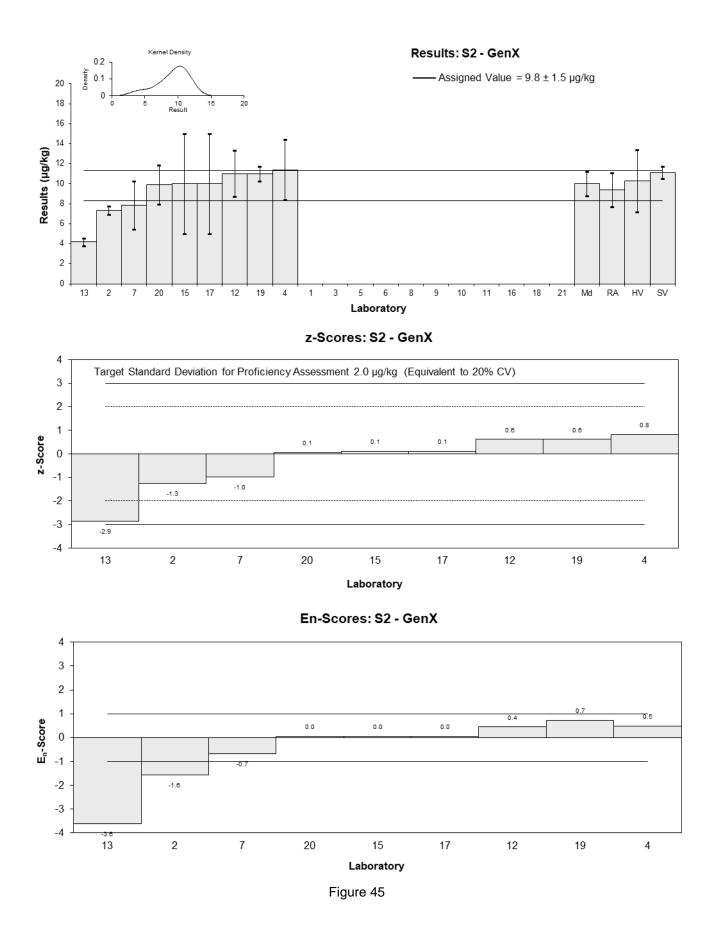
Sample No.	S2
Matrix	Carrot
Analyte	GenX
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	NT	NT	NT		
2	7.34	0.427	99	-1.26	-1.58
3	NS	NS	NS		
4	11.4	3	76	0.82	0.48
5	NT	NT	NT		
6	<1	NR	NT		
7	7.86	2.4	70	-0.99	-0.69
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	NT	NT	NT		
12	11	2.3	81.1	0.61	0.44
13*	4.19	0.382	107	-2.86	-3.62
15	10	5	89	0.10	0.04
16	NR	NR	NR		
17	10	5	94	0.10	0.04
18	NT	NT	NT		
19	11	0.72	76	0.61	0.72
20	9.90	1.98	36	0.05	0.04
21	<1	NR	NT		

\* Outlier, see Section 4.2

Assigned Value	9.8	1.5
Spike Value	11.1	0.6
Homogeneity Value	10.3	3.1
Robust Average	9.4	1.7
Median	10.0	1.2
Mean	9.2	
Ν	9	
Мах	11.4	
Min	4.19	
Robust SD	2.0	
Robust CV	22%	

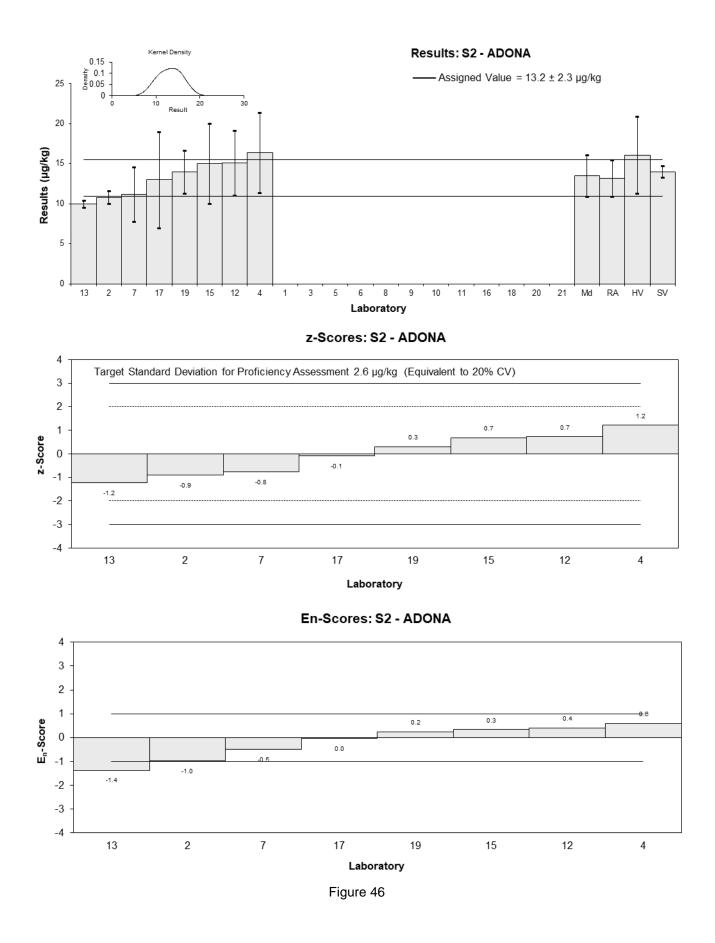


Sample No.	S2
Matrix	Carrot
Analyte	ADONA
Unit	µg/kg

# Participant Results

Lab. Code	Result	Uncertainty	Rec	z	En
1	NT	NT	NT		
2	10.8	0.783	85	-0.91	-0.99
3	NS	NS	NS		
4	16.4	5	NR	1.21	0.58
5	NT	NT	NT		
6	<1	NR	NT		
7	11.2	3.4	81	-0.76	-0.49
8	NS	NS	NS		
9	NT	NT	NT		
10	NS	NS	NS		
11	NT	NT	NT		
12	15.1	4.07	81.1	0.72	0.41
13	9.98	0.456	107	-1.22	-1.37
15	15	5	89	0.68	0.33
16	NR	NR	NR		
17	13	6	94	-0.08	-0.03
18	NT	NT	NT		
19	14	2.7	99	0.30	0.23
20	NT	NT	NT		
21	NT	NT	NT		

Assigned Value	13.2	2.3
Spike Value	14.0	0.7
Homogeneity Value	16.1	4.8
Robust Average	13.2	2.3
Median	13.5	2.6
Mean	13.2	
Ν	8	
Мах	16.4	
Min	9.98	
Robust SD	2.6	
Robust CV	20%	



# 6 DISCUSSION OF RESULTS

### 6.1 Assigned Value

The robust averages of participants' results were used as the assigned values for scored analytes. The robust averages and associated expanded uncertainties were calculated using the procedure described in ISO 13528:2022.<sup>6</sup> Outliers, gross errors and results not included in robust average calculations were removed before the calculation of the assigned value (see Section 4.2 for additional information).<sup>3,4</sup> The calculation of the expanded uncertainty for the robust average is presented in Appendix 3, using Sample S2 PFHpS as an example.

**Traceability**: The consensus of participants' results is not traceable to any external reference, so although expressed in SI units, metrological traceability has not been established.

No assigned values were set for Sample S1 PFTrDA, PFOSA, MeFOSA, EtFOSA, 11Cl-PF3OUdS, and Sample S2 PFBA, as reported results were too variable and recoveries were relatively low. The variability may have been due to difficulties in the analysis caused by the matrix, analyte mass fraction level, properties of the analyte itself, or a combination of these factors. For these analytes without assigned values, participants may still compare their results with the descriptive statistics and spiked value as presented in Section 5.

A comparison of the assigned values (or robust average if the analyte was not scored) and spiked values is presented in Table 51. For this study, the assigned values for scored analytes were within 69% to 93% and 78% to 111% of the spiked values for Samples S1 and S2 respectively. These recoveries are similar to previous NMI PFAS in biota and food PT studies, and provides good support for the assigned values and analyte stability.

Sample	Analyte	Assigned Value (Robust Average) (µg/kg)	Spiked Value (µg/kg)	Assigned Value (Robust Average) / Spiked Value (%)
PFHpS PFOS	PFBS	0.302	0.399	76
	PFPeS	4.19	4.65	90
	PFHxS	1.64	1.89	87
	PFHxS (linear)	1.61	1.89	85
	PFHpS	1.67	2.00	84
	PFOS	3.62	4.77	76
	PFOS (linear)	3.74	4.77	78
	PFNS	8.0	11.5	70
	PFBA	2.03	2.96	69
	PFPeA	1.04	1.13	92
	PFHxA	4.61	5.31	87
	PFHpA	6.02	7.54	80
	PFOA	6.44	7.92	81
	PFNA	0.427	0.503	85
	PFDA	0.74	0.902	82
	PFUdA	1.12	1.21	93
	PFTrDA	(5.4)	8.17	(66)

Table 51 Comparison of Assigned Values (Robust Averages) and Spiked Values

Sample	Analyte	Assigned Value (Robust Average) (µg/kg)	Spiked Value (µg/kg)	Assigned Value (Robust Average) / Spiked Value (%)		
	PFOSA	(3.00)	4.46	(67)		
	MeFOSA	(3.7)	4.99	(74)		
	EtFOSA	(2.66)	3.99	(67)		
	ADONA	4.5	5.64	80		
	9C1-PF3ONS	11.0	14.4	76		
	11Cl-PF3OUdS	(2.27)	4.70	(48)		
	PFBS	0.82	0.891	92		
	PFPeS	8.30	7.47	111		
	PFHxS	6.54	6.61	99		
	PFHxS (linear)	6.44	6.61	97		
	PFHpS	2.99	3.00	100		
	PFOS	1.95	2.12	92		
	PFOS (linear)	2.01	2.12	95		
	PFNS	1.52	1.72	88		
	PFDS	6.76	6.80	99		
	PFBA	(0.78)	1.19	(66)		
S2	PFPeA	2.07	2.20	94		
(Carrot)	PFHxA	7.83	7.45	105		
	PFHpA	1.53	1.50	102		
	PFOA	1.24	1.20	103		
	PFNA	2.36	2.31	102		
	PFDA	9.58	9.47	101		
	PFOSA	4.03	4.95	81		
	MeFOSA	3.98	4.99	80		
	EtFOSA	3.11	3.99	78		
	6:2 FTS	1.69	1.89	89		
	GenX	9.8	11.1	88		
	ADONA	13.2	14.0	94		

# 6.2 Measurement Uncertainty Reported by Participants

Participants were asked to report an estimate of the expanded MU associated with their results and the basis of this uncertainty estimate. It is a requirement of ISO/IEC 17025 that laboratories have procedures to estimate the uncertainty of chemical measurements and to report this in specific circumstances, including when the client's instruction so requires.<sup>8</sup>

Of 597 numeric results reported for spiked analytes in this study, 580 (97%) were reported with an uncertainty. Laboratory **1** did not report uncertainties for linear isomers PFHxS and PFOS in Sample S2 only (uncertainties were reported for these analytes in Sample S1); this participant reported they were accredited to ISO/IEC 17025. Laboratory **3** did not report any uncertainties; this participant reported that they were not accredited.

Laboratories **1** and **8** attached an estimate of MU to at least one non-numeric result reported. An uncertainty expressed as a value should not be attached to a non-value result.<sup>9</sup>

Participants' procedures for estimating their uncertainty are presented in Table 3. Three participants reported using the NATA GAG Estimating and Reporting MU as their guide; NATA no longer publishes this document.<sup>10</sup>

The magnitude of the MUs for analytes in this study was within the range 1.8% to 100% of the reported value. In general, an expanded uncertainty of less than 10% relative is likely to be unrealistically small for the routine analysis of PFAS, while over 50% is likely too large and not fit for purpose. Of the 580 MUs, 436 (75%) were between 10% and 50% relative, 126 were less than 10% relative and 18 were greater than 50% relative.

Uncertainties associated with results returning a satisfactory *z*-score but an unsatisfactory  $E_n$ -score may have been underestimated.

In some cases, results and/or uncertainties were reported with an inappropriate number of significant figures. Including too many significant figures may inaccurately reflect the precision of measurements. The recommended format is to write the uncertainty to no more than two significant figures and then to write the result with the corresponding number of decimal places. For example, instead of  $4.51 \pm 1.2822 \,\mu g/kg$ , it is better to report this as  $4.5 \pm 1.3 \,\mu g/kg$ .<sup>9</sup>

## 6.3 *z*-Score

Target SDs equivalent to 20% PCV were used to calculate *z*-scores. CVs predicted by the Thompson-Horwitz equation,<sup>7</sup> the between-laboratory CVs obtained in this study, and the target SDs (as PCVs) are presented for comparison in Table 52.

Sample	Analyte	Assigned Value (µg/kg)	Thompson-Horwitz CV (%)	Between-Laboratory CV* (%)	Target SD (as PCV) (%)
	PFBS	0.302	22	16	20
	PFPeS	4.19	22	21	20
	PFHxS	1.64	22	21	20
	PFHxS (linear)	1.61	22	21	20
	PFHpS	1.67	22	27	20
	PFOS	3.62	22	27	20
	PFOS (linear)	3.74	22	26	20
	PFNS	8.0	22	33	20
<b>S</b> 1	PFBA	2.03	22	29	20
(Prawn)	PFPeA	1.04	22	14	20
	PFHxA	4.61	22	19	20
	PFHpA	6.02	22	17	20
	PFOA	6.44	22	18	20
	PFNA	0.427	22	20	20
	PFDA	0.74	22	25	20
	PFUdA	1.12	22	27	20
	PFTrDA	Not Set	NA	61	Not Set
	PFOSA	Not Set	NA	37	Not Set

Table 52 Comparison of Thompson-Horwitz CVs, Between-Laboratory CVs, and Target SDs

Sample	Analyte	Assigned Value (µg/kg)	Thompson-Horwitz CV (%)	Between-Laboratory CV* (%)	Target SD (as PCV) (%)
	MeFOSA	Not Set	NA	37	Not Set
	EtFOSA	Not Set	NA	41	Not Set
	ADONA	4.5	22	28	20
	9C1-PF3ONS	11.0	22	22	20
	11Cl-PF3OUdS	Not Set	NA	36	Not Set
	PFBS	0.82	22	16	20
	PFPeS	8.30	22	13	20
	PFHxS	6.54	22	11	20
	PFHxS (linear)	6.44	22	11	20
	PFHpS	2.99	22	12	20
	PFOS	1.95	22	13	20
	PFOS (linear)	2.01	22	11	20
	PFNS	1.52	22	19	20
	PFDS	6.76	22	21	20
	PFBA	Not Set	NA	47	Not Set
S2	PFPeA	2.07	22	17	20
(Carrot)	PFHxA	7.83	22	6.1	20
	PFHpA	1.53	22	18	20
	PFOA	1.24	22	17	20
	PFNA	2.36	22	20	20
	PFDA	9.58	22	13	20
	PFOSA	4.03	22	19	20
	MeFOSA	3.98	22	24	20
	EtFOSA	3.11	22	20	20
	6:2 FTS	1.69	22	21	20
	GenX	9.8	22	17	20
	ADONA	13.2	22	20	20

\* Robust between-laboratory CV (outliers removed where applicable). Shaded cells are between-laboratory CVs for scored analytes which were higher than both the target SD and the Thompson-Horwitz CV.

To account for possible low bias in the consensus value due to laboratories using inefficient analytical or extraction techniques, two *z*-scores were adjusted across Sample S1 PFNS and PFBA. A maximum acceptable result was set to two target SDs more than the spiked value, and results lower than the maximum acceptable result but with a *z*-score greater than 2.0 had their *z*-score adjusted to 2.0. This ensured that laboratories reporting results close to the spiked value were not penalised. *z*-Scores for results higher than the maximum acceptable value were not adjusted, and *z*-scores less than 2.0 were left unaltered.

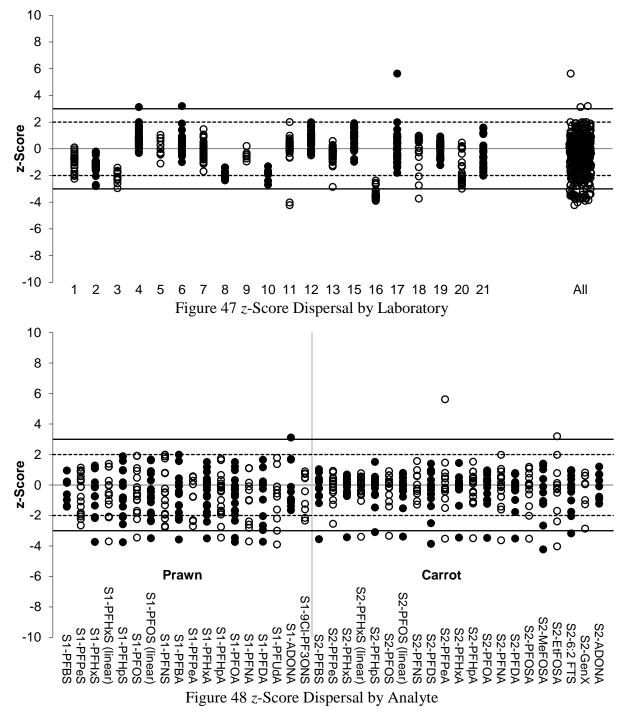
Of 528 results for which *z*-scores were calculated, 453 (86%) returned  $|z| \le 2.0$ , indicating a satisfactory performance.

Sixteen participants analysed both samples, with Laboratories **7**, **12**, **13** and **17** reporting numeric results for all 39 scored analytes. Laboratories **7** and **12** returned satisfactory *z*-scores for all analytes. Laboratories **19** (33), **15** (32) and **9** (7) returned satisfactory *z*-scores for all reported numeric results.

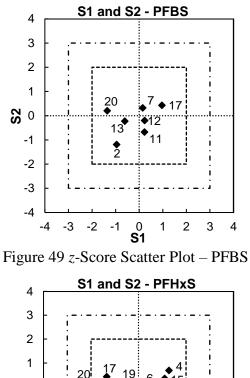
Three participants were sent Sample S1 Prawn only. Laboratory **5** was sent Sample S2 carrot only, and returned satisfactory *z*-scores for all reported numeric results (10).

Laboratory **16** returned questionable or unsatisfactory *z*-scores for all results (31), with all being lower than the assigned value (negative bias; *z*-scores ranging from -3.90 to -2.38).

The dispersal of participants' *z*-scores is presented graphically by laboratory in Figure 47 and by analyte in Figure 48.



Scatter plots of *z*-scores for analytes present in both Samples S1 and S2 are presented in Figures 49 to 63. Scores are predominantly in the upper right and lower left quadrants, indicating that laboratory bias is the major contributor to the variability of results. Points close to the diagonal axis demonstrate excellent repeatability, while points close to the zero demonstrate excellent repeatability and accuracy.



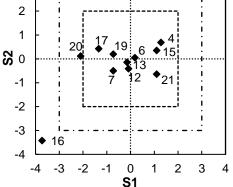


Figure 51 z-Score Scatter Plot – PFHxS

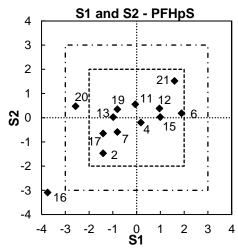


Figure 53 z-Score Scatter Plot – PFHpS

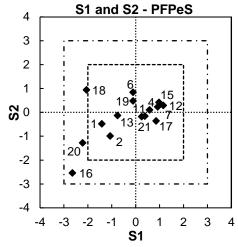


Figure 50 z-Score Scatter Plot – PFPeS

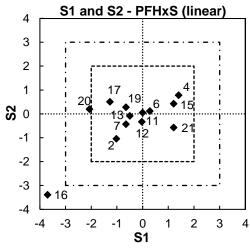


Figure 52 z-Score Scatter Plot – PFHxS (linear)

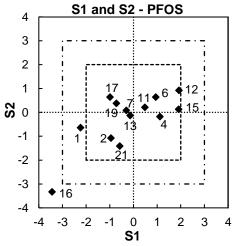


Figure 54 z-Score Scatter Plot – PFOS

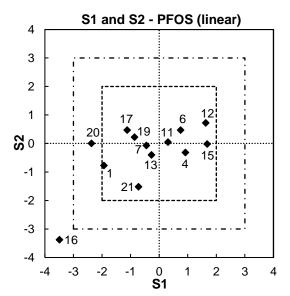
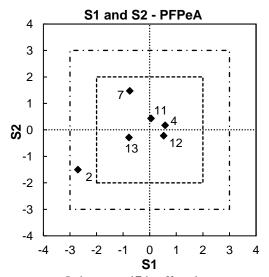


Figure 55 z-Score Scatter Plot – PFOS (linear)



Laboratory **17** is off-scale. Figure 57 *z*-Score Scatter Plot – PFPeA

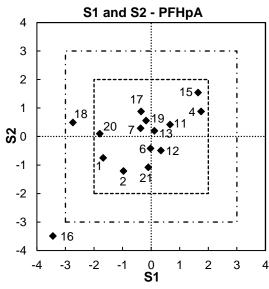
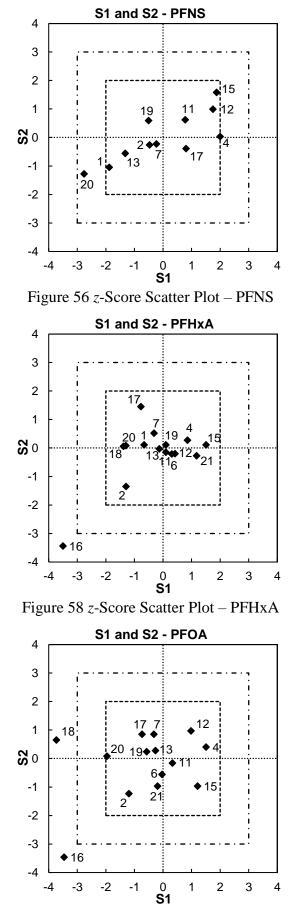
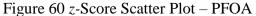


Figure 59 z-Score Scatter Plot – PFHpA





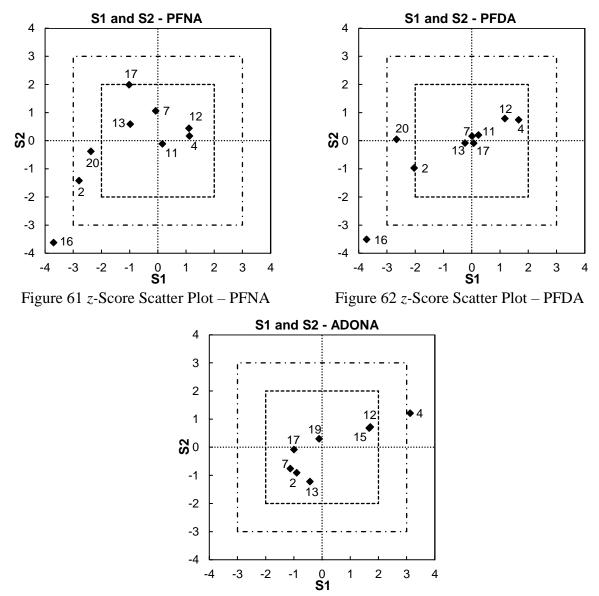


Figure 63 z-Score Scatter Plot – ADONA

## 6.4 *E<sub>n</sub>*-Score

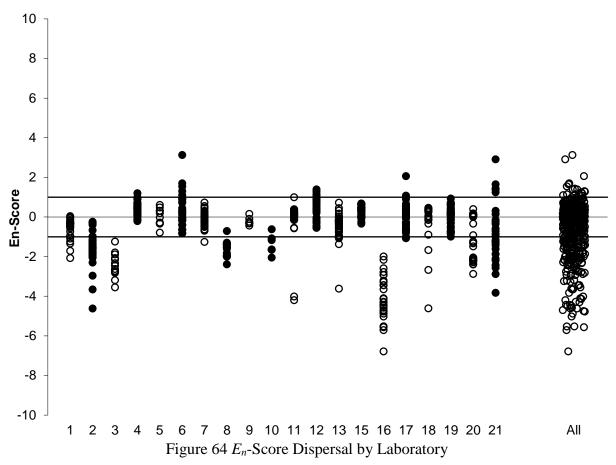
 $E_n$ -scores can be interpreted in conjunction with *z*-scores, as an unsatisfactory  $E_n$ -score can either be caused by issues with measurement, or uncertainty, or both. If a participant did not report any uncertainty with a result, an expanded uncertainty of zero (0) was used to calculate the  $E_n$ -score.  $E_n$ -scores greater than 1.0 were set to 1.0 for results with *z*-scores that were adjusted as discussed in Section 6.3 *z*-Score.

Of 528 results for which  $E_n$ -scores were calculated, 374 (71%) returned  $|E_n| \le 1.0$ , indicating agreement of the participant's result with the assigned value within their respective expanded uncertainties.

No participant returned satisfactory  $E_n$ -scores for all analytes of interest in this study. Of the participants analysing both matrices, Laboratories **19** (33), **15** (32) and **9** (7) returned satisfactory  $E_n$ -scores for all reported results. Of participants analysing Sample S2 carrot only, Laboratory **5** (10) returned satisfactory  $E_n$ -scores for all reported results.

Laboratories 16 (31) and 3 (14) returned unsatisfactory  $E_n$ -scores for all reported results.

The dispersal of participants'  $E_n$ -scores is presented graphically in Figure 64.



## 6.5 Range of PFAS Analysed by Participants

Participants were provided with a list of analytes that may have been spiked into the test samples (Table 1). Of these, 24 different analytes were spiked for this study, with 21 analytes being spiked into Sample S1, and 20 analytes being spiked into Sample S2. For PFHxS and PFOS, participants were requested to report for both linear isomers and total value, however both samples were spiked with linear only isomers. Participants were not required to test for all potential analytes, and were requested to report 'NT' (for 'Not Tested') for any analyte they did not test the samples for.

A summary of participants' testing of the spiked analytes is presented in Table 53.

Of the participants who analysed both samples, Laboratories **4**, **7**, **12**, **13**, **15**, **17** and **19** analysed for all spiked analytes. Of the participants who only received Sample S1 prawn, Laboratories **8** and **10** analysed for all spiked analytes in this sample. Laboratories **6** and **9** reported analysing some analytes in one sample but not the other. All participants tested for at least one spiked analyte, with the proportion of PFAS being analysed by each participant ranging from 31% to 100%.

Out of the spiked analytes in this study, PFBS, PFHxA, PFHpA, PFOA, PFNA and PFDA were tested for by the highest proportion of participants (100% for all). In general, perfluoroalkyl acids were very well represented by participants, with the overall proportion of analysis by participants being 89% and 95% for perfluoroalkane sulfonates and perfluoroalkyl carboxylic acids respectively. A lower proportion of participants analysed the perfluoroalkane sulfonamide (PFOSA), perfluoroalkane sulfonamido (MeFOSA and EtFOSA) and fluorotelomer (6:2 FTS) analytes, being 80%, 75% and 82% respectively. The PFAS replacement compounds (GenX, ADONA, 9CI-PF3ONS and 11CI-PF3OUdS) were analysed by the lowest proportion of participants (65%).

Lab. Code Analyte	1	2	3	4	5	6	7	8	9 <sup>d</sup>	10	11	12	13	15	16	17	18	19	20	21	Proportion of Participants (%)
PFBS	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	100								
PFPeS	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	90										
PFHxS	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	85								
PFHxS (linear)	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	90													
PFHpS	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	90										
PFOS	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	90
PFOS (linear)	$\checkmark$	NT	$\checkmark$	95																	
PFNS	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	75
PFDS <sup>c</sup>	NT	$\checkmark$		$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$		NT		$\checkmark$	88									
PFBA	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	90															
PFPeA	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	90															
PFHxA	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	100								
PFHpA	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	100								
PFOA	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	100								
PFNA	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	100								
PFDA	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	100								
PFUdA <sup>b</sup>	$\checkmark$	$\checkmark$	NT	$\checkmark$		$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	89										
PFTrDA <sup>b</sup>	$\checkmark$	$\checkmark$	NT	$\checkmark$		$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	89										
PFOSA	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	80								
MeFOSA	$\checkmark$	NT	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	75								
EtFOSA	$\checkmark$	NT	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	75								
6:2 FTS <sup>c</sup>	$\checkmark$	$\checkmark$		$\checkmark$	NT	$\checkmark$	$\checkmark$		NT		NT	$\checkmark$	82								

Table 53 Summary of PFAS Analysed by Participants<sup>a</sup>

Lab. Code Analyte	1	2	3	4	5	6	7	8	9 <sup>d</sup>	10	11	12	13	15	16	17	18	19	20	21	Proportion of Participants (%)
GenX <sup>c</sup>	NT	$\checkmark$		$\checkmark$	NT	$\checkmark$	$\checkmark$		NT		NT	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	71
ADONA	NT	$\checkmark$	$\checkmark$	$\checkmark$	NT	√ <sup>e</sup>	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	NT	65
9C1-PF3ONS <sup>b</sup>	NT	$\checkmark$	$\checkmark$	$\checkmark$		NT	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	NT	63
11Cl-PF3OUdS <sup>b</sup>	NT	$\checkmark$	$\checkmark$	$\checkmark$		NT	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$	NT	$\checkmark$	NT	NT	63
Proportion of Analytes (%)	81	92	65	100	50	92	100	100	31	100	77	100	100	100	96	100	77	100	69	88	

<sup>a</sup> Shaded cells indicate that the participant did not enrol for and was not supplied the sample containing that analyte; proportions have been adjusted accordingly.

<sup>b</sup> Spiked into Sample S1 only.

<sup>c</sup> Spiked into Sample S2 only.

<sup>d</sup> Laboratory **9** enrolled for and was supplied both samples in this study, however reported 'NT' for all analytes in Sample S2. This participant's data in this table has been completed according to their Sample S1 results where available.

<sup>e</sup> Laboratory **6** reported analysing for ADONA in Sample S2 but not Sample S1.

## 6.6 PFAS in Food Trigger Points

There are currently no maximum regulatory limits in Australia for PFAS contaminants in food. However, Food Standards Australia New Zealand (FSANZ) has proposed non-regulatory 'trigger points' in a variety of food products for 3 common PFAS compounds, namely PFHxS, PFOS and PFOA, based on food consumption rates and set tolerable daily intakes for these analytes.<sup>11</sup> Where an analyte is found to be exceeding the corresponding trigger point, this may indicate that further investigation is required.

The assigned values in this study and relevant FSANZ trigger points are given in Table 54. With the relatively high trigger points for crustaceans, Sample S1 PFHxS, PFOS and PFOA were all well below the trigger points. For Sample S2, PFHxS and PFOS were above the trigger points, while PFOA was under the trigger point.

Sample	Matrix	Iatrix Classification	PFHxS	(µg/kg)	PFOS (	(µg/kg)	PFOA (µg/kg)		
Sample	wiatiix	Classification	Assigned Value	Trigger Point	Assigned Value	Trigger Point	Assigned Value	Trigger Point	
S1	Prawn	Crustaceans	$1.64\pm0.27$	65	$3.62\pm0.68$	65	$6.44\pm0.85$	520	
S2	Carrot	Vegetables	$6.54\pm0.51$	1.1	$1.95\pm0.17$	1.1	$1.24\pm0.14$	8.8	

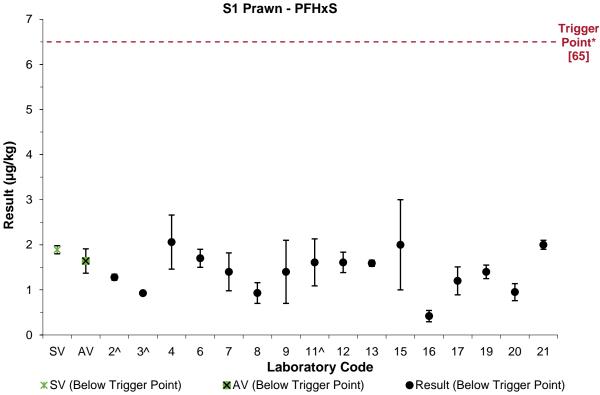
Table 54 Assigned Values and FSANZ Trigger Points for PFHxS, PFOS and PFOA<sup>11</sup>

Figures 65 to 70 show comparisons of the spiked values (SV), assigned values (AV), participants' results, and FSANZ trigger points for these analytes. Where no numeric result was reported, or if a LOR was reported, these results have been excluded from consideration. Where a participant did not report the total value of an analyte, but did report a linear isomers only value, the linear value has been plotted.

In this study, five of the six assessed analytes were either significantly higher or lower than the trigger points, and so it was expected that the vast majority of participants' results should match the assigned values with respect to being above or below the FSANZ trigger points. This was seen in this study, with 97 results of a total 100 (97%) being correctly above or below the trigger point inclusive of uncertainty, and a further two results being correctly above or below the trigger point with uncertainty spanning the trigger point.

Laboratories 2, 4, 6, 7, 11, 12, 13, 17, 19, 20 and 21 correctly identified whether the analyte mass fractions (inclusive of uncertainties) were above or below the trigger points for all six analytes, while Laboratories 18 (4), 3 (3), 5 (3), 8 (3), 9 (3) and 10 (2) did so for all analytes that they reported results for.

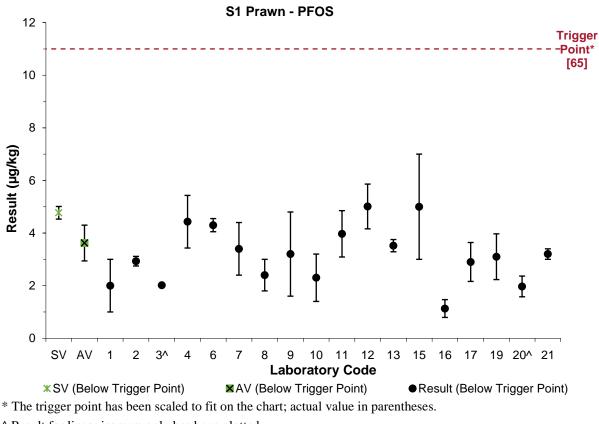
For Sample S2 PFOS, the assigned value above but closer to the trigger point as compared to the other analytes. In this sample, two participants reported results where the uncertainties spanned the trigger point. Laboratory **16** reported a result below the trigger point inclusive of uncertainty, which would have incorrectly indicated no need for further investigation.



\* The trigger point has been scaled to fit on the chart; actual value in parentheses.

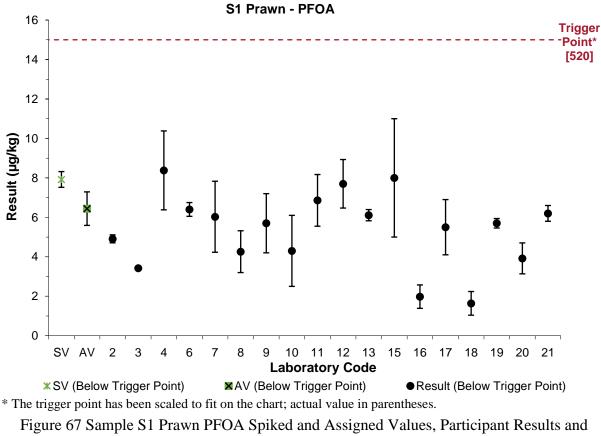
^ Result for linear isomers only has been plotted.

Figure 65 Sample S1 Prawn PFHxS Spiked and Assigned Values, Participant Results and Trigger Point



^ Result for linear isomers only has been plotted.

Figure 66 Sample S1 Prawn PFOS Spiked and Assigned Values, Participant Results and Trigger Point



**Trigger Point** 

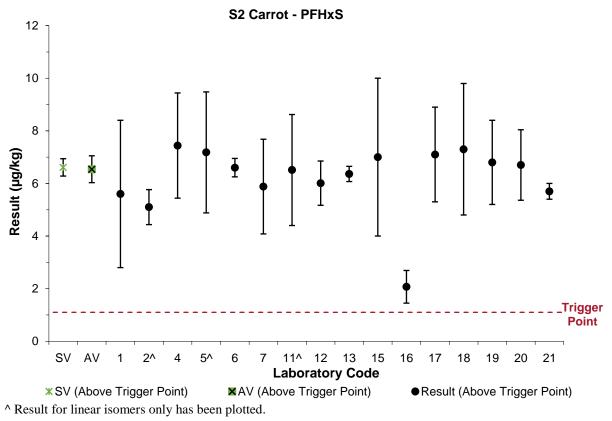


Figure 68 Sample S2 Carrot PFHxS Spiked and Assigned Values, Participant Results and Trigger Point

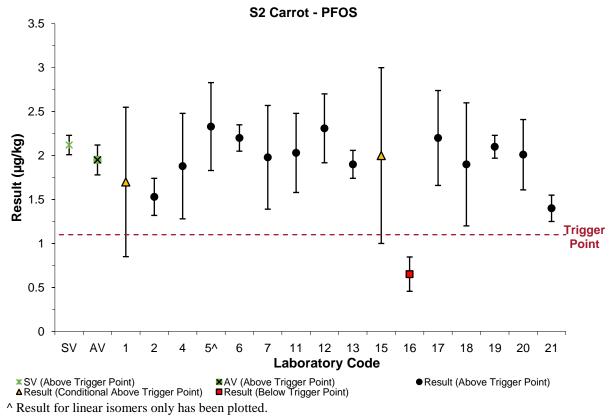
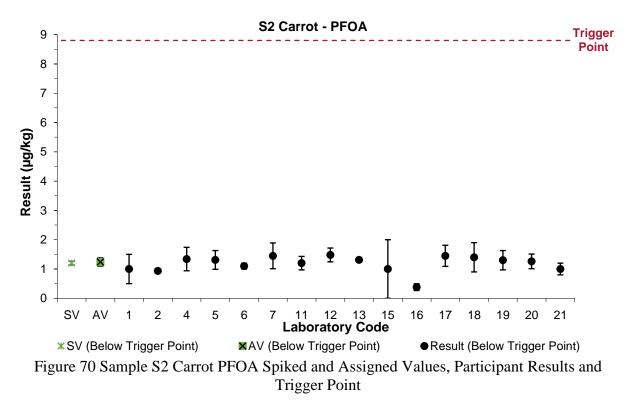


Figure 69 Sample S2 Carrot PFOS Spiked and Assigned Values, Participant Results and Trigger Point



## 6.7 False Negatives

Table 55 presents false negative results. These are analytes present in the samples which a participant tested for, but did not report a result; for example, when participants reported a 'less-than' result (< x) when the assigned value was higher than their limit of reporting (LOR), or did not report anything. For analytes where no assigned value was set, results have only been considered to be false negatives where the robust average and spiked value were significantly higher than the participants' LOR, or if no value was reported.

Lab. Code	Sample	Analyte	Assigned Value (Robust Average) (µg/kg)	Spiked Value (µg/kg)	Result* (µg/kg)
		PFHpS	1.67	2.00	< 1
1	<b>S</b> 1	PFBA	2.03	2.96	< 2
		PFOA	6.44	7.92	< 5
	S1	PFHxS	1.64	1.89	NR
2	51	PFOS (linear)	3.74	4.77	NR
2	S2	PFHxS	6.54	6.61	NR
	52	PFOS (linear)	2.01	2.12	NR
	S1	PFPeA	1.04	1.13	<1
	51	PFUdA	1.12	1.21	<1
6		PFNS	1.52	1.72	<1
0	S2	PFPeA	2.07	2.20	<1
	32	GenX	9.8	11.1	<1
		ADONA	13.2	14.0	<1
8	S1	PFDA	0.74	0.902	<0.5

Lab. Code	Sample	Analyte	Assigned Value (Robust Average) (µg/kg)	Spiked Value (µg/kg)	Result* (µg/kg)
		PFPeS	4.19	4.65	NR
		PFHxS	1.64	1.89	<1
		PFHxS (linear)	1.61	1.89	NR
		PFHpS	1.67	2.00	<1
		PFOS (linear)	3.74	4.77	NR
		PFPeA	1.04	1.13	<1
		PFDA	0.74	0.902	NR
10	S1	PFUdA	1.12	1.21	<1
		PFTrDA	(5.4)	8.17	NR
		PFOSA	(3.00)	4.46	NR
		MeFOSA	(3.7)	4.99	NR
		EtFOSA	(2.66)	3.99	NR
		ADONA	4.5	5.64	NR
		9C1-PF3ONS	11.0	14.4	NR
		11Cl-PF3OUdS	(2.27)	4.70	NR
		PFBS	0.302	0.399	< 0.1
		PFPeA	1.04	1.13	< 0.3
	S1	ADONA	4.5	5.64	NR
16		9C1-PF3ONS	11.0	14.4	NR
16		11Cl-PF3OUdS	(2.27)	4.7	NR
		PFBA	(0.78)	1.19	< 0.3
	S2	GenX	9.8	11.1	NR
		ADONA	13.2	14	NR
		PFHxS	1.64	1.89	<1
		PFHpS	1.67	2	<1
10	61	PFOS	3.62	4.77	<1
18	S1	PFOS (linear)	3.74	4.77	<1
		PFUdA	1.12	1.21	<1
		PFTrDA	(5.4)	8.17	<2
	61	PFPeA	1.04	1.13	< 1.0
10	S1	PFUdA	1.12	1.21	< 1.0
19	52	PFBS	0.82	0.891	NR
	S2	PFBA	(0.78)	1.19	NR
20	S1	PFOS	3.62	4.77	NR
	G 1	PFPeA	1.04	1.13	<1
21	S1	PFUdA	1.12	1.21	<1
	S2	PFNS	1.52	1.72	<1

Lab. Code	Sample	Analyte	Assigned Value (Robust Average) (µg/kg)	Spiked Value (µg/kg)	Result* (µg/kg)
		PFPeA	2.07	2.2	<1
		GenX	9.8	11.1	<1

\* NR results may or may not be false negatives, depending on the participant's actual LOR.

#### 6.8 Reporting of Additional Analytes

Seven participants reported at least one analyte that was not spiked into the test samples by the study coordinator. These results are presented in Table 56.

Lab. Code	Sample	Analyte	Result (µg/kg)	Uncertainty (µg/kg)	Recovery (%)
2	S1	PFDoA	0.025	0.006	56
7	S1	PFDoA	0.120	0.12	81
	S1	PFDS	0.0414	0.0268	93.2
11	51	PFDoA	0.155	0.035	127
	S2	PFDoS	0.0386	0.0204	148.5
12	<b>S</b> 1	PFDoA	0.19	0.0418	71.1
16	<b>S</b> 1	PFDoA	0.1	0.03	165
17	<b>S</b> 1	PFDoA	0.11	0.077	89
	<b>S</b> 1	PFDoA	0.053	0.011	76
20	20 S2		0.281	0.056	83
	32	PFDoA	0.010	0.004	50

Table 56 Non-Spiked Analytes Reported by Participants

#### 6.9 Participants' Methods

Participants were requested to analyse the samples using their normal test method and to report a single result as they would normally report to a client. Methodologies as provided by participants are presented in Appendix 4. A summary is presented below as technique (number of participants):

- Sample Weight
  - o S1: < 1 g (2), 1 1.1 g (8), 2 2.5 g (3), 5 6 g (1)
  - S2: <1 g (1), 1 1.1 g (3), 2 2.5 g (5), 5 6 g (2), 10 g (1)
- Pretreatment
  - S1: homogenisation (10), no pretreatment (2)
  - S2: homogenisation (9), freeze-drying (1), no pre-treatment (2)
- Extraction Technique
  - S1: alkaline digestion (5), QuEChERS (3), SLE (8)
  - S2: alkaline digestion (4), QuEChERS (3), SLE (7), Soxhlet (1)
- Extraction Solvent
  - S1: acetonitrile (4), acetonitrile/acid(/water) (3), acetonitrile/base (2), methanol/base (3), multiple/other (2)

- S2: acetonitrile (3), acetonitrile/acid(/water) (2), acetonitrile/base (1), methanol/base (3), multiple/other (3)
- Extraction Temperature
  - S1: room temperature (13), heated (2)
  - S2: room temperature (10), heated/boiling (3)
- Extraction Time (total)
  - S1: 8 min (1), 30 min (5), 40 min (1), 1 h (2), 2 h (1), 3 h (1), 8 h (1), 16 h (1)
  - S2: 8 min (1), 30 min (5), 1 h (2), 2 h (1), 4 h (1), 8 h (1), 16 h (1)
- Clean-Up
  - S1: SPE / dSPE (carbon: 8, WAX: 2, C18: 2, other / not specified: 6), centrifugation (1), filtration (1), LLE (1)
  - S2: SPE / dSPE (carbon: 7, WAX: 2, C18: 2, other / not specified: 4), centrifugation (1), filtration (1), ion pair separation (1)

• S2: Yes (2), No (6)

• S2: Yes (2), No (11)

- Instrument
  - o S1: LC-MS/MS or LC-QQQ (14), LC-Orbitrap (2)
  - S2: LC-MS/MS or LC-QQQ (12), LC-Orbitrap (2)
- Dilution
  - S1: Yes (2), No (7)
- Guard Column

   S1: Yes (10), No (5)
   S2: Yes (11), No (3)
- Delay Column
  - S1: Yes (15), No (1) S2: Yes (13), No (1)
- Blank Correction
  - S1: Yes (2), No (12)
- Labelled Standard Source
  - S1: Wellington Laboratories (15) S2: Wellington Laboratories (13)
- Recovery Correction

Participants reported a very broad range of recoveries, ranging from 2% to 391%, though the vast majority were between 50% and 150%.

• S1: Yes (11), No (5) • S2: Yes (9), No (5)

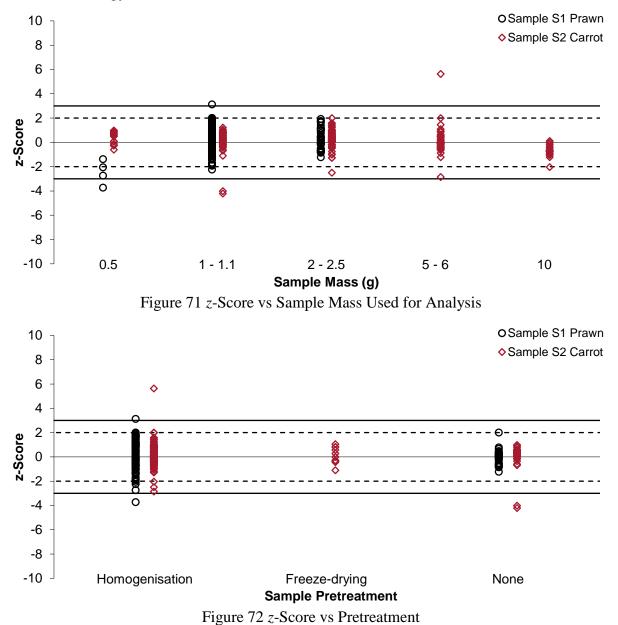
As discussed in Section 4.2, the results from Laboratories **3**, **8**, **10**, **16** and **20** in Sample S1, and from Laboratory **16** in Sample S2, were consistently lower than the spiked value by around the same factor for all analytes. This is an indication of laboratory or method bias. These participants should check their sample or standard preparation/dilution procedures. Therefore, these results have been excluded from the following methodology analysis.

Comparisons of *z*-scores with various extraction and analysis parameters are given in Figures 71 to 78. The most popular methodology for this study was homogenisation as pretreatment, followed by alkaline digestion with basified methanol and SPE clean-up, and then analysis on LC-MS/MS.

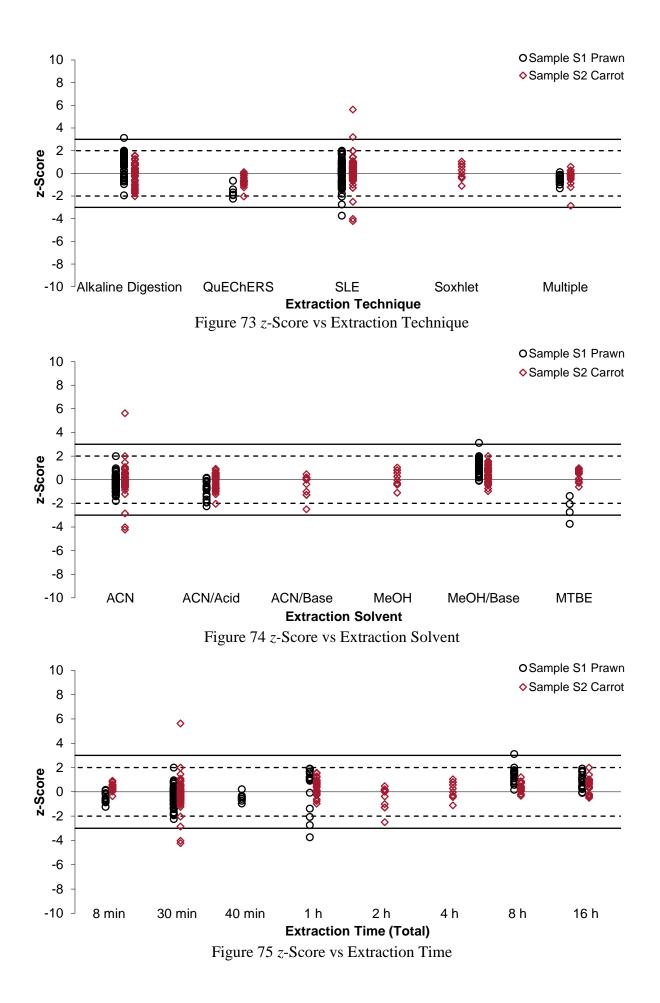
In this study, it was observed that participants using basified methanol as their extraction solvent, or those who had long extraction times (> 8 h), generally achieved higher *z*-scores. As the ratios of assigned values to spiked values were generally below 100% in this study, higher satisfactory *z*-scores may correspond to more efficient extraction parameters.

A participant reported using 0.5 g for Sample S1 prawn analysis, and the results reported by this participant were biased low. Caution should be exercised when a small sample size (e.g. < 1 g) is taken for analysis, as this may not be a suitable representation of the whole sample.

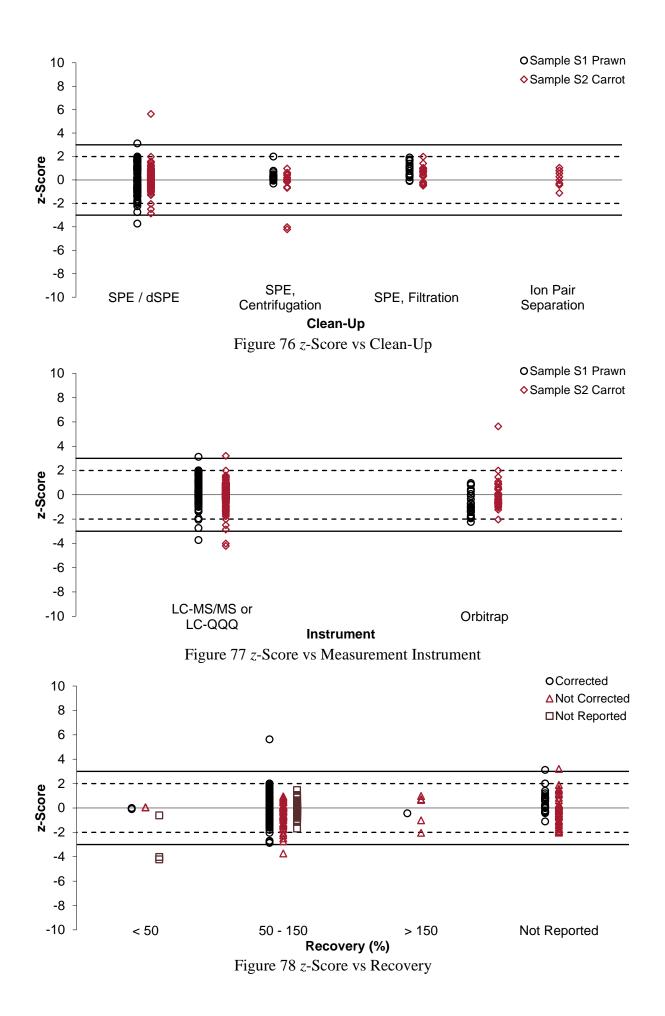
One participant used QuEChERS for extraction; this participant's results were generally satisfactory though biased low. Another participant used MTBE as the extraction solvent; their results for Sample S1 prawn were biased low. These participants may need to review if their methodology introduced bias to their measurements.



AQA 22-14 PFAS in Biota and Food

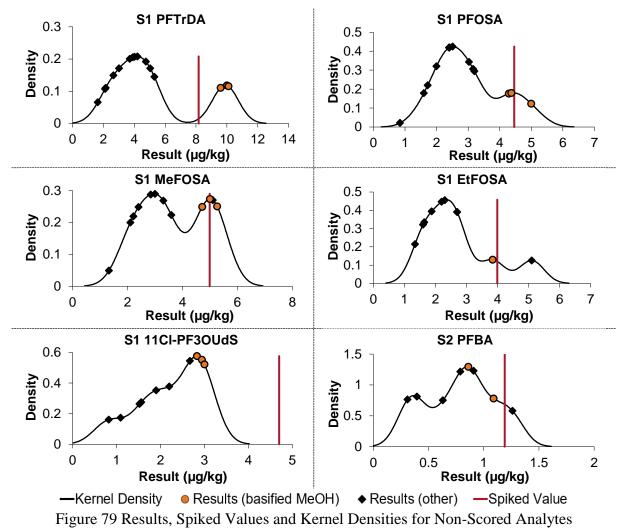


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AQA 22-14 PFAS in Biota and Food

Assigned values were not able to be set for a number of analytes in this study as the results reported by participants were not compatible (Section 6.1). For these analytes, it was seen that participants using basified methanol as their extraction solvent generally reported results closer to the spiked value (Figure 79), similar to what was observed for the scored analytes in this study.



6.10 Total vs Linear Isomers – PFHxS and PFOS

Participants were requested to report both the linear isomers and the total (sum of linear and branched isomers) for PFHxS and PFOS. A summary of results reported by participants is presented in Table 57. The majority of participants reported numeric results for both linear and total.

Table 57 Number of Participants Reporting Numeric PFHxS and PFOS Results

Sampla		PFHxS			PFOS	
Sample	Linear and Total	Linear Only	Total Only	Linear and Total	Linear Only	Total Only
<b>S</b> 1	12	3	1	13	2	3
S2	12	3	1	14	1	1

In this study, both samples were only spiked with linear PFHxS and linear PFOS standards, and therefore the linear to total ratio was expected to be 100% for all.

# PFHxS

Summaries of participants' results for linear and total PFHxS in Samples S1 and S2 are presented in Figures 80 and 81.

Of the participants reporting numeric results for both linear and total PFHxS, the majority correctly reported the same result, or very similar results, for both. However, for Sample S1, Laboratory **13** reported a lower linear value compared to their total value (91% of total), and their results were not in agreement with each other within their respective uncertainties.

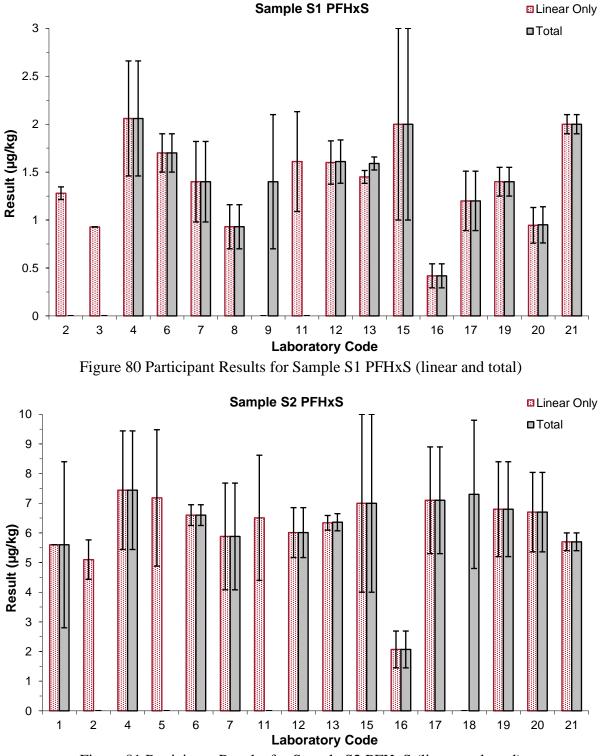


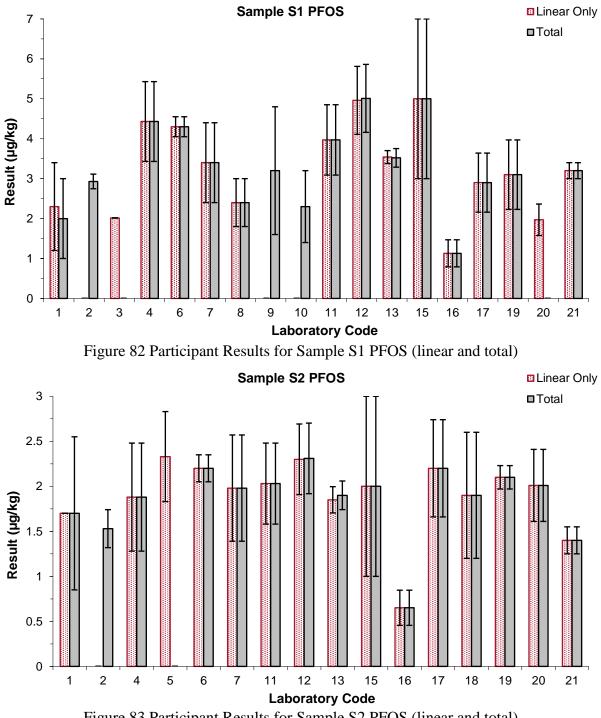
Figure 81 Participant Results for Sample S2 PFHxS (linear and total)

# PFOS

Summaries of participants' results for linear and total PFOS in Samples S1 and S2 are presented in Figures 82 and 83.

Laboratory **20** reported numeric results for both linear only and total PFOS in Sample S2, but only reported a linear result for Sample S1 PFOS (with their total result being 'NR').

Of the participants reporting numeric results for both linear and total PFOS, the majority of participants correctly reported the same result, or very similar results, for both, and all were in agreement with each other within their respective uncertainties. For Sample S1, Laboratory **1** reported a higher value for linear isomers (115% of total).



## 6.11 Effects of Sample Matrix

The samples in this study were spiked prawn (Sample S1) and carrot (Sample S2). A summary of the results reported and *z*-scores obtained by matrix is presented in Table 58.

Participants overall performed better with the carrot matrix, with a higher proportion of numeric results reported and a higher proportion of satisfactory *z*-scores.

Sample	Matrix	Expected Number of Results	Numeric Results Reported	<i>z</i> -Scores Calculated	Satisfactory z-Scores
<b>S</b> 1	Prawn	437	305 (70%)	244	195 (80%)
S2	Carrot	374	292 (78%)	284	258 (91%)

 Table 58 Result Comparison by Matrix

6.12	Summary	of Participants'	<b>Results and</b>	Performances
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Summaries of participants' results and performances for scored analytes in this PT study are presented in Tables 59 and 60, and Figure 84. Table 59 Summary of Participants' Sample S1 Results\*

Lab. Code	PFBS	PFPeS	PFHxS	PFHxS (linear)	PFHpS	PFOS	PFOS (linear)	PFNS	PFBA	PFPeA	PFHxA	PFHpA	PFOA	PFNA	PFDA	PFUdA	ADONA	9C1-PF3ONS
AV	0.302	4.19	1.64	1.61	1.67	3.62	3.74	8.0	2.03	1.04	4.61	6.02	6.44	0.427	0.74	1.12	4.5	11.0
SV	0.399	4.65	1.89	1.89	2.00	4.77	4.77	11.5	2.96	1.13	5.31	7.54	7.92	0.503	0.902	1.21	5.64	14.4
1	< 2	3	< 2	< 2	< 1	2	2.3	5	< 2	< 2	4	4	< 5	< 2	< 5	< 2	NT	NT
2	0.244	3.29	NR	1.28	1.20	2.93	NR	7.25	1.48	0.479	3.42	4.85	4.91	0.188	0.438	0.760	3.69	7.47
3	0.216	2.662	NT	0.928	0.967	NT	2.015	NT	1.104	0.562	2.979	3.958	3.426	0.206	0.304	NT	3.013	6.268
4	<0.5	4.68	2.06	2.06	1.73	4.43	4.43	11.3	2.67	1.16	5.4	8.13	8.38	0.523	0.986	1.52	7.31	12.6
5	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
6	<1	4.1	1.7	1.7	2.3	4.3	4.3	6.4	<5	<1	4.9	6.0	6.4	<1	<1	<1	NT	NT
7	0.311	4.96	1.40	1.40	1.40	3.4	3.4	7.63	1.86	0.885	4.32	5.58	6.03	0.42	0.741	0.994	3.48	12.1
8	<0.5	2.57	0.93	0.93	0.99	2.40	2.40	4.49	1.16	0.55	2.80	4.33	4.26	< 0.5	<0.5	0.63	3.25	5.88
9	<0.5	NT	1.4	NT	NT	3.2	NT	NT	NT	NT	4.8	5.5	5.7	0.4	0.6	NT	NT	NT
10	<1	NR	<1	NR	<1	2.3	NR	3.7	1.5	<1	2.4	3.7	4.3	<1	NR	<1	NR	NR
11	0.315	4.51	NT	1.61	1.65	3.97	3.97	9.24	2.89	1.05	4.70	6.82	6.86	0.441	0.776	1.05	NT	NT
12	0.316	5.15	1.61	1.6	1.99	5.01	4.96	10.8	2.21	1.15	5	6.43	7.7	0.521	0.913	1.43	6.04	12.7
13	0.265	3.55	1.59	1.45	1.34	3.52	3.54	5.89	1.79	0.878	4.49	6.15	6.11	0.343	0.705	0.953	4.11	10.1
15	<1	5	2	2	2	5	5	11	2	<2	6	8	8	<1	<2	<2	6	13
16	< 0.1	1.97	0.418	0.418	0.418	1.13	1.13	NT	0.582	< 0.3	1.38	1.88	1.98	0.111	0.19	0.246	NR	NR
17	0.36	4.9	1.2	1.2	1.2	2.9	2.9	9.3	1.3	1.1	3.9	5.6	5.5	0.34	0.75	1.1	3.6	12
18	<1	2.46	<1	NT	<1	<1	<1	NT	<5	<2	3.34	2.72	1.64	<1	<1	<1	NT	NT
19	< 1.0	4.1	1.4	1.4	1.4	3.1	3.1	7.2	2.1	< 1.0	4.7	5.8	5.7	< 1.0	< 1.0	< 1.0	4.4	8.3
20	0.220	2.33	0.950	0.945	0.811	NR	1.97	3.58	NT	NT	3.41	3.85	3.92	0.224	0.346	0.450	NT	NT
21	<1	4.4	2.0	2.0	2.2	3.2	3.2	4.9	<5	<1	5.7	5.9	6.2	<1	<1	<1	NT	NT

\* AV = Assigned Value; SV = Spiked Value. All values are in  $\mu g/kg$ . Shaded cells are results which returned a questionable or unsatisfactory *z*-score.

Lab. Code	PFBS	PFPeS	PFHxS	PFHxS (linear)	PFHpS	PFOS	PFOS (linear)	PFNS	PFDS	PFPeA	PFHxA	PFHpA	PFOA	PFNA	PFDA	PFOSA	MeFOSA	EtFOSA	6:2 FTS	GenX	ADONA
AV	0.82	8.30	6.54	6.44	2.99	1.95	2.01	1.52	6.76	2.07	7.83	1.53	1.24	2.36	9.58	4.03	3.98	3.11	1.69	9.8	13.2
HV	0.83	8.3	5.9	5.9	2.62	1.88	1.88	1.51	6.4	1.93	7.3	1.52	1.20	2.22	9.4	4.2	-	-	1.91	10.3	16.1
SV	0.891	7.47	6.61	6.61	3.00	2.12	2.12	1.72	6.80	2.20	7.45	1.50	1.20	2.31	9.47	4.95	4.99	3.99	1.89	11.1	14.0
1	0.7	7.5	5.6	5.6	2.6	1.7	1.7	1.2	NT	1.7	8	1.3	1	2.0	8.9	4	3	3	1	NT	NT
2	0.625	6.65	NR	5.10	2.11	1.53	NR	1.44	6.35	1.45	5.71	1.16	0.935	1.69	7.72	2.92	NT	NT	1.62	7.34	10.8
3	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
4	0.957	8.47	7.44	7.44	2.87	1.88	1.88	1.53	7.22	2.14	8.25	1.8	1.34	2.44	11	4.41	<5	<5	1.82	11.4	16.4
5	0.99	NT	NT	7.18	NT	NT	2.33	NT	7.86	1.89	7.38	1.19	1.31	2.35	8.89	NT	NT	NT	NT	NT	NT
6	<1	9.7	6.6	6.6	3.1	2.2	2.2	<1	6.6	<1	7.5	1.4	1.1	2.3	9.2	3.8	5.1	5.1	1.9	<1	<1
7	0.873	8.68	5.88	5.88	2.64	1.98	1.98	1.45	6.80	2.68	8.65	1.62	1.45	2.86	9.88	4.12	3.15	2.06	1.65	7.86	11.2
8	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
9	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
10	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
11	0.709	8.04	NT	6.51	3.32	2.03	2.03	1.71	8.08	2.25	7.59	1.66	1.20	2.31	9.97	3.54	0.620	0.610	NT	NT	NT
12	0.787	8.78	6.01	6.01	3.22	2.31	2.3	1.82	8.67	1.98	7.51	1.38	1.48	2.57	11.1	4.64	4.8	4.35	2.02	11	15.1
13	0.782	8.08	6.36	6.34	3	1.9	1.85	1.35	5.57	1.95	7.79	1.59	1.31	2.64	9.42	4.04	3.91	3.11	1.54	4.19	9.98
15	<1	9	7	7	3	2	2	2	8	2	8	2	1	2	11	5	5	<5	2	10	15
16	0.238	4.09	2.07	2.07	1.14	0.651	0.651	NT	1.55	0.612	2.44	0.462	0.381	0.652	2.85	1.19	1.86	1.63	0.618	NR	NR
17	0.89	7.7	7.1	7.1	2.6	2.2	2.2	1.4	6.0	4.4	10.1	1.8	1.45	3.3	9.4	4.9	3.6	2.8	2.0	10	13
18	<1	9.86	7.3	NT	3.04	1.9	1.9	NT	5.94	2.4	7.92	1.68	1.4	2.78	10.96	<5	<5	2.94	2.02	NT	NT
19	NR	9.1	6.8	6.8	3.2	2.1	2.1	1.7	8.0	2.3	8.0	1.7	1.3	2.6	8.9	4.1	4.3	3.6	1.7	11	14
20	0.853	6.17	6.70	6.70	3.27	2.01	2.01	1.13	3.38	NT	7.95	1.56	1.26	2.18	9.68	NT	NT	NT	1.35	9.90	NT
21	<1	8.0	5.7	5.7	3.9	1.4	1.4	<1	4.9	<1	7.4	1.2	1.0	1.6	6.2	2.4	3.0	3.2	1.1	<1	NT

# Table 60 Summary of Participants' Sample S2 Results\*

\* AV = Assigned Value; HV = Homogeneity Value; SV = Spiked Value. All values are in µg/kg. Shaded cells are results which returned a questionable or unsatisfactory z-score.

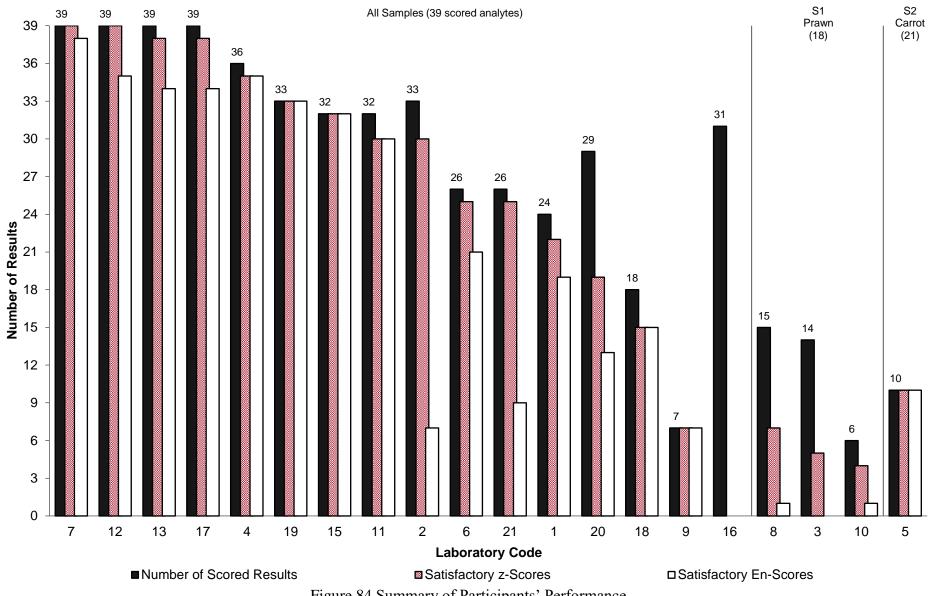


Figure 84 Summary of Participants' Performance

## 6.13 Comparison with Previous PFAS in Biota and Food Studies

NMI has coordinated PFAS in Biota and Food PT studies since 2016. A summary of participation and reported results rates over the last seven studies (2016 to 2022) is presented in Figure 85. Proportions of PFAS analysed and numeric results reported have remained relatively high over this period, despite the increased number of spiked analytes as compared to the original studies.

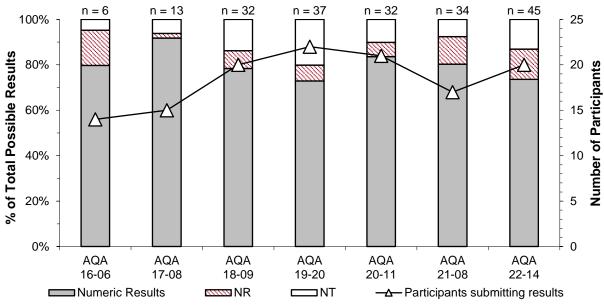


Figure 85 Summary of Participation and Reported Results in PFAS in Biota and Food PT Studies (n = number of spiked analytes).

A summary of the satisfactory performance (presented as a percentage of the total number of scores for each study) in PFAS in Biota and Food PT studies over the last seven studies (2016 to 2022) is presented in Figure 86. The target SD used to calculate *z*-scores has been kept constant at 20% PCV, which enables comparison between different studies. Proportions of satisfactory scores has remained relatively consistent, with the average proportion of satisfactory scores over this period being 89% for *z*-scores and 76% for  $E_n$ -scores.

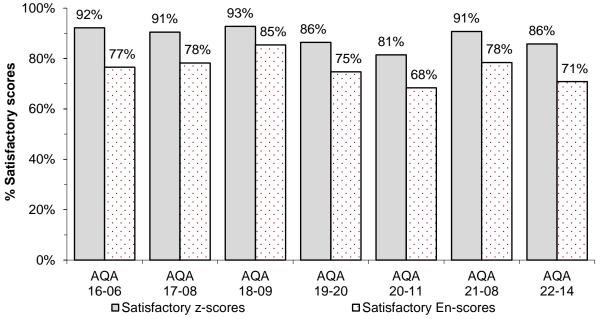


Figure 86 Summary of Participants' Performance for PFAS in Biota and Food PT Studies

The number of analytes assessed in each study has increased significantly as compared to the initial PFAS in Biota and Food study, and the studies have increased in size and complexity. As a point of comparison, PFOS and PFOA have been assessed in every study, and a summary of the proportion of satisfactory scores for these analytes over the last seven studies is presented in Figure 87.

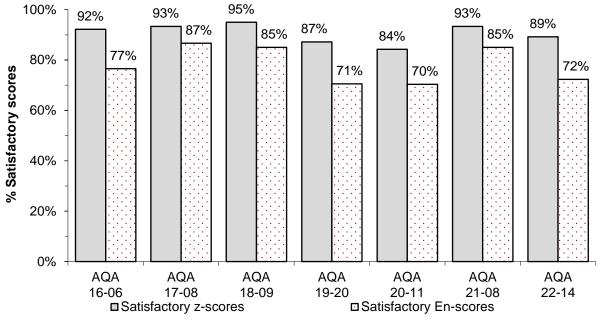
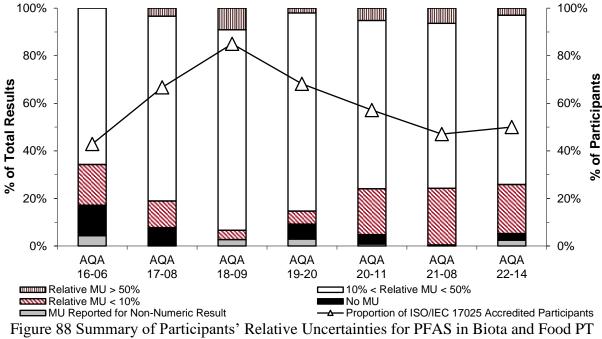


Figure 87 Summary of Participants' Performance for PFOS and PFOA in Biota and Food PT Studies

Individual performance history reports are emailed to participants at the end of each PT study; the consideration of *z*-scores over time provides much more useful information than a single *z*-score. Over time, laboratories should expect at least 95% of their *z*-scores to lie within the range  $|z| \le 2.0$ . Scores in the range  $2.0 \le |z| < 3.0$  can occasionally occur, however these should be interpreted in conjunction with the other scores obtained by that laboratory. For example, a trend of *z*-scores on one side of the zero line is an indication of method or laboratory bias.

As discussed in Section 6.2, it is a requirement of ISO/IEC 17025 that laboratories report their uncertainty. Figure 88 presents a summary of relative uncertainties as reported by participants over the last seven studies (2016 to 2022). Over this period, the vast majority of results were reported with uncertainties (95%), despite only around 60% participants reporting that they were accredited to ISO/IEC 17025. A small proportion of reported results consisted of numeric results with no uncertainty, or non-numeric results with uncertainties; the proportion of such results has reduced in recent studies as compared to the first PFAS in Biota and Food study run in 2016. Over the last three studies in particular, there has been an increased number of participants reporting potentially unrealistically small or large relative uncertainties for routine PFAS measurements (i.e. less than 10% or larger than 50% relative).

Participants reporting results with satisfactory *z*-scores, but with smaller relative uncertainties and unsatisfactory  $E_n$ -scores, may need to assess whether their uncertainties have been underestimated.



Studies

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## **APPENDIX 1 SAMPLE PREPARATION**

**Sample S1:** Prawns were blended to yield 550.6 g of puree. The pureed prawns were placed in a tray and sprayed with a spiking solution containing PFAS analytes in methanol. The prawns were thoroughly mixed, before being divided into patties of no more than 6 cm in diameter, placed on a tray, covered, and placed into the freezer overnight at -80 °C. The frozen patties were then ground using a Retsch SM2000 Knife Mill which was kept cold using liquid nitrogen and dry ice. The dry ice was then allowed to sublime off, before 5 g portions of the spiked prawns were packed into sample tubes. The tubes were labelled, shrink-wrapped, and then stored at -80 °C prior to dispatch.

**Sample S2:** Organic carrots were bought from a Sydney organic fruit and vegetable wholesaler. The carrots were rinsed, cut and blended. A stainless steel tray was lined with aluminium foil and the carrot was spread evenly over the tray. The tray was tilted at 45 degrees and a prepared composite in methanol was sprayed over the carrot with regular mixing steps to homogenise the carrot. The spiked carrot was then formed into patties of no more than 6 cm in diameter and placed on trays which were covered with baking paper. The trays were then placed into a freezer over the weekend at -80 °C. The frozen patties were then ground using a Retsch SM2000 Knife Mill which was kept cold using liquid nitrogen and dry ice. The dry ice was then allowed to sublime off, before of at least 25 g of the spiked carrot was packed into sample tubes. The tubes were labelled, shrink-wrapped, and then stored at -80 °C prior to dispatch.

## APPENDIX 2 HOMOGENEITY AND STABILITY

No homogeneity or stability testing was conducted on Sample S1 prawn. This sample was prepared and packaged using a process previously demonstrated to produce homogeneous and stable PT samples; in particular, stability for PFAS analytes in prawn at room temperature has been demonstrated for at least two months previously.<sup>5</sup>

## A2.1 Homogeneity and Stability Testing – Sample S2 Carrot

## **Homogeneity Testing**

Homogeneity and stability was conducted for Sample S2 carrot, which was prepared using a new method for PFAS in fresh produce, though the process used was similar to the preparation of PFAS in biota or meat samples.

Samples were analysed by the NMI Australian Ultra Trace Laboratory. For homogeneity testing, measurements were made under repeatability conditions in a random order. Samples were prepared in duplicate by accurately weighing 1 g of the sample then spiking with  $100 \,\mu L$ of labelled internal standard in methanol. The samples were extracted by overnight tumbling in alkaline methanol (0.01 N potassium hydroxide), then centrifuged and a portion was purified by passing through activated carbon (SUPLCLEAN ENVI-CARB, 500 mg, 120-400 Mesh) eluted using methanol. After evaporation under nitrogen, the extract was reconstituted to 600 µL in mobile phase and spiked with 20 µL labelled recovery standard in methanol. All chemicals were analytical reagents or LCMS grade solvents. Instrument analysis was performed using an Ultra Performance Liquid Chromatography (UPLC) coupled with a Liquid Chromatography Qtrap Mass Spectrometer (ABSciex 6500+), operating in multiple reaction monitoring mode. 2 µL of extract was injected onto a Waters Acquity BEH C18 column (2.1 mm x 100 mm x 1.7 µm, 130 Å) with a mobile phase gradient consisting of water:methanol (2 mM ammonium acetate). Two mass transitions were monitored for each target analyte and labelled internal standard, and abundance ratios checked. The instrument mass accuracy was calibrated annually during preventative maintenance, and the six point calibration curve established for each analytical batch. A solvent batch blank was extracted and analysed with each batch, and sample results were reported if results were at least three times the level of any analyte detected in the batch blank. Quantification was based on the use of the labelled internal standards using relative retention factors from the multipoint calibration, and was corrected for internal standard recoveries. The analysis was based on USEPA Draft Method 1633 and used calibration, internal and recovery standards supplied by Wellington Laboratories.

Homogeneity checks were based on that described by Thompson and Fearn,<sup>12</sup> which is also the procedure as described in the International Harmonized Protocol.<sup>4</sup> The results are presented in Tables 61 to 76. Samples were found to be sufficiently homogeneous for use in a PT study with a target SD (as PCV) of 20%.

Container	Result	(µg/kg)		
Number	Replicate 1	Replicate 2		
5	0.85	0.84		
18	0.77	0.88		
29	0.84	0.80		
40	0.81	0.92		
52	0.85	0.83		
66	0.81	0.79		
75	0.90	0.80		
Mean	0.83			
CV	5.2%			

## Table 61 Sample S2 PFBS Homogeneity Testing

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.331	0.727	Pass
$s_{an}/\sigma$	0.309	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	0.009	Pass

## Table 62 Sample S2 PFPeS Homogeneity Testing

Container	Result	(µg/kg)		
Number	Replicate 1	Replicate 2		
5	8.5	8.2		
18	8.0	8.5		
29	8.9	7.7		
40	7.4	8.5		
52	8.3	8.5		
66	8.4	8.3		
75	9.5	7.7		
Mean	8.3			
CV	6.4	1%		

## Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.519	0.727	Pass
$s_{an}/\sigma$	0.404	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	1.171	Pass

## Table 63 Sample S2 PFHxS Homogeneity Testing

Container	Result (µg/kg)					
Number	Replicate 1	Replicate 2				
5	5.8	6.0				
18	6.1	6.4				
29	6.3	6.1				
40*	5.1	6.3				
52	5.6	6.2				
66	5.8	5.8				
75	5.7	5.4				
Mean	5.9					
CV	6.3%					

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.681	0.781	Pass
s <sub>an</sub> /σ	0.169	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.051	0.349	Pass

\* Results from container 40 were not included in the test for homogeneity, being identified as Cochran outliers due to the difference between replicates.<sup>12</sup>

Container	Result	(µg/kg)		
Number	Replicate 1	Replicate 2		
5	2.58	2.52		
18	2.69	2.43		
29	2.67	2.77		
40	2.67	3.02		
52	2.31	2.69		
66	2.67	2.52		
75	2.87	2.26		
Mean	2.62			
CV	7.8	3%		

# Table 64 Sample S2 PFHpS Homogeneity Testing

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.498	0.727	Pass
$s_{an}/\sigma$	0.442	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	0.129	Pass

# Table 65 Sample S2 PFOS Homogeneity Testing

Container	Result (µg/kg)		
Number	Replicate 1	Replicate 2	
5	1.86	1.89	
18	2.05	1.94	
29	1.99	1.67	
40	1.98	1.76	
52	1.77	1.83	
66	1.96	1.87	
75	1.91	1.80	
Mean	1.88		
CV	5.6%		

## Thompson and Fearn Homogeneity ${\rm Tests}^{12}$

Test	Value	Critical	Result
Cochran	0.542	0.727	Pass
$s_{an}/\sigma$	0.308	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	0.046	Pass

# Table 66 Sample S2 PFNS Homogeneity Testing

Container	Result (µg/kg)	
Number	Replicate 1	Replicate 2
5	1.62	1.51
18	1.53	1.49
29	1.64	1.44
40	1.54	1.42
52	1.56	1.67
66	1.56	1.12
75	1.51	1.53
Mean	1.51	
CV	8.8%	

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.716	0.727	Pass
s <sub>an</sub> /σ	0.467	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	0.046	Pass

Container	Result (µg/kg)		
Number	Replicate 1	Replicate 2	
5	6.3	6.4	
18	6.6	6.8	
29	6.9	5.9	
40	6.5	7.0	
52	6.0	5.7	
66	6.6	5.9	
75	6.3	5.9	
Mean	6.4		
CV	6.4%		

# Table 67 Sample S2 PFDS Homogeneity Testing

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.493	0.727	Pass
s <sub>an</sub> /σ	0.311	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.011	0.528	Pass

# Table 68 Sample S2 PFPeA Homogeneity Testing

Container	Result (µg/kg)		
Number	Replicate 1	Replicate 2	
5	1.84	1.96	
18	1.98	1.80	
29	1.97	1.93	
40	2.05	1.88	
52	1.99	1.87	
66	1.80	1.96	
75	2.04	2.00	
Mean	1.93		
CV	4.3%		

## Thompson and Fearn Homogeneity ${\rm Tests}^{12}$

Test	Value	Critical	Result
Cochran	0.285	0.727	Pass
$s_{an}/\sigma$	0.243	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	0.041	Pass

## Table 69 Sample S2 PFHxA Homogeneity Testing

Container	Result (µg/kg)		
Number	Replicate 1	Replicate 2	
5	7.3	7.7	
18	7.3	8.2	
29	7.0	6.5	
40	7.3	6.9	
52	7.4	7.7	
66	7.0	6.8	
75	7.9	7.4	
Mean	7.3		
CV	6.3%		

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.411	0.727	Pass
s <sub>an</sub> /σ	0.249	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.090	0.596	Pass

Container	Result (µg/kg)		
Number	Replicate 1	Replicate 2	
5	1.58	1.47	
18	1.64	1.40	
29	1.52	1.46	
40	1.52	1.69	
52	1.58	1.46	
66	1.46	1.53	
75	1.40	1.55	
Mean	1.52		
CV	5.5%		

#### Table 70 Sample S2 PFHpA Homogeneity Testing

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.397	0.727	Pass
$s_{an}/\sigma$	0.331	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	0.032	Pass

#### Table 71 Sample S2 PFOA Homogeneity Testing

Container	Result (µg/kg)	
Number	Replicate 1	Replicate 2
5	1.17	1.19
18	1.22	1.19
29	1.22	1.27
40*	1.18	1.06
52	1.18	1.19
66	1.24	1.25
75	1.23	1.21
Mean	1.20	
CV	4.1%	

Test	Value	Critical	Result
Cochran	0.649	0.781	Pass
$s_{an}/\sigma$	0.078	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.001	0.012	Pass

\* Results from container 40 were not included in the test for homogeneity, being identified as Cochran outliers due to the difference between replicates.<sup>12</sup>

#### Table 72 Sample S2 PFNA Homogeneity Testing

Container	Result (µg/kg)	
Number	Replicate 1	Replicate 2
5	2.32	2.25
18	2.17	2.23
29	2.22	2.02
40	2.25	2.11
52	2.35	2.25
66	2.24	2.17
75	2.35	2.20
Mean	2.22	
CV	4.0%	

Thompson and Fearn Homogeneity Tests<sup>12</sup>

-		e	
Test	Value	Critical	Result
Cochran	0.380	0.727	Pass
$s_{an}/\sigma$	0.193	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.001	0.048	Pass

Container	Result (µg/kg)		
Number	Replicate 1	Replicate 2	
5	9.5	10.1	
18	9.4	9.5	
29	9.1	9.6	
40	10.2	9.5	
52	9.0	9.3	
66	9.8	8.4	
75	9.1	8.8	
Mean	9.4		
CV	5.3%		

#### Table 73 Sample S2 PFDA Homogeneity Testing

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.615	0.727	Pass
$s_{an}/\sigma$	0.263	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.002	1.015	Pass

#### Table 74 Sample S2 6:2 FTS Homogeneity Testing

Container	Result (µg/kg)		
Number	Replicate 1	Replicate 2	
5	1.71	1.69	
18	1.95	1.76	
29	1.85	2.26	
40	2.08	2.17	
52	1.48	2.00	
66	1.74	1.78	
75	2.12	2.12	
Mean	1.91		
CV	12%		

#### Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.562	0.727	Pass
$s_{an}/\sigma$	0.488	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.017	0.077	Pass

#### Table 75 Sample S2 GenX Homogeneity Testing

Container	Result (µg/kg)	
Number	Replicate 1	Replicate 2
5	10.2	10.7
18	10.1	10.0
29	10.4	10.2
40	10.3	10.4
52	10.8	10.3
66	10.3	10.0
75	10.5	10.4
Mean	10.3	
CV	2.3%	

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.393	0.727	Pass
$s_{an}/\sigma$	0.109	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.005	0.881	Pass

Container	Result (µg/kg)				
Number	Replicate 1	Replicate 2			
5*	24.8	17.1			
18	15.8	13.3			
29	15.8	14.8			
40	15.9	16.4			
52	14.7	15.7			
66	15.8	14.7			
75	15.7	15.1			
Mean	16.1				
CV	17	%			

#### Table 76 Sample S2 ADONA Homogeneity Testing

Thompson and Fearn Homogeneity Tests<sup>12</sup>

Test	Value	Critical	Result
Cochran	0.603	0.781	Pass
$s_{an}/\sigma$	0.302	0.500	Pass
s <sup>2</sup> <sub>sam</sub>	0.000	3.318	Pass

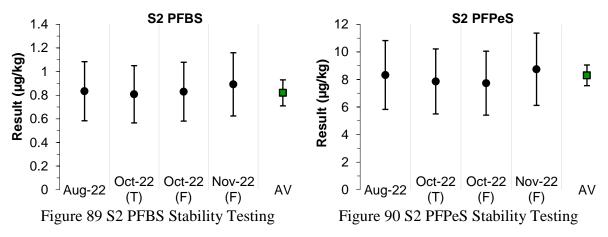
\* Results from container 5 were not included in the test for homogeneity, being identified as Cochran outliers due to the difference between replicates.<sup>12</sup>

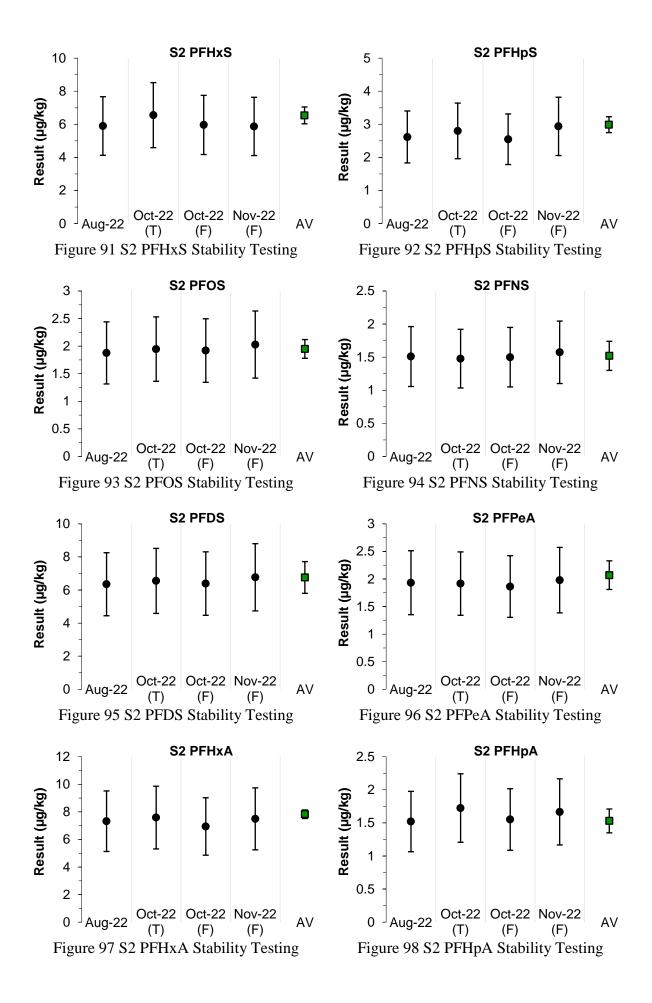
#### **Stability Testing**

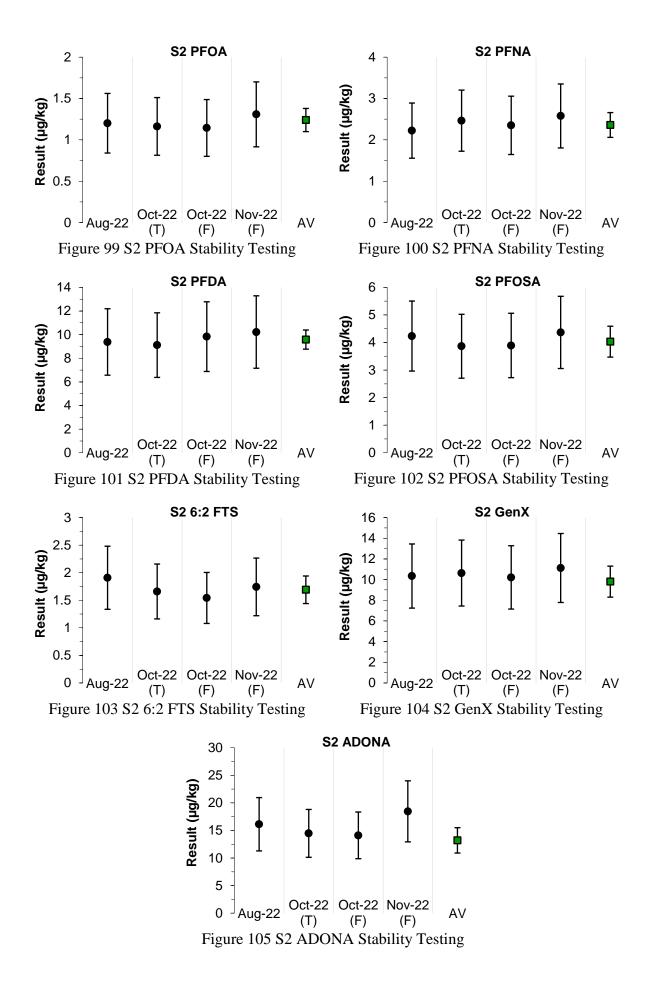
Sample S2 carrot analysis was performed as described above.

The carrot samples were analysed at an initial time point in August 2022 (the start of the PT study). On the sample dispatch date, a sample was set aside and packaged in the same way as the samples dispatched to participants. This was stored at ambient conditions until all samples had been delivered to the participants (October 2022), before being analysed, to reflect transportation stability. Additional samples were stored at freezer temperature, to reflect long-term storage temperature at a participant's laboratory; samples were taken for analysis at October 2022 and November 2022 (the conclusion of the PT study).

Results were in good agreement with each other and the assigned value within their respective uncertainties (Figures 89 to 105, T = Transportation Stability and F = Freezer Stability). The samples were also shown to be adequately stable when assessed against the criteria specified in ISO 13528:2022.<sup>6</sup>

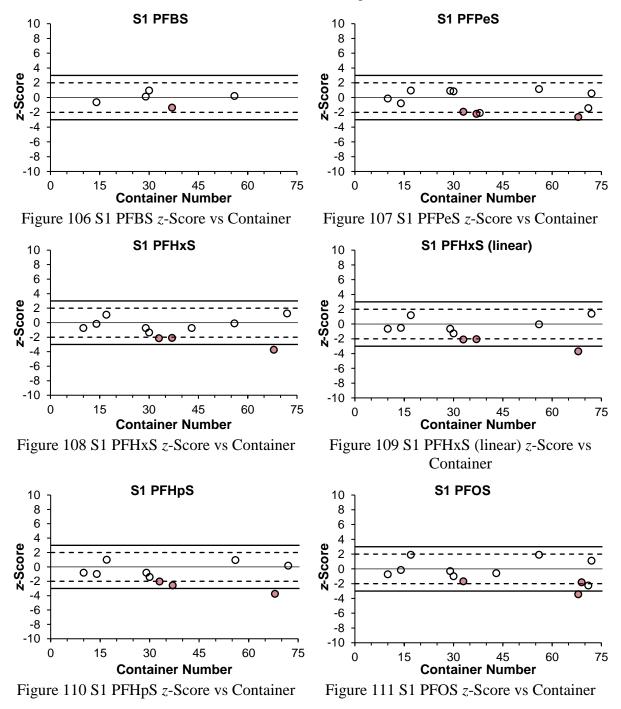


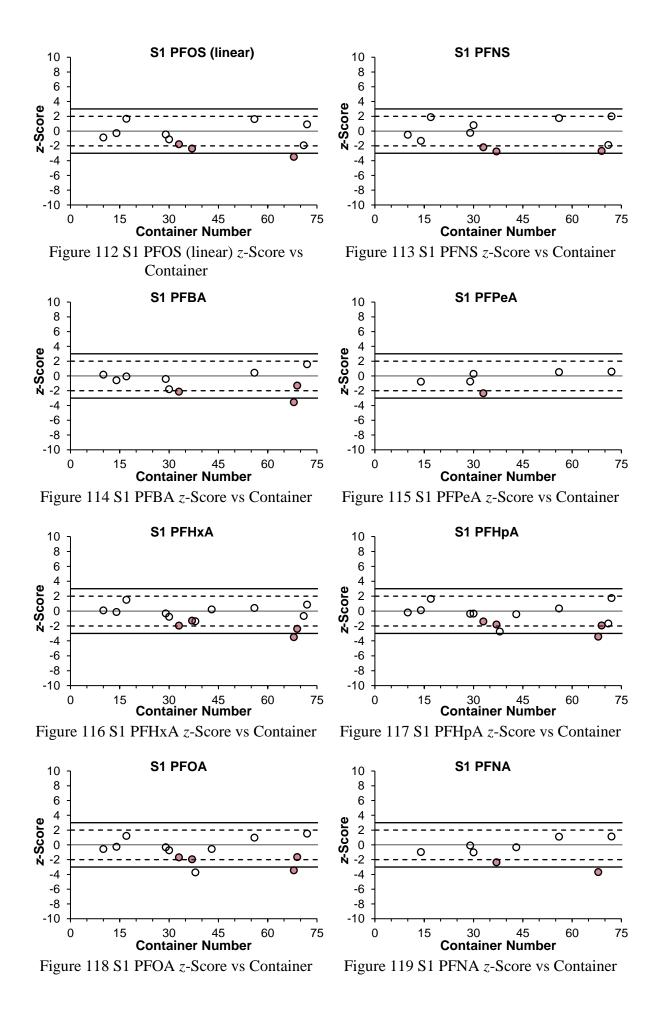


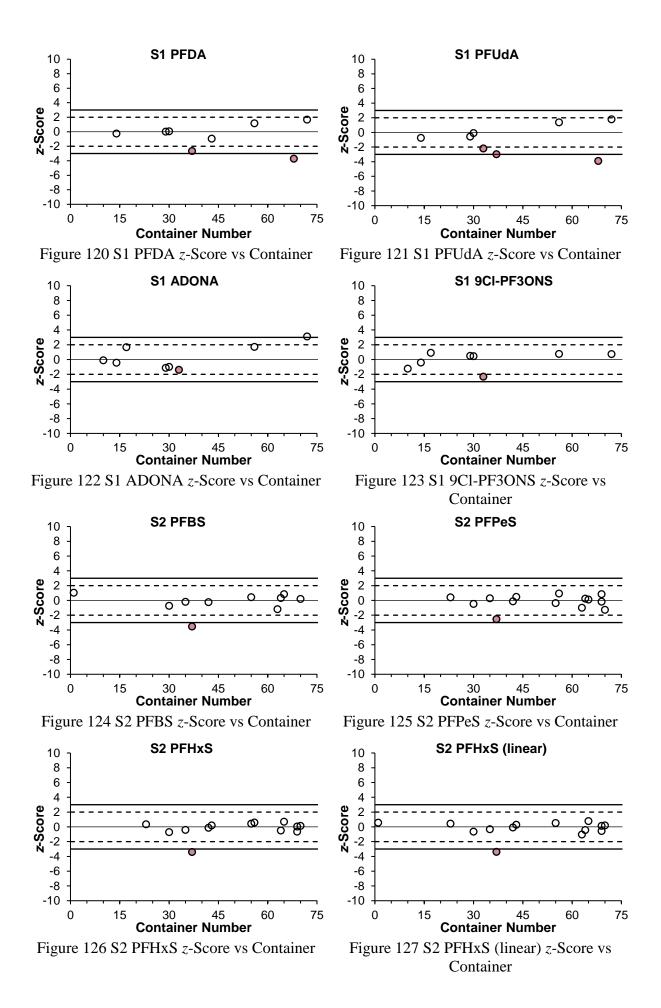


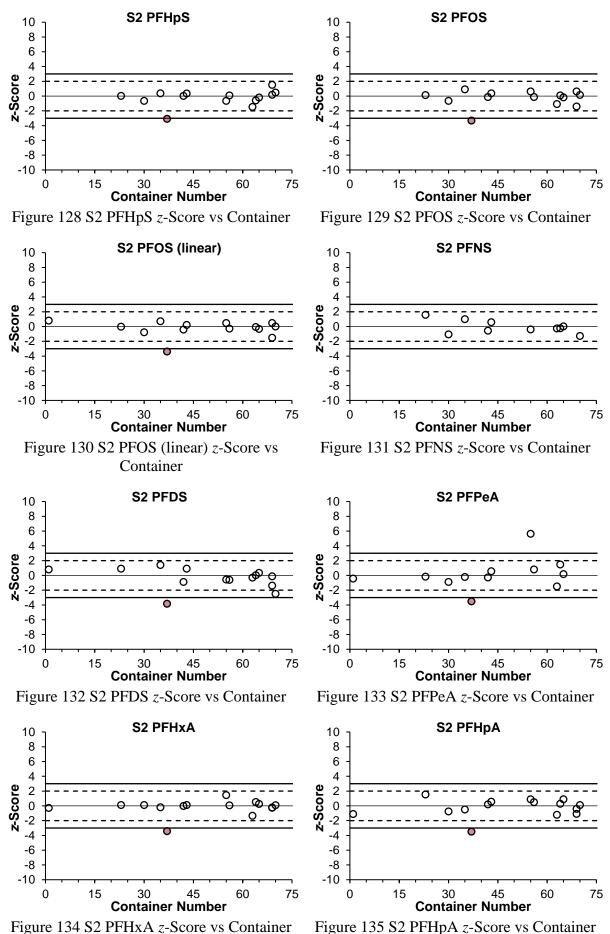
#### A2.2 Comparison of Results and Container Numbers

Comparisons of *z*-scores obtained to the container number analysed by participants for all scored analytes are presented for information in Figures 106 to 144 (results have been included when the participant was sent one sample set only), with results excluded from statistical calculations as described in Section 4.2 being shaded.

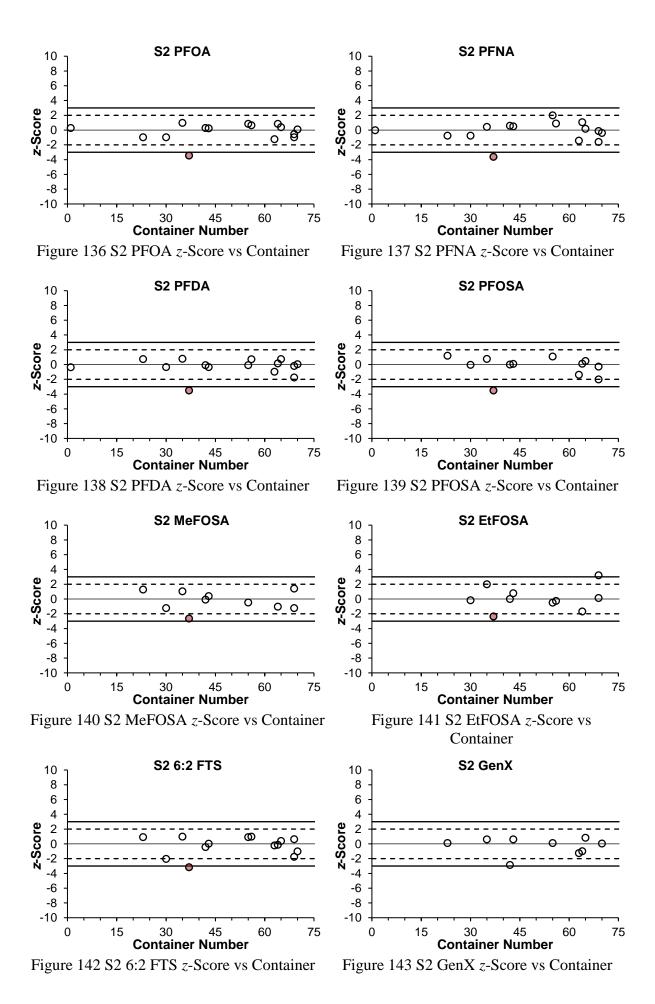


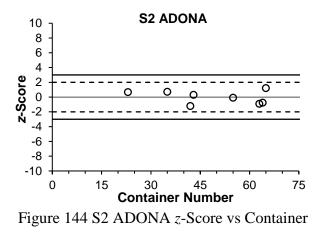






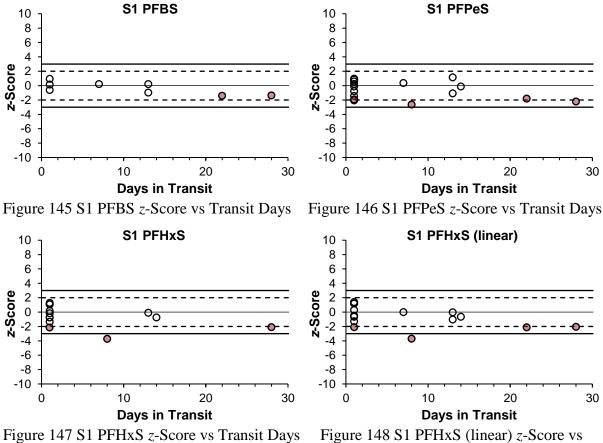
148



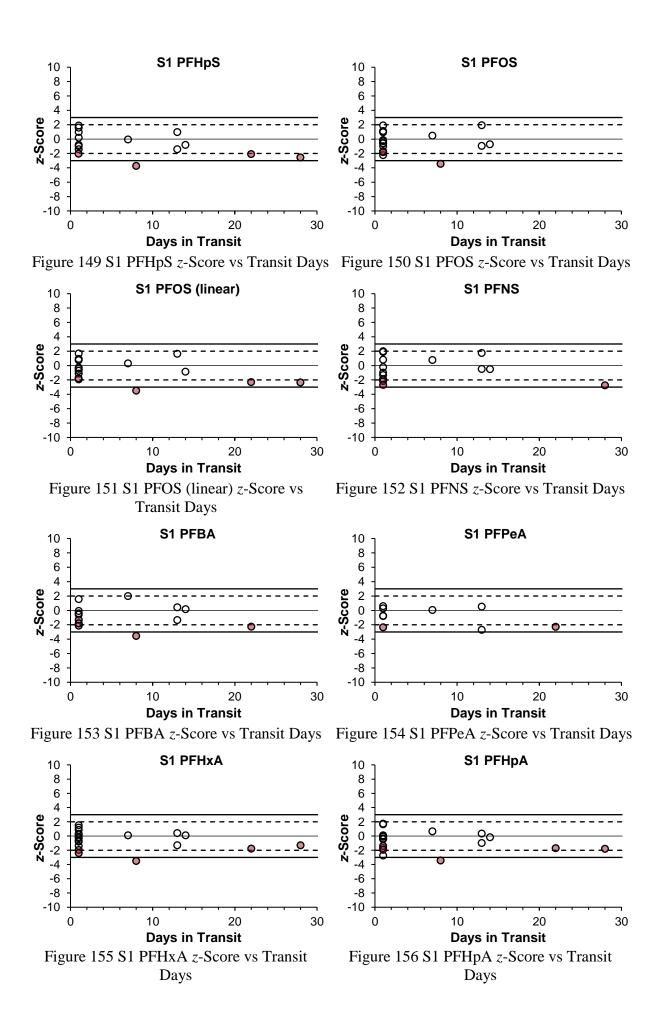


A2.3 Comparison of Results and Days in Transit

Comparisons of participants' results to the number of days the samples spent in transit for all scored analytes are presented for information in Figures 145 to 183, with results excluded from statistical calculations as described in Section 4.2 being shaded.



Transit Days



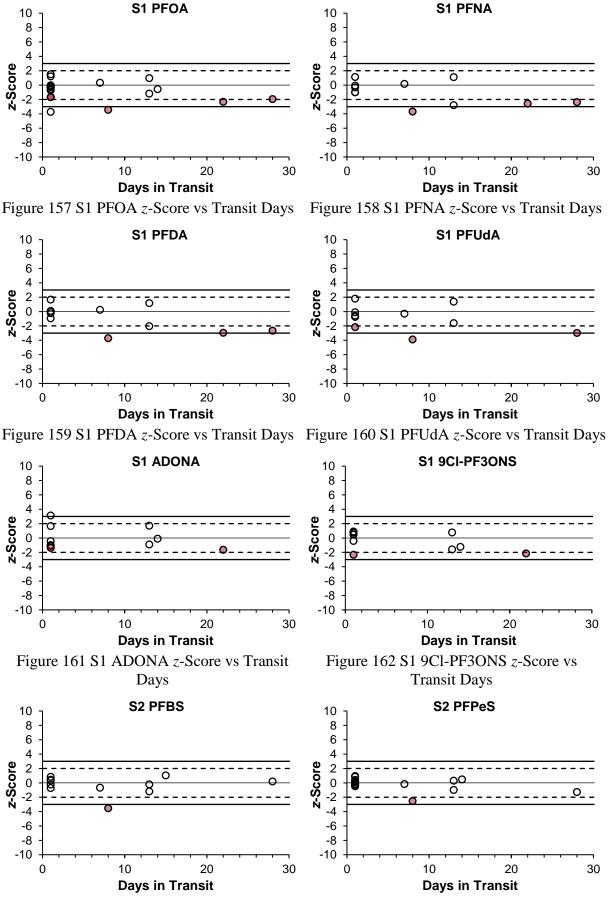


Figure 163 S2 PFBS *z*-Score vs Transit Days

Figure 164 S2 PFPeS z-Score vs Transit Days

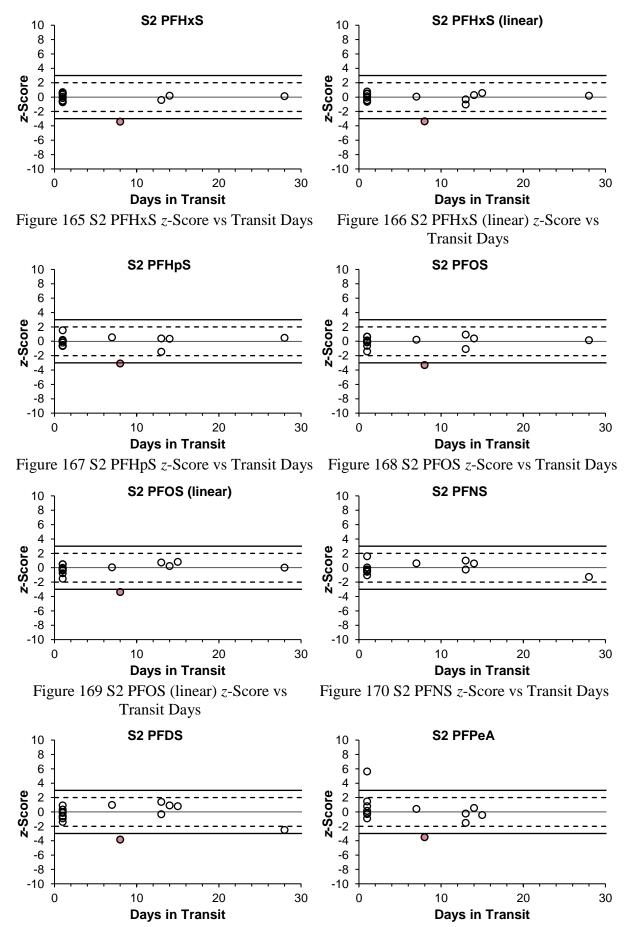
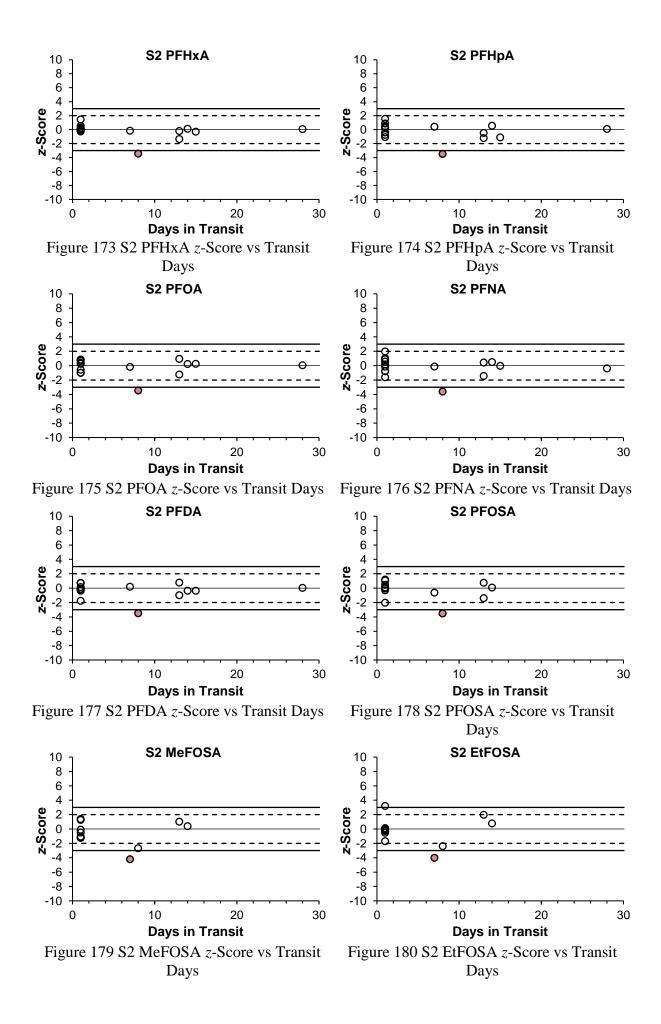
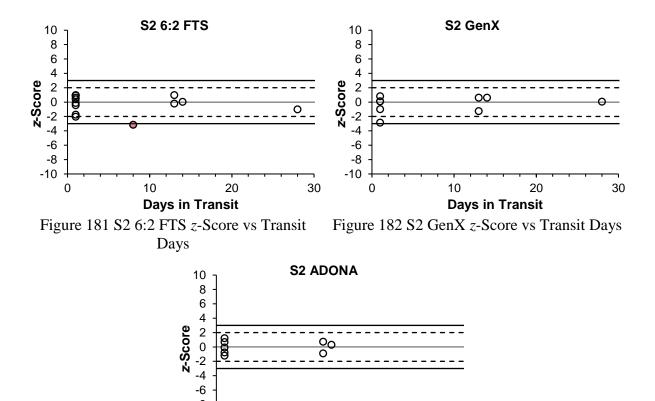


Figure 171 S2 PFDS z-Score vs Transit Days Figure 172 S2 PFPeA z-Score vs Transit Days





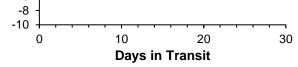


Figure 183 S2 ADONA z-Score vs Transit Days

## APPENDIX 3 ROBUST AVERAGE AND ASSOCIATED UNCERTAINTY, z-SCORE AND $E_{rr}$ -SCORE CALCULATIONS

#### A3.1 Robust Average and Associated Uncertainty

Robust averages were calculated using the procedure described in ISO 13528:2022.<sup>6</sup> The associated uncertainties were estimated as according to Equation 4.

$$u_{rob\ av} = \frac{1.25 \times S_{rob\ av}}{\sqrt{p}} \qquad Equation\ 4$$

where:

<i>Urob av</i>	is the standard uncertainty of the robust average
Srob av	is the standard deviation of the robust average
р	is the number of results

The expanded uncertainty  $(U_{rob av})$  is the standard uncertainty multiplied by a coverage factor of 2 at approximately 95% confidence level.

A worked example is set out below in Table 77.

Table 77 Uncertainty Estimate for Robust Average of Sample S2 PFHpS

Number of Results (p)	14
Robust Average	2.99 µg/kg
$S_{rob av}$	0.36 µg/kg
$u_{rob av}$	0.12 µg/kg
k	2
Urob av	0.24 µg/kg

Therefore, the robust average for Sample S2 PFHpS is  $2.99 \pm 0.24 \ \mu g/kg$ .

#### A3.2 z-Score and En-Score Calculations

For each participant's result, a *z*-score and  $E_n$ -score are calculated according to Equations 2 and 3 respectively (Section 0).

A worked example is set out below in Table 78.

Table 78 z-Score and En-Score for Sample S1 PFBS Result Reported by Laboratory 2

Participant Result (µg/kg)	Assigned Value (µg/kg)	Target Standard Deviation	z-Score	<i>E</i> <sub>n</sub> -Score
$0.244 \pm 0.011$	$0.302 \pm 0.048$	20% as PCV, or: 0.2 × 0.302 = 0.0604 μg/kg	$z\text{-Score} = \frac{0.244 - 0.302}{0.0604}$ $= -0.96$	$E_n\text{-Score} = \frac{0.244 - 0.302}{\sqrt{0.011^2 + 0.048^2}}$ $= -1.18$

#### APPENDIX 4 PARTICIPANTS' TEST METHODS

Participants' responses to the methodology questionnaire are presented in Tables 79 to 129. Some responses may have been modified so that the participant cannot be identified.

Lab. Code	S1 Sample Weight (g)	Sample Pretreatment	Extraction Technique	Extraction Solvent	Extraction Temperature	Extraction Time	Clean-Up
1	1	Homogenisation Geno/Grinder 14min & Centrifuge 10min	QuEChERS - modified AOAC	Acetonitrile with 1% Acetic Acid	Room	Sonicate 30 min at 30-35 degrees	envicarb
2							
3							
4	1	Homogenisation	Alkaline Digestion	KOH-methanol	Room temp	8 hrs	Active carbon SPE
5				NS			
6			Solid-Liquid Extraction				
7							
8	6.1	Homogenisation	QuEChERS	Acetonitrile/Sodium Hydroxide	Room temperature	180mins	Solid-Phase Extraction
9	1		Alkaline Digestion	Acetonitrile	Room temperature	2 x 20 minutes	GCB and WAX SPE
10	2		Solid-Liquid Extraction	Acidified acetonitrile/water	Room		
11	1	No	Solid-Liquid Extraction	Acetonitrile	Room temperature	30 min	SPE-WAX, ultracentrifugation
12	2.01 (as received)	Homogenisation	Solid-Liquid Extraction	KOH-Methanol	Ambient Room Temperature	16 hours	Activated carbon/SPE/filtration

Table 79 Participant Methodology – Sample S1 Prawn Extraction

Lab. Code	S1 Sample Weight (g)	Sample Pretreatment	Extraction Technique	Extraction Solvent	Extraction Temperature	Extraction Time	Clean-Up
13	1	Homogenisation	Digestion with 200mM NaOH in methanol, then extraction with acetonitrile.	Acetonitrile	Room Temperature	2 x 15min	liquid-liquid extraction with n- hexane, then Bond Elut Carbon SPE
15	1g	Homogenisation	Alkaline Digestion	Basified MeOH	Room	60 mins	Envicarb
16	1.1	Homogenisation	QuEChERS	Methanol + Ammonium- ACN and Acetone	40 °C	30 min	Solid-Phase Extraction
17	1	Homogenisation	Solid-Liquid Extraction	Acetonitrile	Ambient	30 mins	Solid-Phase Extraction
18	0.5 grams	Homogenisation	Solid-Liquid Extraction	MTbE	Room	60 minutes	C18 & Activated Carbon
19	2.076 and 2.106 (duplicate)	NA	Solid-Liquid Extraction Merris-Minimix shaker	2% formic acid in acetonitrile	Room temperature	8 min	dSPE (C18, Envicarb, MgSO4)
20	0.7	Homogenisation	Solid-Liquid Extraction	0.1 % NH3 in ACN	room temperature	2 h	Solid-Phase Extraction two different SPE cartridges
21			Alkaline Digestion				

## Table 80 Participant Methodology – Sample S1 Prawn Instrumental Technique and Analysis

Lab. Code	Instrument	Guard Column	Instrument Column	Dilution Factor	Delay Column?	Blank Correction?	Standard Method?
1	Orbitrap	C18 3mm	Kinetex C18 100x3mm 2.6 um		Yes	Yes	In house
2							US FDA Foods Program Compendium of Analytical Laboratory Methods; method C-010.02
3							
4	LC-MSMS or LC-QQQ	No	C18, 50 mm	No	Yes	No	No
5	NS						

Lab. Code	Instrument	Guard Column	Instrument Column	Dilution Factor	Delay Column?	Blank Correction?	Standard Method?
6	LC-MSMS or LC-QQQ	C18	2.1x100mn 1.9um		Yes	No	USEPA 537
7							
8	LC-MSMS or LC-QQQ	None	InfinityLab Poroshell HPH-C18 column, 2.1x50mm, 2.7micron	0.16393	No	No	
9	LC-MSMS or LC-QQQ	Nil	Shimadzu Shim-pack XR-ODS III (1.6 μm, 50 mm x 2.0 mm)	No	Yes	No	No
10	LC-MSMS or LC-QQQ				Yes		
11	LC-MSMS or LC-QQQ	Gemini NX-C18; 4mm x 2.0 mm ID	NX-C18; 15cm x 2mm x 3um	No	Yes	Yes	No
12	LC-MSMS or LC-QQQ	Phenomenex Evo C18 (2µm, 2 mm x 2.1 mm)	BEH C18 (1.7μm, 50 mm x 2.1 mm)	No	Yes	No	No
13	LC-MSMS or LC-QQQ	Phenomenex Evo C18 (2 mm x 2.1 mm)	Phenomenex Evo C18 (100 mm x 2.1 mm x 2.6 um)	No	Yes	No	Isotopic Dilution
15	LC-MSMS or LC-QQQ	Pre-column Filter 0.2µm	C18 50mm x 2.1mm x 1.8µm	50	Yes	No	No. In-house
16	LC-MSMS or LC-QQQ	UltraShield UHPLC 0.2 µm Restek	Raptor C18 1.8 µm 50 x 2.1 mm Restek		yes	no	
17	Orbitrap	C18	C18		Yes		
18	LC-MSMS or LC-QQQ	nil	C18 1.6µm, 2.0mm x 50mm	No	Yes	No	
19	LC-MSMS or LC-QQQ	NA	Zorbax XDB-C18, 100 mm x 2.1 mm, 1.8μm	NA	Yes	No	No
20	LC-MSMS or LC-QQQ	C18 Column, 2.1 x 5 mm, 3.5 µm	C18 Column, 130Å, 3 x 50 mm, 3.5 µm		Yes	No	
21	LC-MSMS or LC-QQQ	C18	2.1x100mn 1.9um		Yes	No	USEPA 537

Lab. Code	Labelled Standard Source	Recovery Correction?	Labelled Standards Additional Information
1	Wellington	No	
2	Wellington	Yes	d5NN EtFOSAA added before instrument analysis
3			
4	Wellington	Yes	
5			NS
6	Wellington	no	
7			
8	Wellington	Yes	
9	Wellington	Yes	
10			
11	Wellington	Yes	
12	Wellington	Yes	
13	Wellington	Yes	
15	Wellington	Yes	
16		Yes	
17	Wellington	Yes	Results corrected by ISTD added before instrumentation
18	Wellington Laboratories	No	
19	Wellington Laboratory	Yes	
20	Wellington	No	
21	Wellington	no	

## Table 81 Participant Methodology – Sample S1 Prawn Labelled Standards

#### Table 82 Labelled Standards for S1 PFBS

Lab. Code	Before Extraction	Before Instrument Analysis
1	Sodium perfluoro-1- [2,3,4 13C3] butanesulfonate M3PFBS	
2	M3 PFBS	
3		
4	13C3 PFBS	
5	N	S
6		M3PFBS
7		
8	13C3-PFBS	
9	13C3-PFBS	13C8-PFOS
10		
11	18O2-PFHxS	18O2-PFOS
12	13C3-PFBS	18O2-PFHxS
13	13C3-PFBS	13C3-PFHxS
15	13C3-PFBS	N/A
16	13C3-PFBS	13C4-PFOA
17	PFOS-C8	PFBS-13C3
18		13C3-PFBS
19	M3PFBS	NA
20	13C3-PFBS	13C2-PFHxA
21		M3PFBS

-	
Before Extraction	Before Instrument Analysis
M PFHxS	
13C3 PFBS	
N	S
13C3-PFBS	
Ν	Т
18O2-PFHxS	18O2-PFOS
13C3-PFHxS	18O2-PFHxS
18O2-PFHxS	13C3-PFHxS
18O2-PFHxS	N/A
13C3-PFBS	13C4-PFOA
PFOS-C8	PFOS-C4
	16O2-PFHxS
M5PFHxA	NA
linear 13C3-PFHxS	13C2-PFHxA
	M PFHxS 13C3 PFBS 13C3 PFBS 13C3-PFBS 13C3-PFHxS 18O2-PFHxS 18O2-PFHxS 18O2-PFHxS 18O2-PFHxS 13C3-PFBS PFOS-C8 M5PFHxA

Table 83 Labelled Standards for S1 PFPeS

#### Table 84 Labelled Standards for S1 PFHxS

Lab. Code	Before Extraction	Before Instrument Analysis
1	Sodium perfluoro- 1-[1,2,3 13C3] hexanesulfonate M3PFHxS	
2		
3	N	Т
4	13C3 PFHxS	PFHxS18O2
5	N	IS
6		
7		
8	18O2-PFHXS	
9	18O2-PFHxS	13C8-PFOS
10		
11	N	Т
12	13C3-PFHxS	18O2-PFHxS
13	18O2-PFHxS	13C3-PFHxS
15	18O2-PFHxS	N/A
16	18O2-PFHxS	13C4-PFOA
17	PFOS-C8	PFHxS-18O2
18		16O2-PFHxS
19	M3PFHxS	NA
20	linear 13C3-PFHxS	
21		

## Table 85 Labelled Standards for S1 PFHxS (linear)

	· · · · ·		
Lab. Code	Before Extraction	Before Instrument Analysis	
1			
2	M PFHxS		
3			
4	13C3 PFHxS		
5	N	IS	
6		M3PFHxS	
7			
8	18O2-PFHXS		
9	N	T	
10			
11	18O2-PFHxS	18O2-PFOS	
12	13C3-PFHxS	18O2-PFHxS	
13	18O2-PFHxS	13C3-PFHxS	
15	18O2-PFHxS	N/A	
16	18O2-PFHxS	13C4-PFOA	
17	PFOS-C8	PFHxS-18O2	
18	N	NT	
19	M3PFHxS	NA	
20	linear 13C3-PFHxS		
21		M3PFHxS	

Table 86 Labelled Standards for S1 PFHpS

Before Instrument Lab. Before Extraction Code Analysis 1 2 M PFHxS 3 13C3 PFHxS 4 NS 5 6 7 8 18O2-PFHXS 9 NT 10 18O2-PFHxS 1802-PFOS 11 13C8-PFOS 13C4-PFOS 12 13 18O2-PFHxS 13C3-PFHxS 15 13C4-PFOS N/A 18O2-PFHxS 13C4-PFOA 16 PFOS-C8 PFOS-C4 17 18 13C8-PFOS 19 M3PFHxS NA 20 linear 13C3-PFHxS 21

Table 87 Labelled Standards for S1 PFOS

Lab. Code	Before Extraction	Before Instrument Analysis
1	Sodium perfluoro- 1-[ 13C8] ocatanesulfonate M8PFOS	
2	13C PFOS	
3	N	Т
4	13C8 PFOS	13C4 PFOS
5	N	IS
6		
7		
8	13C8-PFOS	
9	13C4-PFOS	13C8-PFOS
10		
11	13C4-PFOS	18O2-PFOS
12	13C8-PFOS	13C4-PFOS
13	13C4-PFOS	13C8-PFOS
15	13C4-PFOS	N/A
16	13C4-PFOS	13C4-PFOA
17	PFOS-C8	PFOS-C4
18	13C4-PFOS	13C8-PFOS
19	M8PFOS	NA
20	linear 13C8-PFOS	13C5-PFNA
21		

## Table 88 Labelled Standards for S1 PFOS (linear)

	()	
Lab. Code	Before Extraction	Before Instrument Analysis
1		
2		
3		
4	13C8 PFOS	
5	Ν	IS
6	MPFOS	M8PFOS
7		
8	13C8-PFOS	
9	N	νT
10		
11	13C4-PFOS	18O2-PFOS
12	13C8-PFOS	13C4-PFOS
13	13C4-PFOS	13C8-PFOS
15	13C4-PFOS	N/A
16	13C4-PFOS	13C4-PFOA
17	PFOS-C8	PFOS-C4
18		13C8-PFOS
19	M8PFOS	NA
20	linear 13C8-PFOS	
21	MPFOS	M8PFOS

Table 89 Labelled Standards for S1 PFNS Before Instrument Lab. Before Extraction Code Analysis 1 2 13C PFOS 3 NT 13C8 PFOS 4 NS 5 6 7 8 13C8-PFOS 9 NT

> 13C4-PFOS 13C8-PFOS

> 13C4-PFOS

13C4-PFOS

PFOS-C8

M8PFOS

linear 13C8-PFOS

1802-PFOS

13C4-PFOS

13C8-PFOS

N/A

PFBS-13C3

NA

NT

NT

10

11

12 13

15

16

17

18

19

20

21

Table 90 Labelled Standards for S1 PFBA

Lab.	Before Extraction	Before Instrument
Code	Defore Extraction	Analysis
1	Perfluoro-n- [13C4]butanoic acid MPFBA	
2	M3 PFBA	
3		
4	13C4 PFBA	13C3 PFBA
5	N	IS
6	M3PFBA	MPFBA
7		
8	13C4-PFBA	
9	NT	
10		
11	13C4-PFBA	13C8-PFOA
12	13C4-PFBA	13C3-PFBA
13	13C4-PFBA	13C3-PFBA
15	13C4-PFBA	N/A
16	13C4-PFBA	13C4-PFOA
17	PFOS-C8	PFBA-13C4
18		13C4-PFBA
19	M4PFBA	NA
20	N	Т
21	M3PFBA	MPFBA

### Table 91 Labelled Standards for S1 PFPeA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [13C5]pentanoic acid M5PFPeA	
2	M3 PFPeA	
3		
4	13C5 PFPeA	
5	N	IS
6		M5PFPeA
7		
8	13C5-PFPEA	
9	N	T
10		
11	13C5-PFPeA	13C8-PFOA
12	13C5-PFPeA	13C2-PFHxA
13	13C4-PFPeA	13C5 -PFPeA
15	13C3-PFPeA	N/A
16	13C5-PFPeA	13C4-PFOA
17	PFOS-C8	PFPeA-13C3
18		13C5-PFPeA
19	M5PFPeA	NA
20	NT	
21		M5PFPeA

Table 92 Labelled Standards for S1 PFHxA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [1,2,3,4,6- 13C5]hexanoic acid M5PFHxA	
2	M3 PFHxA	
3		
4	13C5 PFHxA	13C2 PFHxA
5	N	S
6		M5PFHxA
7		
8	13C2-PFHXA	
9	13C2-PFHxA	13C8-PFOA
10		
11	13C5-PFHxA	13C8-PFOA
12	13C5-PFHxA	13C2-PFHxA
13	13C2-PFHxA	13C5 -PFPeA
15	13C2-PFHxA	N/A
16	13C2-PFHxA	13C4-PFOA
17	PFOS-C8	PFHxA-13C2
18		13C5-PFHxA
19	M5PFHxA	NA
20	13C5-PFHxA	13C2-PFHxA
21		M5PFHxA

Table 93 Labelled Standards for S1 PFHpA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [1,2,3,4- 13C4]heptanoic acid M4PFHpA	
2	M3 PFHxA	
3		
4	13C4 PFHpA	
5	Ν	IS
6		M4PFHpA
7		
8	13C4-PFHPA	
9	13C4-PFHpA	13C8-PFOA
10		
11	13C4-PFHpA	13C8-PFOA
12	13C4-PFHpA	13C4-PFOA
13	13C3-PFHpA	13C8-PFOA
15	13C4-PFHpA	N/A
16	13C4-PFHpA	13C4-PFOA
17	PFOS-C8	PFHpA-13C4
18		13C4-PFHpA
19	MPFHpA	NA
20	13C4-PFHpA	13C2-PFHxA
21		M4PFHpA

#### Table 94 Labelled Standards for S1 PFOA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [13C8]octanoic acid M8PFOA	
2	13C PFOA	
3		
4	13C8 PFOA	13C4 PFOA
5	N	IS
6	M2PFOA	M8PFOA
7		
8	13C8-PFOA	
9	13C4-PFOA	13C8-PFOA
10		
11	13C4-PFOA	13C8-PFOA
12	13C8-PFOA	13C4-PFOA
13	13C4-PFOA	13C8-PFOA
15	13C4-PFOA	N/A
16	13C8-PFOA	13C4-PFOA
17	PFOS-C8	PFOA-13C4
18	13C8-PFOA	13C4-PFOA
19	M8PFOA	NA
20	13C8-PFOA	13C5-PFNA
21	M2PFOA	M8PFOA

Before Extraction	Before Instrument Analysis
Perfluoro-n- [13C9]nonanoic acid M9PFNA	
13C PFOA	
13C9 PFNA	13C5 PFNA
N	S
	M9PFNA
13C5-PFNA	
13C5-PFNA	13C8-PFOA
13C9-PFNA	13C5-PFNA
13C9-PFNA	13C5-PFNA
13C5-PFNA	13C8-PFOA
13C5-PFNA	N/A
13C5-PFNA	13C4-PFOA
PFOS-C8	PFNA-13C5
	13C5-PFNA
M9PFNA	NA
13C9-PFNA	13C5-PFNA
	M9PFNA
	Perfluoro-n- [13C9]nonanoic acid M9PFNA 13C PFOA 13C9 PFNA 13C9 PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA 13C5-PFNA

Table 95 Labelled Standards for S1 PFNA

#### Table 96 Labelled Standards for S1 PFDA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [1,2,3,4,6- 13C6]decanoic acid M6PFDA	
2	13C PFOA	
3		
4	13C6 PFDA	13C2 PFDA
5	N	IS
6	MPFDA	M6PFDA
7		
8	13C6-PFDA	
9	13C2-PFDA	13C8-PFOA
10		
11	13C2-PFDA	13C5-PFNA
12	13C6-PFDA	13C2-PFDA
13	13C2-PFDA	13C8-PFOA
15	13C2-PFDA	N/A
16	13C2-PFDA	13C4-PFOA
17	PFOS-C8	PFDA-13C2
18		13C6-PFDA
19	M6PFDA	NA
20	13C6-PFDA	13C5-PFNA
21	MPFDA	M6PFDA

#### Table 97 Labelled Standards for S1 PFUdA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [1,2,3,4,6,7- 13C7]undecanoic acid M7PFUdA	
2	MPFUdA	
3	N	Τ
4	13C7 PFUnA	
5	N	IS
6		M7PFUdA
7		
8	13C2-PFUDA	
9	N	Τ
10		
11	13C2-PFUdA	13C5-PFNA
12	13C7-PFUnA	13C2-PFDA
13	13C2-PFUdA	13C8-PFOA
15	13C2-PFUdA	N/A
16	13C2-PFUnA	13C4-PFOA
17	PFOS-C8	PFUNDA-13C2
18		13C2-PFUnDA
19	M7PFUnDA	NA
20	13C7-PFUdA	13C5-PFNA
21		M7PFUdA

Lab. Code	Before Extraction	Before Instrument Analysis					
1							
2	MPFDoA						
3	NT						
4	13C2 PFDoA						
5	N	IS					
6							
7							
8	13C2-PFTEDA						
9	N	ΙΤ					
10							
11	13C2-PFHxDA	13C2-PFTeDA					
12	13C2-PFDoA; 13C2-PFTeDA	13C2-PFDA					
13	13C2-PFTeDA	13C8-PFOA					
15	13C2-PFTeDA	N/A					
16	13C2-PFTeDA	13C4-PFOA					
17	PFOS-C8	PFTeDA-13C2					
18		13C2-PFTeDA					
19	MPFDoDA	NA					
20	13C2-PFDoA						
21							

Table 98 Labelled Standards for S1 PFTrDA

#### Table 99 Labelled Standards for S1 PFOSA

Lab.	Before Extraction	Before Instrument		
Code	Defore Extraction	Analysis		
1	Perfluoro-1-[13C8] otanesulfonamide			
2	M8 FOSA			
3	N	Т		
4	13C8 PFOSA			
5	N	IS		
6		M8-FOSA		
7				
8	13C8-FOSA			
9	NT			
10				
11	13C8-PFOSA	13C2-PFTeDA		
12	13C8-PFOSA	13C4-PFOS		
13	13C8-FOSA			
15	13C8-FOSA	N/A		
16	13C8-PFOSA	13C4-PFOA		
17	PFOS-C8	FOSA-13C8		
18		13C8-FOSA		
19	MPFOSA	NA		
20	N	Т		
21		M8-FOSA		

#### Table 100 Labelled Standards for S1 MeFOSA

Lab. Code	Before Extraction	Before Instrument Analysis
1	N-methyl-d3- perfluoro-1- octancesulfonamide	
2	N	Т
3	N	T
4	d3-N-MeFOSA	
5	N	IS
6		d-N-MeFOSA
7		
8	d3-N-MEFOSA	
9	N	T
10		
11	13C8-PFOSA	13C2-PFTeDA
12	D3-N-MeFOSA	13C4-PFOS
13	D3-N-Me FOSA	
15	D3-M PFOSA	N/A
16	d3-N-MeFOSA	13C4-PFOA
17	PFOS-C8	MeFOSA-D3
18		d3-MeFOSA
19	d-NMeFOSA-M	NA
20	N	Т
21		d-N-MeFOSA

Lab. Code	Before Extraction	Before Instrument Analysis
1	N-ethyl-d5- perfluoro-1- octanesulfonamide	
2	N	T
3	N	T
4	d5-N-EtFOSA	
5	N	IS
6		d-N-EtFOSA
7		
8	d5-N-ETFOSA	
9	N	T
10		
11	13C8-PFOSA	13C2-PFTeDA
12	D5-N-EtFOSA	13C4-PFOS
13	D5-N-Et FOSA	
15	D5-E PFOSA	N/A
16	d5-N-EtFOSA	13C4-PFOA
17	PFOS-C8	EtFOSA-D5
18		d5-EtFOSA
19	d-NEtFOSA-M	NA
20	N	T
21		d-N-EtFOSA

Table 101 Labelled Standards for S1 EtFOSA

#### Table 102 Labelled Standards for S1 ADONA

Lab. Code	Before Extraction	Before Instrument Analysis			
1	NT				
2	13C PFOA				
3					
4	13C3 HFPO-DA				
5	Ν	IS			
6	N	IT			
7					
8	13C8-PFOS				
9	N	IT			
10					
11	N	ſΤ			
12	13C3-HFPO-DA	13C2-PFHxA			
13	13C4-PFOA				
15	13C4-PFHpA	N/A			
16					
17	PFOS-C8	FOSA-13C8			
18	N	IT			
19	MPFHpA	NA			
20	NT				
21	NT				

#### Table 103 Labelled Standards for S1 9Cl-PF3ONS

#### Table 104 Labelled Standards for S1 11Cl-PF3OUdS

Lab. Code	Before Extraction	Before Instrument Analysis			
1	NT				
2	MPFHxS				
3					
4	13C3 HFPO-DA				
5	Ν	NS			
6	Ν	Τ			
7					
8	13C8-PFOS				
9	NT				
10					
11	Ν	T			
12	13C3-HFPO-DA	13C2-PFHxA			
13	13C4-PFOS	13C8-PFOS			
15	13C4-PFOS	N/A			
16					
17	PFOS-C8				
18	N	νT			
19	M8PFOS	NA			
20	NT				
21	NT				

Lab. Code	Before Extraction	Before Instrument Analysis			
1	Ν	Τ			
2	MPFHxS				
3					
4	13C3 HFPO-DA				
5	Ν	IS			
6	Ν	T			
7					
8	13C2-PFHXA				
9	Ν	νT			
10					
11	Ν	νT			
12	13C3-HFPO-DA	13C2-PFHxA			
13	13C4-PFOS	13C8-PFOS			
15	13C4-PFOS	N/A			
16					
17	PFOS-C8				
18	Ν	NT			
19	MPFDoDA	NA			
20	NT				
21	Ν	Τ			

Lab. Code	S2 Sample Weight (g)	Sample Pretreatment	Extraction Technique	Extraction Solvent	Extraction Temperature	Extraction Time	Clean-Up
1	10	Homogenisation Geno/Grinder 14min & Centrifuge 10min	QuEChERS - modified AOAC	Acetonitrile with 1% Acetic Acid	Room	Sonicate 30 min at 30-35 degrees	envicarb
2							
3				NS			
4	1	1 Homogenisation Alkaline Digestion KOH-methanol Room temp 8 h		8 hrs	Active carbon SPE		
5	1	Freeze-drying	Soxhlet	MeOH	boiling point	4h	ion pair separation
6			Solid-Liquid Extraction				
7							
8	NS						
9							
10				NS			
11	1	No	Solid-Liquid Extraction	Acetonitrile	Room temperature	30 min	SPE-WAX, ultracentrifugation
12	2.14 (as received)	Homogenisation	Solid-Liquid Extraction	KOH-Methanol	Ambient Room Temperature	16 hours	Activated Carbon/SPE/Filtration
13	5	Homogenisation	Digestion with 10mM NaOH before QuEChERS extraction	Acetonitrile	Room temperature	30 min	Bond Elut Carbon, then SPE with Strata X-AW cartridges
15	2g	Homogenisation	Alkaline Digestion	Basified MeOH	Room	60 mins	Envicarb
16	2.5	Homogenisation	QuEChERS	Methanol + Ammonium-ACN and Acetone	40 °C	30 min	Solid-Phase Extraction
17	5	Homogenisation	Solid-Liquid Extraction	Acetonitrile	Ambient	30	Solid-Phase Extraction
18	0.5 grams	Homogenisation	Solid-Liquid Extraction	MTbE	Room	60 minutes	C18 & Activated Carbon

## Table 105 Participant Methodology – Sample S2 Carrot Extraction

Lab. Code	S2 Sample Weight (g)	Sample Pretreatment	Extraction Technique	Extraction Solvent	Extraction Temperature	Extraction Time	Clean-Up
19	2.024 and 2.082 (duplicate)	NA	Solid-Liquid Extraction (SLE) Merris-Minimix shaker	2% formic acid in acetonitrile	Room temperature	8 min	dSPE (C18, Envicarb, MgSO4)
20	2.5	Homogenisation	Solid-Liquid Extraction	0.1 % NH3 in ACN	room temperature	2 hours	Solid-Phase Extraction two different SPE cartridges
21			Alkaline Digestion				

## Table 106 Participant Methodology – Sample S2 Carrot Instrumental Technique and Analysis

Lab. Code	Instrument	Guard Column	Instrument Column	Dilution Factor	Delay Column?	Blank Correction?	Standard Method?	
1	Orbitrap	C18 3mm	Kinetex C18 100x3mm 2.6 um		Yes	Yes	In house	
2							US FDA Foods Program Compendium of Analytical Laboratory Methods; method C-010.02	
3	NS							
4	LC-MSMS or LC-QQQ	No	C18, 50 mm	No	Yes	No	No	
5	LC-MSMS or LC-QQQ	C18	Biphenyl 150x 2.5 mm	yes 1:10; 1:100	No	No	no	
6	LC-MSMS or LC-QQQ	C18	2.1x100mn 1.9um		Yes	No	USEPA 537	
7								
8			NS					
9								
10	NS							

Lab. Code	Instrument	Guard Column	Instrument Column	Dilution Factor	Delay Column?	Blank Correction?	Standard Method?
11	LC-MSMS or LC-QQQ	Gemini NX-C18; 4mm x 2.0mm ID	NX-C18; 15cm x 2mm x 3um	No	Yes	Yes	
12	LC-MSMS or LC-QQQ	Phenomenex Evo C18 (2µm, 2 mm x 2.1 mm)	BEH C18 (1.7μm, 50 mm x 2.1 mm)	No	Yes	No	No
13	LC-MSMS or LC-QQQ	Phenomenex Evo C18 (2 mm x 2.1 mm)	Phenomenex Evo C18 (100 mm x 2.1 mm x 2.6 um)	No	Yes	No	Isotopic Dilution
15	LC-MSMS or LC-QQQ	Pro column Hiltor () Jum ( (18 )		50	Yes	No	No. In-house
16	LC-MSMS or LC-QQQ	UltraShield UHPLC 0.2 µm Restek	Raptor C18 1.8 µm 50 x 2.1 mm Restek		yes	no	
17	Orbitrap	C18	C18		Yes		
18	LC-MSMS or LC-QQQ nil C		C18 1.6µm, 2.0mm x 50mm	No	Yes	No	
19	LC-MSMS or LC-QQQ	NA	Zorbax XDB-C18, 100 mm x 2.1 mm, 1.8µm	NA	Yes	No	No
20	LC-MSMS or LC-QQQ	C18 Column, 2.1 x 5 mm, 3.5 μm	C18 Column, 130Å, 3 x 50 mm, 3.5 µm		Yes	No	
21	LC-MSMS or LC-QQQ	C18	2.1x100mn 1.9um		Yes	No	USEPA 537

## $Table \ 107 \ Participant \ Methodology-Sample \ S2 \ Carrot \ Labelled \ Standards$

Lab. Code	Labelled Standard Source	Recovery Correction?	Labelled Standards Additional Information
1	Wellington	No	
2	Wellington	Yes	d5NN EtFOSAA added before instrument analysis
3			NS
4	Wellington	Yes	

Lab. Code	Labelled Standard Source	Recovery Correction?	Labelled Standards Additional Information
5	Wellington	Yes	
6	Wellington	no	
7			
8			NS
9			
10			NS
11			
12	Wellington	Yes	
13	Wellington	Yes	
15	Wellington	Yes	
16		Yes	
17	Wellington	Yes	Results corrected by ISTD added before instrumentation
18	Wellington Laboratories	No	
19	Wellington Laboratory	Yes	
20	Wellington	No	
21	Wellington	no	

#### Table 108 Labelled Standards for S2 PFBS

Lab. Code	Before Extraction	Before Instrument Analysis
1	Sodium perfluoro- 1-[2,3,4 13C3] butanesulfonate M3PFBS	
2	M3 PFBS	
3	N	S
4	13C3 PFBS	
5		
6		M3PFBS
7		
8	NS	
9	NT	
10	NS	
11	18O2-PFHxS	18O2-PFOS
12	13C3-PFBS	18O2-PFHxS
13	13C3-PFBS	13C3-PFHxS
15	13C3-PFBS	N/A
16	13C3-PFBS	13C4-PFOA
17	PFOS-C8	PFBS-13C3
18		13C3-PFBS
19	M3PFBS	NA
20	13C3-PFBS	13C2-PFHxA
21		M3PFBS

Before Extraction	Before Instrument Analysis
M PFHxS	
N	S
13C3 PFBS	
Ν	Т
NS	
NT	
NS	
18O2-PFHxS	18O2-PFOS
13C3-PFHxS	18O2-PFHxS
18O2-PFHxS	13C3-PFHxS
18O2-PFHxS	N/A
13C3-PFBS	13C4-PFOA
PFOS-C8	PFOS-C4
	16O2-PFHxS
M5PFHxA	NA
linear 13C3-PFHxS	13C2-PFHxA
	M PFHxS N 13C3 PFBS N 13C3 PFBS N N N N 13C3-PFHxS 13C3-PFHxS 18O2-PFHxS 18O2-PFHxS 18O2-PFHxS 18O2-PFHxS 13C3-PFBS PFOS-C8 M5PFHxA

Table 109 Labelled Standards for S2 PFPeS

Table 110 Labelled Standards for S2 PFHxS

Lab. Code	Before Extraction	Before Instrument Analysis
1	Sodium perfluoro- 1-[1,2,3 13C3] hexanesulfonate M3PFHxS	
2		
3	N	IS
4	13C3 PFHxS	PFHxS18O2
5	N	Т
6		
7		
8	NS	
9	NT	
10	NS	
11	NT	
12	13C3-PFHxS	18O2-PFHxS
13	18O2-PFHxS	13C3-PFHxS
15	18O2-PFHxS	N/A
16	18O2-PFHxS	13C4-PFOA
17	PFOS-C8	PFHxS-18O2
18		16O2-PFHxS
19	M3PFHxS	NA
20	linear 13C3-PFHxS	
21		

# Table 111 Labelled Standards for S2 PFHxS (linear)

(mittai)				
Lab. Code	Before Extraction	Before Instrument Analysis		
1				
2	M PFHxS			
3	N	S		
4	13C3 PFHxS			
5	yes			
6		M3PFHxS		
7				
8	NS			
9	NT			
10	NS			
11	18O2-PFHxS	18O2-PFOS		
12	13C3-PFHxS	18O2-PFHxS		
13	18O2-PFHxS	13C3-PFHxS		
15	18O2-PFHxS	N/A		
16	18O2-PFHxS	13C4-PFOA		
17	PFOS-C8	PFHxS-18O2		
18	NT			
19	M3PFHxS	NA		
20	linear 13C3-PFHxS			
21		M3PFHxS		

		1
Lab. Code	Before Extraction	Before Instrument Analysis
1		
2	M PFHxS	
3	N	IS
4	13C3 PFHxS	
5	N	Т
6		
7		
8	NS	
9	NT	
10	NS	
11	18O2-PFHxS	18O2-PFOS
12	13C8-PFOS	13C4-PFOS
13	18O2-PFHxS	13C3-PFHxS
15	13C4-PFOS	N/A
16	18O2-PFHxS	13C4-PFOA
17	PFOS-C8	PFOS-C4
18		13C8-PFOS
19	M3PFHxS	NA
20	linear 13C3-PFHxS	
21		

Table 112 Labelled Standards for S2 PFHpS

#### Table 113 Labelled Standards for S2 PFOS

Lab. Code	Before Extraction	Before Instrument Analysis
1	Sodium perfluoro- 1-[13C8] ocatanesulfonate M8PFOS	
2	13C PFOS	
3	N	S
4	13C8 PFOS	13C4 PFOS
5	N	Т
6		
7		
8	NS	
9	NT	
10	NS	
11	13C4-PFOS	18O2-PFOS
12	13C8-PFOS	13C4-PFOS
13	13C4-PFOS	13C8-PFOS
15	13C4-PFOS	N/A
16	13C4-PFOS	13C4-PFOA
17	PFOS-C8	PFOS-C4
18	13C4-PFOS	13C8-PFOS
19	M8PFOS	NA
20	linear 13C8-PFOS	13C5-PFNA
21		

# Table 114 Labelled Standards for S2 PFOS (linear)

	()		
Lab. Code	Before Extraction	Before Instrument Analysis	
1			
2			
3	N	IS	
4	13C8 PFOS		
5	yes		
6	MPFOS	M8PFOS	
7			
8	NS		
9	NT		
10	N	NS	
11	13C4-PFOS	18O2-PFOS	
12	13C8-PFOS	13C4-PFOS	
13	13C4-PFOS	13C8-PFOS	
15	13C4-PFOS	N/A	
16	13C4-PFOS	13C4-PFOA	
17	PFOS-C8	PFOS-C4	
18		13C8-PFOS	
19	M8PFOS	NA	
20	linear 13C8-PFOS		
21	MPFOS	M8PFOS	

Lab. Code	Before Extraction	Before Instrument Analysis
1		
2	13C PFOS	
3	N	IS
4	13C8 PFOS	
5	N	T
6		
7		
8	NS	
9	NT	
10	NS	
11	13C4-PFOS	18O2-PFOS
12	13C8-PFOS	13C4-PFOS
13	13C4-PFOS	13C8-PFOS
15	13C4-PFOS	N/A
16	NT	
17	PFOS-C8	PFBS-13C3
18	NT	
19	M8PFOS	NA
20	linear 13C8-PFOS	
21		
-		

Table 115 Labelled Standards for S2 PFNS

#### Table 116 Labelled Standards for S2 PFDS

Lab. Code	Before Extraction	Before Instrument Analysis
1	N	T
2	13C PFOS	
3	Ň	IS
4	13C8 PFOS	
5		
6		
7		
8	NS	
9	NT	
10	NS	
11	13C4-PFOS	18O2-PFOS
12	13C8-PFOS	13C4-PFOS
13	13C4-PFOS	13C8-PFOS
15	13C4-PFOS	N/A
16	13C2-PFUnA	13C4-PFOA
17	PFOS-C8	PFBA-13C4
18		13C8-PFOS
19	M8PFOS	NA
20	linear 13C8-PFOS	
21		

#### Table 117 Labelled Standards for S2 PFBA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [13C4]butanoic acid MPFBA	
2	M3 PFBA	
3	N	IS
4	13C4 PFBA	13C3 PFBA
5	yes	
6	M3PFBA	MPFBA
7		
8	NS	
9	NT	
10	NS	
11	13C4-PFBA	13C8-PFOA
12	13C4-PFBA	13C3-PFBA
13	13C4-PFBA	13C3-PFBA
15	13C4-PFBA	N/A
16	13C4-PFBA	13C4-PFOA
17	PFOS-C8	PFBA-13C4
18		13C4-PFBA
19	M4PFBA	NA
20	N	T
21	M3PFBA	MPFBA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [13C5]pentanoic acid M5PFPeA	
2	M3 PFPeA	
3	Ν	IS
4	13C5 PFPeA	
5		
6		M5PFPeA
7		
8	NS	
9	NT	
10	NS	
11	13C5-PFPeA	13C8-PFOA
12	13C5-PFPeA	13C2-PFHxA
13	13C4-PFPeA	13C5 -PFPeA
15	13C3-PFPeA	N/A
16	13C5-PFPeA	13C4-PFOA
17	PFOS-C8	PFPeA-13C3
18		13C5-PFPeA
19	M5PFPeA	NA
20	NT	
21		M5PFPeA

Table 118 Labelled Standards for S2 PFPeA

Table 119 Labelled Standards for S2 PFHxA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [1,2,3,4,6- 13C5]hexanoic acid M5PFHxA	
2	M3 PFHxA	
3	N	IS
4	13C5 PFHxA	13C2 PFHxA
5	yes	
6		M5PFHxA
7		
8	N	IS
9	NT	
10	N	IS
11	13C5-PFHxA	13C8-PFOA
12	13C5-PFHxA	13C2-PFHxA
13	13C2-PFHxA	13C5 -PFPeA
15	13C2-PFHxA	N/A
16	13C2-PFHxA	13C4-PFOA
17	PFOS-C8	PFHxA-13C2
18		13C5-PFHxA
19	M5PFHxA	NA
20	13C5-PFHxA	13C2-PFHxA
21		M5PFHxA

### Table 120 Labelled Standards for S2 PFHpA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [1,2,3,4- 13C4]heptanoic acid M4PFHpA	
2	M3 PFHxA	
3	Ν	١S
4	13C4 PFHpA	
5		
6		M4PFHpA
7		
8	NS	
9	NT	
10	NS	
11	13C4-PFHpA	13C8-PFOA
12	13C4-PFHpA	13C4-PFOA
13	13C3-PFHpA	13C8-PFOA
15	13C4-PFHpA	N/A
16	13C4-PFHpA	13C4-PFOA
17	PFOS-C8	PFHpA-13C4
18		13C4-PFHpA
19	MPFHpA	NA
20	13C4-PFHpA	13C2-PFHxA
21		M4PFHpA

Tuble 121 Euberied Standards for 52 11 071		
Before Extraction	Before Instrument Analysis	
Perfluoro-n- [13C8]octanoic acid M8PFOA		
13C PFOA		
N	IS	
13C8 PFOA	13C4 PFOA	
yes		
M2PFOA	M8PFOA	
NS		
NT		
NS		
13C4-PFOA	13C8-PFOA	
13C8-PFOA	13C4-PFOA	
13C4-PFOA	13C8-PFOA	
13C4-PFOA	N/A	
13C8-PFOA	13C4-PFOA	
PFOS-C8	PFOA-13C4	
13C8-PFOA	13C4-PFOA	
M8PFOA	NA	
13C8-PFOA	13C5-PFNA	
M2PFOA	M8PFOA	
	Perfluoro-n-         [13C8]octanoic         acid M8PFOA         13C PFOA         N         13C8 PFOA         yes         M2PFOA         N         13C4-PFOA         13C4-PFOA         13C4-PFOA         13C4-PFOA         13C8-PFOA         13C8-PFOA	

Table 121 Labelled Standards for S2 PFOA

#### Table 122 Labelled Standards for S2 PFNA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [13C9]nonanoic acid M9PFNA	
2	13C PFOA	
3	Ν	IS
4	13C9 PFNA	13C5 PFNA
5	yes	
6		M9PFNA
7		
8	NS	
9	NT	
10	N	IS
11	13C9-PFNA	13C5-PFNA
12	13C9-PFNA	13C5-PFNA
13	13C5-PFNA	13C8-PFOA
15	13C5-PFNA	N/A
16	13C5-PFNA	13C4-PFOA
17	PFOS-C8	PFNA-13C5
18		13C5-PFNA
19	M9PFNA	NA
20	13C9-PFNA	13C5-PFNA
21		M9PFNA

#### Table 123 Labelled Standards for S2 PFDA

Lab. Code	Before Extraction	Before Instrument Analysis
1	Perfluoro-n- [1,2,3,4,6- 13C6]decanoic acid M6PFDA	
2	13C PFOA	
3	N	IS
4	13C6 PFDA	13C2 PFDA
5	yes	
6	MPFDA	M6PFDA
7		
8	NS	
9	NT	
10	NS	
11	13C2-PFDA	13C5-PFNA
12	13C6-PFDA	13C2-PFDA
13	13C2-PFDA	13C8-PFOA
15	13C2-PFDA	N/A
16	13C2-PFDA	13C4-PFOA
17	PFOS-C8	PFDA-13C2
18		13C6-PFDA
19	M6PFDA	NA
20	13C6-PFDA	13C5-PFNA
21	MPFDA	M6PFDA

Table 12+ Eaberied Standards for 52 11 05/1		
Before Extraction	Before Instrument Analysis	
Perfluoro-1-[13C8] otanesulfonamide		
M8 FOSA		
N	S	
13C8 PFOSA		
N	Т	
	M8-FOSA	
NS		
NT		
NS		
13C8-PFOSA	13C2-PFTeDA	
13C8-PFOSA	13C4-PFOS	
13C8-FOSA		
13C8-FOSA	N/A	
13C8-PFOSA	13C4-PFOA	
PFOS-C8	FOSA-13C8	
	13C8-FOSA	
MPFOSA	NA	
NT		
	M8-FOSA	
	Before Extraction Perfluoro-1-[13C8] otanesulfonamide M8 FOSA M8 FOSA M8 FOSA M8 FOSA M8 FOSA M8 FOSA M8 M8 FOS M8 FOSA M8 FOS M	

Table 124 Labelled Standards for S2 PFOSA

#### Table 125 Labelled Standards for S2 MeFOSA

Lab. Code	Before Extraction	Before Instrument Analysis	
1	N-methyl-d3- perfluoro-1- octancesulfonamide		
2	N	Т	
3	N	IS	
4	d3-N-MeFOSA		
5	NT		
6		d-N-MeFOSA	
7			
8	NS		
9	NT		
10	N	NS	
11	13C8-PFOSA	13C2-PFTeDA	
12	D3-N-MeFOSA	13C4-PFOS	
13	D3-N-Me FOSA		
15	D3-M PFOSA	N/A	
16	d3-N-MeFOSA	13C4-PFOA	
17	PFOS-C8	MeFOSA-D3	
18		d3-MeFOSA	
19	d-NMeFOSA-M	NA	
20	N	Т	
21		d-N-MeFOSA	

#### Table 126 Labelled Standards for S2 EtFOSA

Lab. Code	Before Extraction	Before Instrument Analysis
1	N-ethyl-d5- perfluoro-1- octanesulfonamide	
2	N	Т
3	N	IS
4	d5-N-EtFOSA	
5	NT	
6		d-N-EtFOSA
7		
8	NS	
9	NT	
10	NS	
11	13C8-PFOSA	13C2-PFTeDA
12	D5-N-EtFOSA	13C4-PFOS
13	D5-N-Et FOSA	
15	D5-E PFOSA	N/A
16	d5-N-EtFOSA	13C4-PFOA
17	PFOS-C8	EtFOSA-D5
18		d5-EtFOSA
19	d-NEtFOSA-M	NA
20	N	T
21		d-N-EtFOSA

Table 127 Labelled Standards for S2 6:2 FTS

Lab. Code	Before Extraction	Before Instrument Analysis
1	M2-6:2FTS - 1H,1H,2H,2H- perfluoro1-[1,2-13C2]- octane sulfonate (6:2)	
2	13C2D4 6:2 FTS	
3	NS	
4	13C2 6:2 FTS	
5	NT	
6		M2-6:2FTS
7		
8	NS	
9	NT	
10	NS	
11	NT	
12	13C2-6:2 FTS	13C2-D4-6:2 FTS
13	13C2-6:2 FTS	
15	13C2,12C6 6:2-FTS	N/A
16	13C2-6:2FTS	13C4-PFOA
17	PFOS-C8	6:2 FTS-13C2
18		13C2-6:2 FTS
19	M6:2 FTS	NA
20	13C2-6:2 FTS	
21		M2-6:2FTS

Table 128 Labelled Standards for S2 GenX

Lab. Code	Before Extraction	Before Instrument Analysis
1	NT	
2	M3 HFPO	
3	NISTITIO N	10
		5
4	13C3 HFPO-DA	
5	N	Т
6		
7		
8	N	IS
9	NT	
10	NS	
11	NT	
12	13C3-HFPO-DA	13C2-PFHxA
13	13C4-PFOA	
15	13C312C3HF11O3	N/A
16		
17	PFOS-C8	PFPeA-13C3
18	NT	
19	M3HFPO-DA	NA
20	13C3-GenX	
21		

Lab. Code	Before Extraction	Before Instrument Analysis
1	N	T
2	13C PFOA	
3	Ň	IS
4	13C3 HFPO-DA	
5	N	ΪΤ
6		
7		
8	NS	
9	NT	
10	NS	
11	NT	
12	13C3-HFPO-DA	13C2-PFHxA
13	13C4-PFOA	
15	13C4-PFHpA	N/A
16		
17	PFOS-C8	FOSA-13C8
18	NT	
19	MPFHpA	NA
20	NT	
21	NT	

#### Table 129 Labelled Standards for S2 ADONA

#### APPENDIX 5 ACRONYMS AND ABBREVIATIONS

-	
4:2 FTS	4:2 Fluorotelomer sulfonate
6:2 FTS	6:2 Fluorotelomer sulfonate
8:2 FTS	8:2 Fluorotelomer sulfonate
10:2 FTS	10:2 Fluorotelomer sulfonate
9Cl-PF3ONS	9-chlorohexadecafluoro-3-oxanonane-1-sulfonate
11Cl-PF3OUdS	11-chloroeicosafluoro-3-oxaundecane-1-sulfonate
ACN	Acetonitrile
ADONA	Ammonium 4,8-dioxa-3H-perfluorononanoate
AV	Assigned Value
CITAC	Cooperation on International Traceability in Analytical Chemistry
CRM	Certified Reference Material
CV	Coefficient of Variation
dSPE	Dispersive SPE
EtFOSA	N-Ethyl perfluorooctane sulfonamide
EtFOSAA	N-Ethyl perfluorooctane sulfonamido acetic acid
EtFOSE	N-Ethyl perfluorooctane sulfonamido ethanol
FSANZ	Food Standards Australia New Zealand
GAG	General Accreditation Guidance (NATA)
GenX	Ammonium 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy) propanoate
GUM	Guide to the Expression of Uncertainty in Measurement
HV	Homogeneity Value
IEC	International Electrotechnical Commission
ISO	International Organization for Standardization
LC	Liquid Chromatography
LLE	Liquid-Liquid Extraction
LOR	Limit of Reporting
Max	Maximum
Md	Median
MeFOSA	N-Methyl perfluorooctane sulfonamide
MeFOSAA	N-Methyl perfluorooctane sulfonamido acetic acid
MeFOSE	N-Methyl perfluorooctane sulfonamido ethanol
MeOH	Methanol
Min	Minimum
MS	Mass Spectrometry

MS/MS	Tandem Mass Spectrometry
MTBE	Methyl <i>tert</i> -butyl ether
MU	Measurement Uncertainty
Ν	Number of numeric results
NA	Not Applicable
NATA	National Association of Testing Authorities, Australia
NMI	National Measurement Institute, Australia
NR	Not Reported
NS	Not Supplied
NT	Not Tested
PCV	Performance Coefficient of Variation
PFAS	Per- and Polyfluoroalkyl Substances
PFBA	Perfluorobutanoic acid
PFBS	Perfluorobutane sulfonate
PFDA	Perfluorodecanoic acid
PFDoA	Perfluorododecanoic acid
PFDoS	Perfluorododecane sulfonate
PFDS	Perfluorodecane sulfonate
PFHpA	Perfluoroheptanoic acid
PFHpS	Perfluoroheptane sulfonate
PFHxA	Perfluorohexanoic acid
PFHxS	Perfluorohexane sulfonate
PFNA	Perfluorononanoic acid
PFNS	Perfluorononane sulfonate
PFOA	Perfluorooctanoic acid
PFOS	Perfluorooctane sulfonate
PFOSA	Perfluorooctane sulfonamide
PFPeA	Perfluoropentanoic acid
PFPeS	Perfluoropentane sulfonate
PFTeDA	Perfluorotetradecanoic acid
PFTrDA	Perfluorotridecanoic acid
PFTrDS	Perfluorotridecane sulfonate
PFUdA	Perfluoroundecanoic acid
PFUdS	Perfluoroundecane sulfonate
PT	Proficiency Testing

QQQ	Triple Quadrupole Mass Spectrometry
QuEChERS	Quick, Easy, Cheap, Effective, Rugged and Safe extraction method
RA	Robust Average
Rec	Recovery
RM	Reference Material
S <sub>an</sub>	Analytical standard deviation
SD	Standard Deviation
SI	International System of Units
SLE	Solid-Liquid Extraction
SPE	Solid-Phase Extraction
SS	Spiked Samples
S <sub>sam</sub>	Between-sample standard deviation
SV	Spiked Value (or formulated concentration of a PT sample)
UPLC	Ultra Performance Liquid Chromatography
USEPA	United States Environmental Protection Agency
WAX	Weak Anion Exchange
σ	Target standard deviation for proficiency assessment

#### **END OF REPORT**