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National
Measurement
Institute

Proficiency Test Final Report AQA 23-10 Organic Compounds and Pesticides in Wastewater

October 2023

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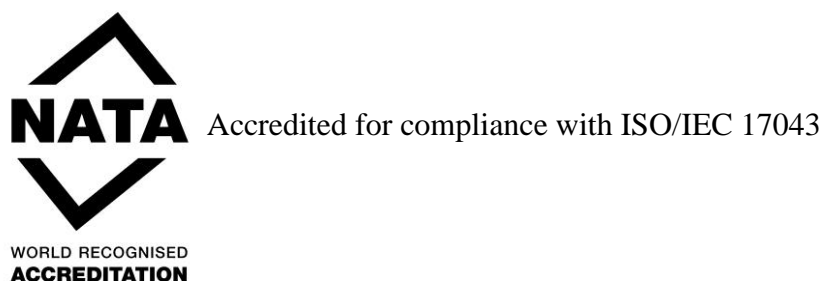


TABLE OF CONTENTS

SUMMARY	5
1 INTRODUCTION	7
1.1 NMI Proficiency Testing Program	7
1.2 Study Aims	7
1.3 Study Conduct	7
2 STUDY INFORMATION	8
2.1 Study Timetable	8
2.2 Participation and Laboratory Code	8
2.3 Selection of Analytes	8
2.4 Test Material Preparation	9
2.5 Homogeneity and Stability of Test Materials	9
2.6 Test Material Storage and Dispatch	10
2.7 Instructions to Participants	10
2.8 Interim Report and Preliminary Report	10
3 PARTICIPANT LABORATORY INFORMATION	11
3.1 Participants' Test Methods	11
3.2 Basis of Participants' Measurement Uncertainty Estimates	11
3.3 Participants' Comments	12
4 PRESENTATION OF RESULTS AND STATISTICAL ANALYSIS	13
4.1 Results Summary	13
4.2 Outliers and Extreme Outliers	13
4.3 Assigned Value	13
4.4 Robust Average and Robust Between-Laboratory Coefficient of Variation	13
4.5 Performance Coefficient of Variation	13
4.6 Target Standard Deviation for Proficiency Assessment	14
4.7 z-Score	14
4.8 E_r -Score	14
4.9 Traceability and Measurement Uncertainty	14
5 TABLES AND FIGURES	15
6 DISCUSSION OF RESULTS	61
6.1 Assigned Value	61
6.2 Measurement Uncertainty Reported by Participants	62
6.3 z-Score	62
6.4 E_r -Score	65
6.5 False Negatives	65
6.6 Reporting of Additional Analytes	66
6.7 Range of PAHs and Pesticides Analysed by Participants	66
6.8 Participants' Analytical Methods	67
6.9 Certified Reference Materials	72
6.10 Comparison with TRH and Total BTEX in River Water	73
6.11 Summary of Participants' Results and Performances	74
7 REFERENCES	76
APPENDIX 1 SAMPLE PREPARATION	77

A1.1 Diesel Fuel and Water Preparation	77
A1.2 Test Sample Preparation	77
APPENDIX 2 ASSESSMENT OF HOMOGENEITY AND STABILITY	78
A2.1 Homogeneity	78
A2.2 Stability	78
APPENDIX 3 ROBUST AVERAGE AND ASSOCIATED UNCERTAINTY, z-SCORE AND E_r -SCORE CALCULATIONS	80
A3.1 Robust Average and Associated Uncertainty	80
A3.2 z-Score and E_r -Score Calculation	80
APPENDIX 4 PARTICIPANTS' TEST METHODS	81
APPENDIX 5 ACRONYMS AND ABBREVIATIONS	85

SUMMARY

AQA 23-10 Organic Compounds and Pesticides in Wastewater commenced in June 2023. Twelve laboratories registered to participate, and eleven participants submitted results.

The sample set consisted of four wastewater samples. Samples were prepared in the NMI Sydney laboratory by spiking wastewater with various analytes.

Of a possible 253 results, 206 numeric results (81%) were submitted. Twenty-two results were a 'less than' value ($< x$) or Not Reported (NR), and 25 results were Not Tested (NT).

The assigned values for all scored analytes were the robust averages of participants' results. The associated uncertainties were estimated from the robust standard deviations of the participants' results.

Traceability: The consensus of participants' results is not traceable to any external reference, so although expressed in SI units, metrological traceability has not been established.

The outcomes of the study were assessed against the aims as follows:

- *Assess the ability of participants to correctly identify organic compounds and pesticides in wastewater.*

Laboratories **3, 4, 6, 7** and **10** reported numeric results for all 16 scored analytes.

Two participants did not report numeric results for analytes which they tested for and were present in the test samples (total of two results). Three participants reported numeric results for analytes not spiked into the test samples (total of five results).

- *Compare the performance of participants and assess their accuracy in the measurement of organic compounds and pesticides in wastewater.*

Laboratory performance was assessed using both z -scores and E_n -scores.

Of 162 z -scores, 157 (97%) returned a score of $|z| \leq 2.0$, indicating a satisfactory performance.

Of 151 E_n -scores, 128 (85%) returned a score of $|E_n| \leq 1.0$, indicating agreement of the participant's result with the assigned value within their respective expanded uncertainties.

Laboratory **4** achieved satisfactory z - and E_n -scores across all 16 scored analytes.

- *Evaluate the participants' methods for the measurement of organic compounds and pesticides in wastewater.*

For Sample S1 TRH, most participants used liquid-liquid extraction with dichloromethane, with analysis on GC-FID.

For Sample S2 BTEX, all participants used purge-and-trap GC-MS(/MS).

For Sample S3 PAHs, a wide variety of extraction techniques and solvents were used, however all participants used GC-MS(/MS) for analysis.

For Sample S4 Pesticides, a wide variety of procedures were employed by participants.

- *Develop the practical application of traceability and measurement uncertainty, and provide participants with information that will be useful in assessing their uncertainty estimates.*

Of 206 numeric results, 201 (98%) were reported with an expanded measurement uncertainty. The magnitude of reported uncertainties was within the range of 1.3% to 50%. Participants used a wide variety of procedures to estimate their uncertainty.

- *Produce materials that can be used in method validation and as control samples.*

The test samples of this study are homogeneous and are well characterised. Surplus samples are available for purchase from NMI and can be used for quality control and method validation purposes.

1 INTRODUCTION

1.1 NMI Proficiency Testing Program

The National Measurement Institute (NMI) is responsible for Australia's national measurement infrastructure, providing a range of services including a chemical proficiency testing program.

Proficiency testing (PT) is the 'evaluation of participant performance against pre-established criteria by means of interlaboratory comparison'.¹ NMI PT studies target chemical testing in areas of high public significance such as trade, environment, law enforcement and food safety. NMI offers studies in:

- pesticide residues in soil and water, fruit, vegetables and herbs;
- petroleum hydrocarbons and volatile organic compounds in soil and water;
- inorganic analytes in soil, water, filters, food and pharmaceuticals;
- per- and polyfluoroalkyl substances in soil, water, biota and food;
- controlled drug assay, drugs in wipes and clandestine laboratory; and
- allergens in food.

1.2 Study Aims

The aims of the study were to:

- assess the ability of participants to correctly identify organic compounds and pesticides in wastewater;
- compare the performance of participants and assess their accuracy in the measurement of organic compounds and pesticides in wastewater;
- evaluate the participants' methods for the measurement of organic compounds and pesticides in wastewater;
- develop the practical application of traceability and measurement uncertainty, and provide participants with information that will be useful in assessing their uncertainty estimates; and
- produce materials that can be used in method validation and as control samples.

The choice of the test method was left to the participating laboratories.

1.3 Study Conduct

The conduct of NMI PT studies is described in the NMI Study Protocol for Proficiency Testing.² The statistical methods used are described in the NMI Chemical Proficiency Testing Statistical Manual.³ These documents have been prepared with reference to ISO/IEC 17043,¹ and The International Harmonized Protocol for the Proficiency Testing of Analytical Chemistry Laboratories.⁴

NMI is accredited by the National Association of Testing Authorities, Australia (NATA) to ISO/IEC 17043 as a provider of proficiency testing schemes.¹ This study falls within the scope of NMI's accreditation.

2 STUDY INFORMATION

2.1 Study Timetable

The timetable of the study was:

Invitations sent	13/06/2023
Samples sent	3/07/2023
Results due	7/08/2023
Interim Report	10/08/2023
Preliminary Report	12/09/2023

The release of the Preliminary Report was delayed due to an internal investigation relating to Sample S2 (see Section 2.8).

2.2 Participation and Laboratory Code

Twelve laboratories registered to participate in this study, and all participants were assigned a confidential laboratory code number for this study. Eleven participants submitted results.

2.3 Selection of Analytes

When selecting analytes and spiking values for this study, consideration was given to:

- a variety of analytes amenable to gas and/or liquid chromatography; and
- feedback and suggestions from participants and other stakeholders.

For Sample S1, participants were requested to measure semi-volatile hydrocarbons (>C10-C40) and total recoverable hydrocarbons (TRH). For Sample S2, participants were requested to measure volatile hydrocarbons (C6-C10), and benzene, toluene, ethylbenzene and xylenes (BTEX). For Sample S3, participants were provided with a list of potential poly-aromatic hydrocarbons (PAHs) that were spiked the sample (Table 1). For Sample S4, participants were provided with a list of potential pesticides that were spiked into the sample (Table 2).

Table 1 List of Possible PAHs for Sample S3

Naphthalene	Fluorene	Benz[<i>a</i>]anthracene	Benzo[<i>a</i>]pyrene
Acenaphthylene	Phenanthrene	Chrysene	Indeno[1,2,3- <i>cd</i>]pyrene
Acenaphthene	Fluoranthene	Benzo[<i>b</i>]fluoranthene	Dibenz[<i>a,h</i>]anthracene
Anthracene	Pyrene	Benzo[<i>k</i>]fluoranthene	Benzo[<i>g,h,i</i>]perylene

Table 2 List of Possible Pesticides for Sample S4

Aldicarb	Dicamba	Omethoate
Aldrin	Dieldrin	Parathion
Atrazine	Dimethoate	Parathion-methyl
Azinphos-methyl	Diuron	Pendimethalin
Chlorpyrifos	Endosulfan	Permethrin
Chlordane	Ethion	Picloram
Chlorfenvinphos	Fenthion	Piperonyl butoxide
Clopyralid	Heptachlor	Pirimicarb

Cyfluthrin	Imazapyr	Pirimphos-ethyl
Cypermethrin	Lindane	Pirimphos-methyl
2,4-D	Malathion	Propiconazole
DDT	MCPA	Simazine
Deltamethrin	Metolachlor	2,4,5-T
Diazinon	Metsulfuron-methyl	Tetrachlorvinphos

2.4 Test Material Preparation

The test samples were prepared by spiking processed wastewater with various analytes to obtain the concentrations listed in Table 3. Additional information on the preparation of the samples is given in Appendix 1.

Table 3 Spiked Values of Test Samples

Sample	Analyte	Spiked Value ($\mu\text{g/L}$)	Uncertainty* ($\mu\text{g/L}$)
S1	TRH	2810	140
S2	Benzene	69.2	3.5
	Toluene	261	13
	Ethylbenzene	29.0	1.5
	Xylenes	129	6
	Total BTEX	489	24
S3	Acenaphthene	8.08	0.40
	Acenaphthylene	1.09	0.05
	Benz[<i>a</i>]anthracene	6.50	0.32
	Benzo[<i>a</i>]pyrene	3.62	0.18
	Fluorene	3.10	0.15
	Phenanthrene	1.95	0.10
S4	Atrazine	11.6	0.6
	Chlorpyrifos	20.1	1.0
	Dicamba	7.49	0.37
	Dieldrin	5.01	0.25
	Ethion	3.99	0.20
	Lindane	10.0	0.5
	Simazine	5.05	0.25

* Estimated expanded uncertainty at time of spiking at approximately 95% confidence using a coverage factor of 2.

2.5 Homogeneity and Stability of Test Materials

No homogeneity or stability testing was conducted for this study. The samples were prepared, packaged and stored using a process that has been demonstrated to produce sufficiently homogeneous and stable samples in previous NMI PT studies with similar analytes and matrices.

Participants' results gave no reason to question the homogeneity or transportation stability of the samples (Appendix 2). Assigned values were set if there was a reasonable consensus between participants' results.

2.6 Test Material Storage and Dispatch

After preparation, the samples were stored at 4 °C. Samples were packaged into insulated polystyrene foam boxes with cooler bricks and dispatched by courier on 3 July 2023.

The following items were packaged with the samples:

- a covering letter which included a description of the test samples and instructions for participants; and
- a form for participants to confirm the receipt and condition of the test samples.

An Excel spreadsheet for the electronic reporting of results was emailed to participants.

2.7 Instructions to Participants

Participants were instructed as follows:

- Quantitatively analyse the samples using your routine test method.
- Do not test for volatile hydrocarbons (C6-C10) or BTEX components in Sample S1.
- Participants need not test for all listed analytes.
- For each analyte in each sample, report a single result in units of $\mu\text{g/L}$ expressed as if reporting to a client, applying the limit of reporting of the method used for analysis. This is the figure that will be used in all statistical analysis in the study report. Also, reported the associated expanded uncertainty in units of $\mu\text{g/L}$ (e.g. $2000 \pm 200 \mu\text{g/L}$), if determined.
- If an analyte was not tested for, please report 'NT' as its result.
- Give details of your methodology and basis of uncertainty estimate as requested by the results sheet.
- Return the completed results sheet by 31 July 2023 by email to proficiency@measurement.gov.au.

The results due date was later extended to 7 August 2023 due to issues affecting some participants.

2.8 Interim Report and Preliminary Report

An Interim Report was emailed to all participants on 10 August 2023.

A Preliminary Report was emailed to all participants on 12 September 2023. The Preliminary Report included a summary of the results reported by participants, assigned values, performance coefficient of variations, z -scores and E_n -scores for each analyte in this study. No data from the Preliminary Report has been changed in the present Final Report.

The release of this Preliminary Report was delayed because of an internal investigation for Sample S2 ethylbenzene. This was due to a greater variability of participants' results as compared to the other BTEX analytes. This variability was also observed in the internal investigation, and so this analyte was not scored.

3 PARTICIPANT LABORATORY INFORMATION

3.1 Participants' Test Methods

Participants were requested to provide information about their test methods. Responses are presented in Appendix 4.

3.2 Basis of Participants' Measurement Uncertainty Estimates

Participants were requested to provide information about the basis of their measurement uncertainty (MU) estimates. Responses are presented in Table 4. Some responses were modified so that the participant cannot be identified.

Table 4 Basis of Measurement Uncertainty Estimate

Lab. Code	Approach to Estimating MU	Information Sources for MU Estimation*		Guide Document for Estimating MU
		Precision	Method Bias	
1	Top Down - precision and estimates of the method and laboratory bias	Control samples - SS Duplicate analysis Instrument calibration	Instrument calibration Recoveries of SS Standard purity	Eurachem/CITAC Guide
2	Top Down - precision and estimates of the method and laboratory bias	Control samples - CRM Duplicate analysis Instrument calibration	CRM Instrument calibration Recoveries of SS	Eurachem/CITAC Guide
3	Top Down - precision and estimates of the method and laboratory bias	Control samples - CRM	Recoveries of SS	Eurachem/CITAC Guide
4	Top Down - precision and estimates of the method and laboratory bias	Control samples - RM Duplicate analysis Instrument calibration	Instrument calibration Recoveries of SS	ISO/GUM
5	Top Down - precision and estimates of the method and laboratory bias	Control samples - SS Duplicate analysis Instrument calibration	Instrument calibration Recoveries of SS	Eurachem/CITAC Guide
6	Top Down - precision and estimates of the method and laboratory bias	Control samples - RM Duplicate analysis	CRM Instrument calibration	Eurachem/CITAC Guide
7	Bottom Up (ISO/GUM, fish bone/cause and effect diagram)	Control samples Duplicate analysis Instrument calibration	Instrument calibration Laboratory bias from PT studies Recoveries of SS	Eurachem/CITAC Guide
8	Top Down - precision and estimates of the method and laboratory bias	Control samples - CRM Duplicate analysis Instrument calibration	CRM Instrument calibration	Eurachem/CITAC Guide
10	Top Down - reproducibility (standard deviation) from PT studies used directly	Control samples - SS Duplicate analysis Instrument calibration		Eurachem/CITAC Guide
11	Top Down - precision and estimates of the method and laboratory bias	Control samples Instrument calibration	Instrument calibration Recoveries of SS Standard purity	Eurachem/CITAC Guide
12	Bottom Up (ISO/GUM, fish bone/cause and effect diagram)	Duplicate analysis		ISO/GUM

* SS = Spiked Samples, RM = Reference Material, CRM = Certified Reference Material

3.3 Participants' Comments

Participants were invited to make comments or suggestions on the samples, this study, or possible future studies. Such feedback may be useful in improving future studies. Participants' comments received for this study are presented in Table 5, along with the study coordinator's response where applicable. Some responses may be modified so that the participant cannot be identified.

Table 5 Participants' Comments

Lab. Code	Sample	Participant's Comments	Study Coordinator's Response
3	S1	A 100mL aliquot was subsampled from the 500mL bottle and prepared as per procedure.	
	S2	The laboratory had noted variations associated with the concentration of Ethylbenzene in this study. Each vial had a slightly different concentration to the next and confirmed on the same instrumentation. The laboratory obtained Ethylbenzene concentrations between 2.5 to 18.8 ug/L. A raised limit of report of 20 ug/L was used due to the high %RPD seen amongst each vial.	Further investigation was performed by NMI for this analyte, and some variability was observed. Therefore, no assigned value has been set for Sample S2 ethylbenzene to ensure participants were not negatively affected with regards to scoring.
	All	Please prepare more samples in the 100mL approach for SV analysis.	Thank you for your feedback. We are looking into providing 100 mL bottles as an option for other future PT studies.
11	S4	Propazine detected at 0.05 ug/L	

4 PRESENTATION OF RESULTS AND STATISTICAL ANALYSIS

4.1 Results Summary

Participant results are listed in Tables 6 to 28 with summary statistics: robust average, median, mean, number of numeric results (N), maximum (Max), minimum (Min), robust standard deviation (Robust SD) and robust coefficient of variation (Robust CV). Bar charts of results and performance scores are presented in Figures 2 to 24. An example chart with interpretation guide is shown in Figure 1.

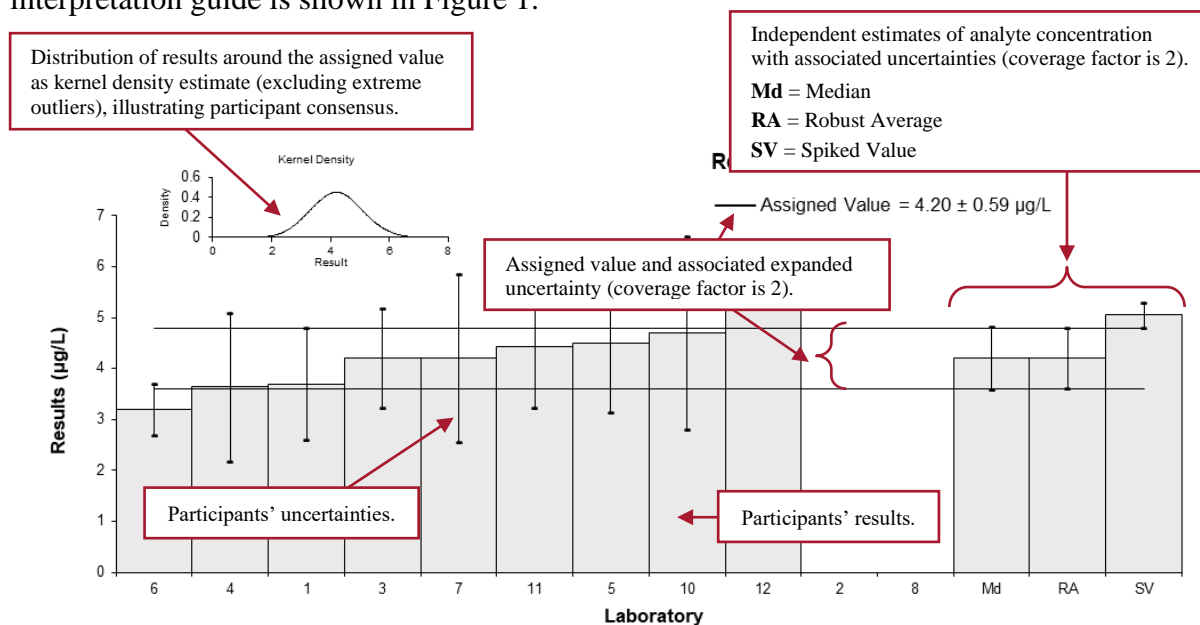


Figure 1 Guide to Presentation of Results

4.2 Outliers and Extreme Outliers

Outliers were results less than 50% and greater than 150% of the robust average, and these were removed before the calculation of the assigned value.^{3,4} Extreme outliers were obvious blunders, e.g. results reported with incorrect units or for a different analyte or sample, and such results were removed for the calculation of all summary statistics.^{3,4}

4.3 Assigned Value

The assigned value is defined as the 'value attributed to a particular property of a proficiency test item'.¹ In this PT study, this property is the concentration of the analytes in the samples. Assigned values were the robust averages of participants' results, and the expanded uncertainties were estimated from the associated robust SDs (Appendix 3).

4.4 Robust Average and Robust Between-Laboratory Coefficient of Variation

The robust averages and associated expanded uncertainties, and robust CVs (a measure of the variability of participants' results) were calculated using the procedure described in ISO 13528.⁵

4.5 Performance Coefficient of Variation

The performance coefficient of variation (PCV) is a fixed measure of the between-laboratory variation that in the judgement of the study coordinator would be expected from participants given the analyte concentrations. The PCV is not the CV of participants' results; it is set by the study coordinator and is based on the analyte concentrations and experience from previous studies, and is supported by mathematical models such as the Thompson-Horwitz equation.⁶ By setting a fixed and realistic value for the PCV, a participant's performance does not depend on other participants' performances and can be compared from study to study.

4.6 Target Standard Deviation for Proficiency Assessment

The target standard deviation for proficiency assessment (σ) is the product of the assigned value (X) and the PCV, as presented in Equation 1.

$$\sigma = X \times PCV \quad \text{Equation 1}$$

4.7 z-Score

For each participant result, a z-score is calculated according to Equation 2.

$$z = \frac{(\chi - X)}{\sigma} \quad \text{Equation 2}$$

where:

- z is z-score
- χ is a participant's result
- X is the assigned value
- σ is the target standard deviation from Equation 1

For the absolute value of a z-score:

- $|z| \leq 2.0$ is satisfactory;
- $2.0 < |z| < 3.0$ is questionable; and
- $|z| \geq 3.0$ is unsatisfactory.

To account for potential low bias in the consensus value due to inefficient methodologies, scores may be adjusted for a 'maximum acceptable result' (see Section 6.3).

4.8 E_n -Score

The E_n -score is complementary to the z-score in the assessment of laboratory performance. The E_n -score includes expanded uncertainty and is calculated according to Equation 3.

$$E_n = \frac{(\chi - X)}{\sqrt{U_\chi^2 + U_X^2}} \quad \text{Equation 3}$$

where:

- E_n is E_n -score
- χ is a participant's result
- X is the assigned value
- U_χ is the expanded uncertainty of the participant's result
- U_X is the expanded uncertainty of the assigned value

For the absolute value of an E_n -score:

- $|E_n| \leq 1.0$ is satisfactory; and
- $|E_n| > 1.0$ is unsatisfactory.

4.9 Traceability and Measurement Uncertainty

Laboratories accredited to ISO/IEC 17025 must establish and demonstrate the traceability and MU associated with their test results.⁷

Guidelines for quantifying uncertainty in analytical measurement are described in the Eurachem/CITAC Guide.⁸

5 TABLES AND FIGURES

Table 6

Sample Details

Sample No.	S1
Matrix	Wastewater
Analyte	>C10-C16
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty
1	950	300
2	903	261.87
3	770	326
4	420	130
5	400	120
6	940	282
7	250	85
8	700	190
10	510	200
11	1056	296
12	705	20

Statistics

Assigned Value	Not Set	
Spike Value	Not Spiked	
Robust Average	690	230
Median	710	260
Mean	690	
N	11	
Max	1056	
Min	250	
Robust SD	300	
Robust CV	43%	

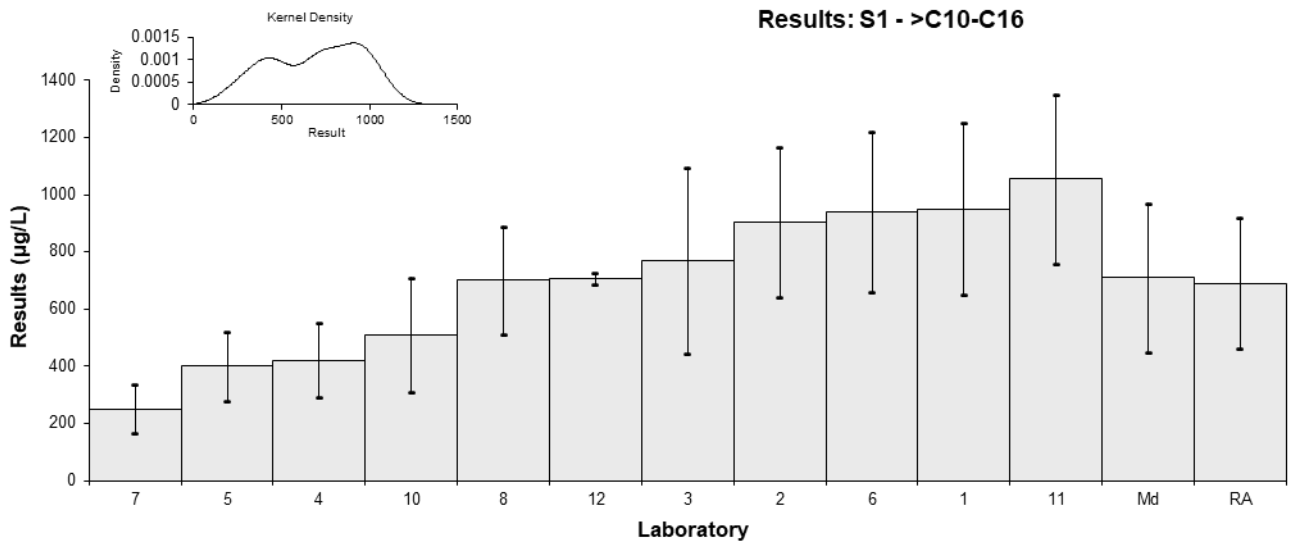


Figure 2

Table 7

Sample Details

Sample No.	S1
Matrix	Wastewater
Analyte	>C16-C34
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty
1	840	300
2	1111	333.3
3	810	258
4	459	140
5	<300	NR
6	890	270
7	310	120
8	790	300
10	350	140
11	1273	344
12	520	20

Statistics

Assigned Value	Not Set	
Spike Value	Not Spiked	
Robust Average	740	290
Median	800	350
Mean	740	
N	10	
Max	1273	
Min	310	
Robust SD	360	
Robust CV	49%	

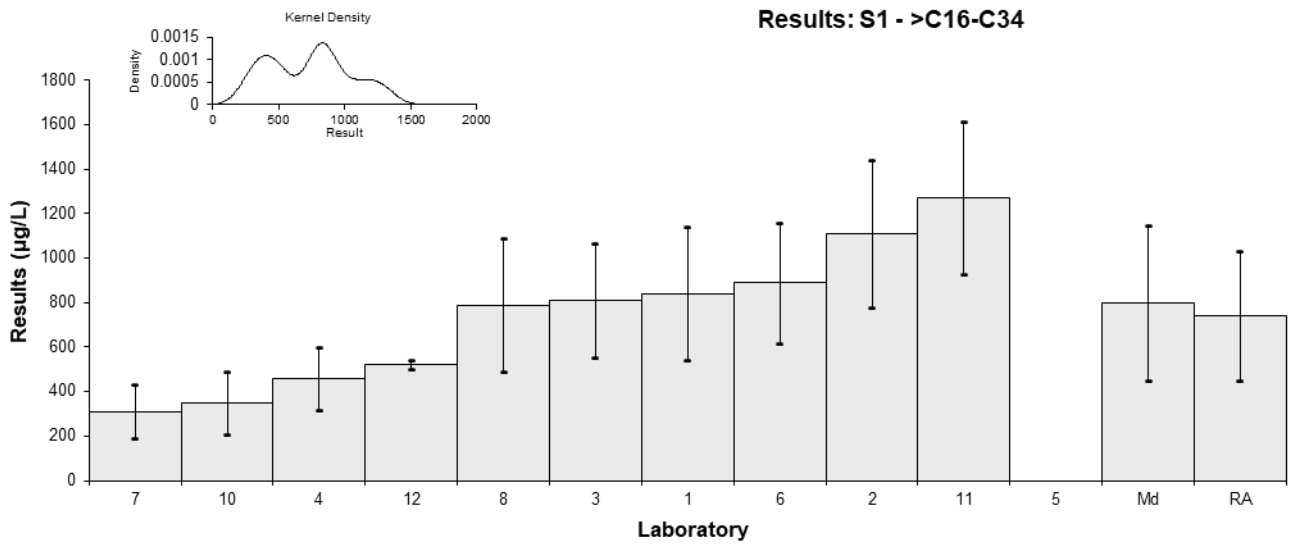


Figure 3

Table 8

Sample Details

Sample No.	S1
Matrix	Wastewater
Analyte	>C34-C40
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty
1	<500	NR
2	<200	NR
3	<100	NR
4	<50	NR
5	<300	NR
6	<100	NR
7	<100	62
8	<100	NR
10	<100	NR
11	222	58
12	<20	20

Statistics

Assigned Value	Not Set	
Spike Value	Not Spiked	
Robust Average	NA (N<6)	
Median	NA (N<3)	
Mean	NA (N<2)	
N	1	
Max	222	
Min	222	
Robust SD	NA (N<6)	
Robust CV	NA (N<6)	

Results: S1 - >C34-C40

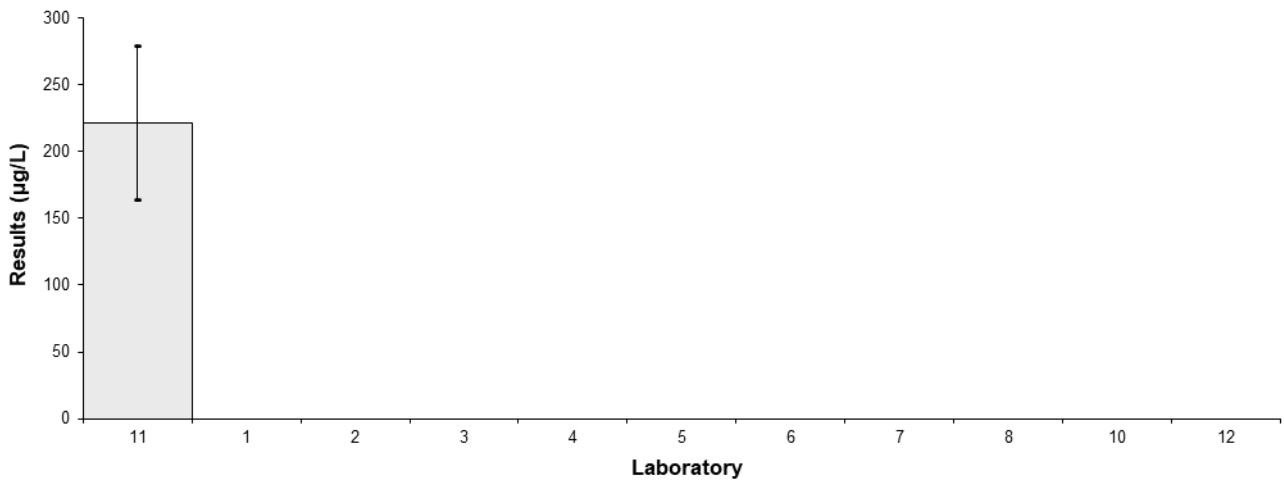


Figure 4

Table 9

Sample Details

Sample No.	S1
Matrix	Wastewater
Analyte	TRH
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E _n
1	1790	500	1.44	0.61
2	2014	NR	2.00▼	
3	1580	601	0.68	0.26
4	900	270	-1.76	-0.97
5	800	240	-2.12	-1.20
6	1830	NR	1.58	1.02
7*	490	98	-3.24	-2.04
8	1490	415	0.36	0.17
10	860	340	-1.91	-0.97
11*	2560	770	2.00▼	
12	1220	50	-0.61	-0.39

* Outlier, see Section 4.2; ▼ Adjusted Score, see Section 6.3

Statistics

Assigned Value	1390	430
Spike Value	2810	140
Robust Average	1400	510
Max Acceptable Result	3930	
Median	1490	590
Mean	1410	
N	11	
Max	2560	
Min	490	
Robust SD	670	
Robust CV	48%	

Laboratory 2 did not report a TRH value. The study coordinator summed the individual hydrocarbon ranges, and no estimate of uncertainty of the TRH result was made.

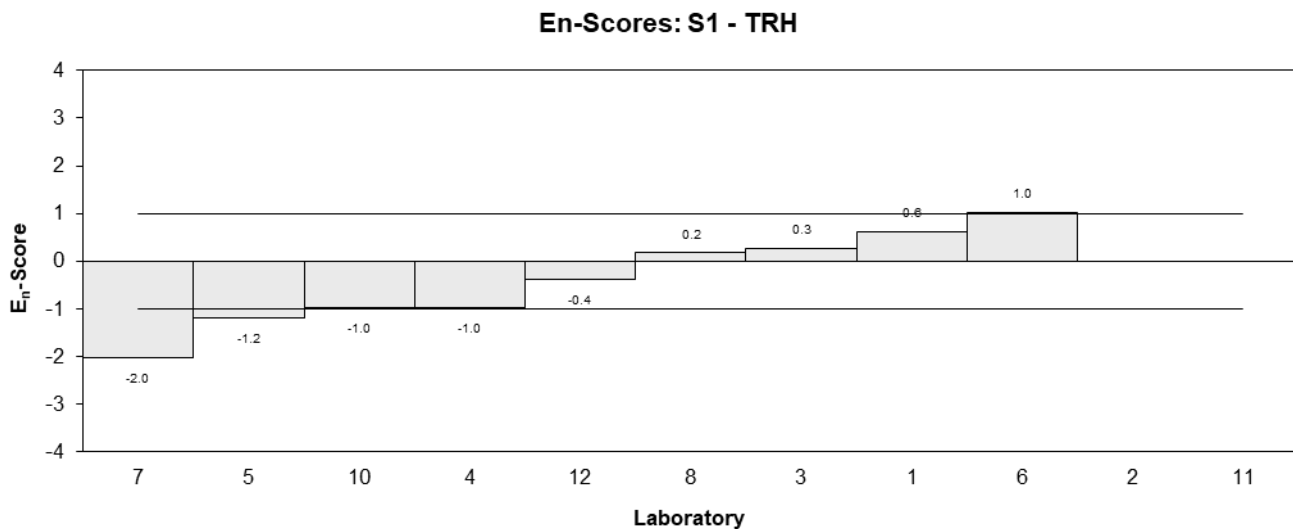
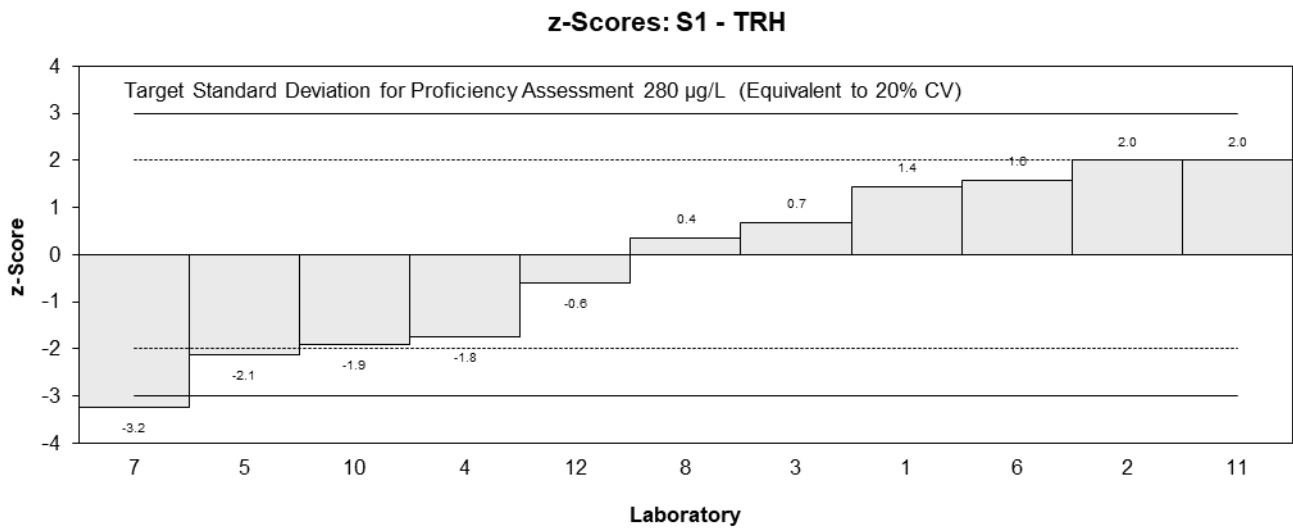
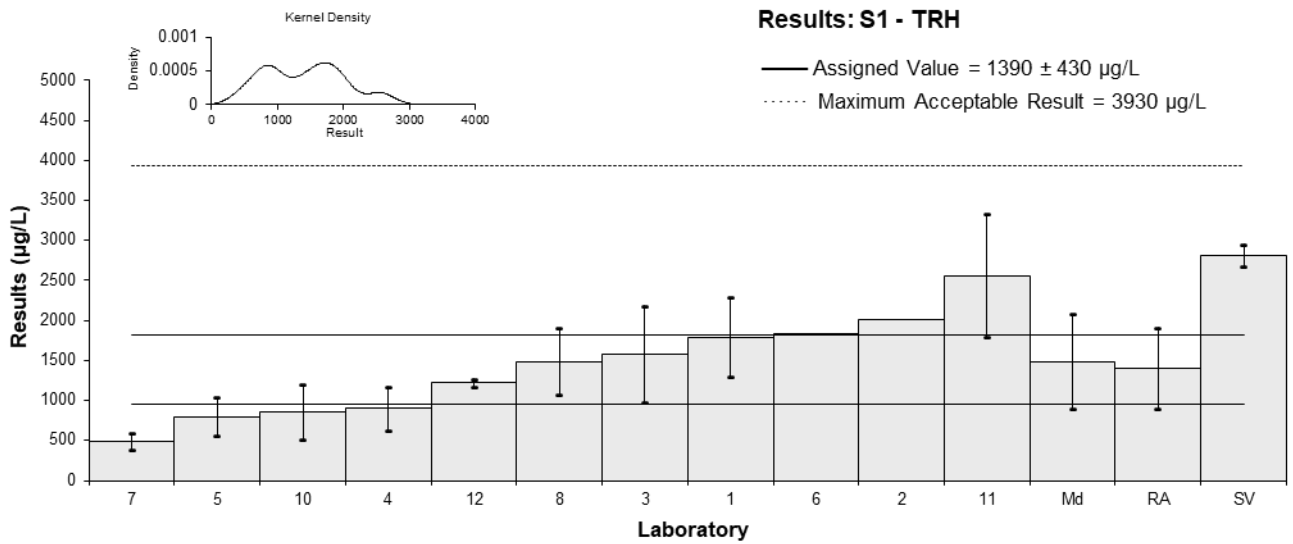


Figure 5

Table 10

Sample Details

Sample No.	S2
Matrix	Wastewater
Analyte	C6-C10
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty
1	NT	NT
2	1016	209.296
3	976	202.4
4	1190	360
5	NT	NT
6	970	191
7	1150	255
8	1060	237
10	580	240
11	NT	NT
12	NT	NT

Statistics

Assigned Value	Not Set	
Spike Value	Not Spiked	
Robust Average	1020	140
Median	1020	60
Mean	990	
N	7	
Max	1190	
Min	580	
Robust SD	140	
Robust CV	14%	

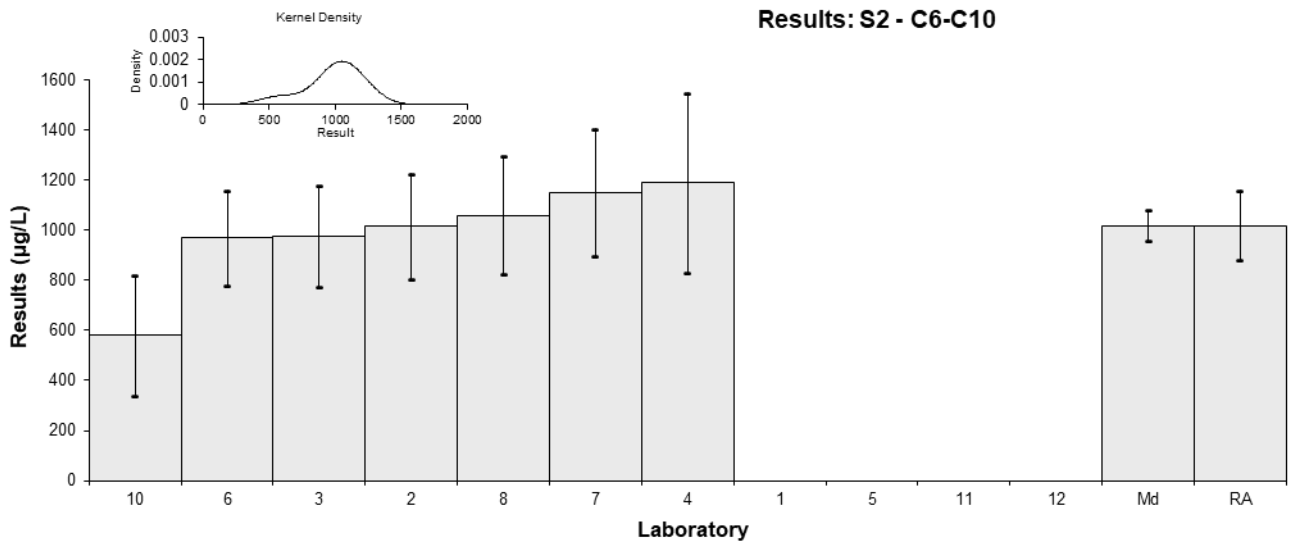


Figure 6

Table 11

Sample Details

Sample No.	S2
Matrix	Wastewater
Analyte	Benzene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	NT	NT		
2	56	9.856	-0.54	-0.61
3	68	12	0.41	0.40
4	70.02	21	0.57	0.33
5	54.5	16.35	-0.66	-0.48
6	65	10	0.18	0.20
7	63	12.4	0.02	0.01
8	64	11	0.10	0.10
10	67	13	0.33	0.30
11	58	1.3	-0.38	-0.90
12	NT	NT		

Statistics

Assigned Value	62.8	5.2
Spike Value	69.2	3.5
Robust Average	62.8	5.2
Median	64.0	4.9
Mean	62.8	
N	9	
Max	70.02	
Min	54.5	
Robust SD	6.2	
Robust CV	9.9%	

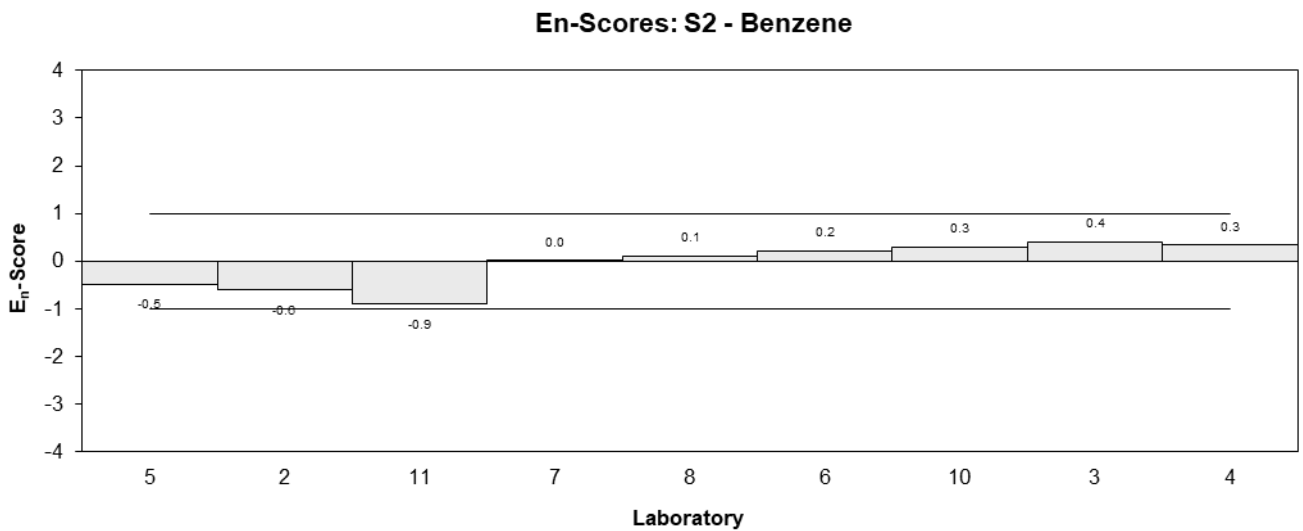
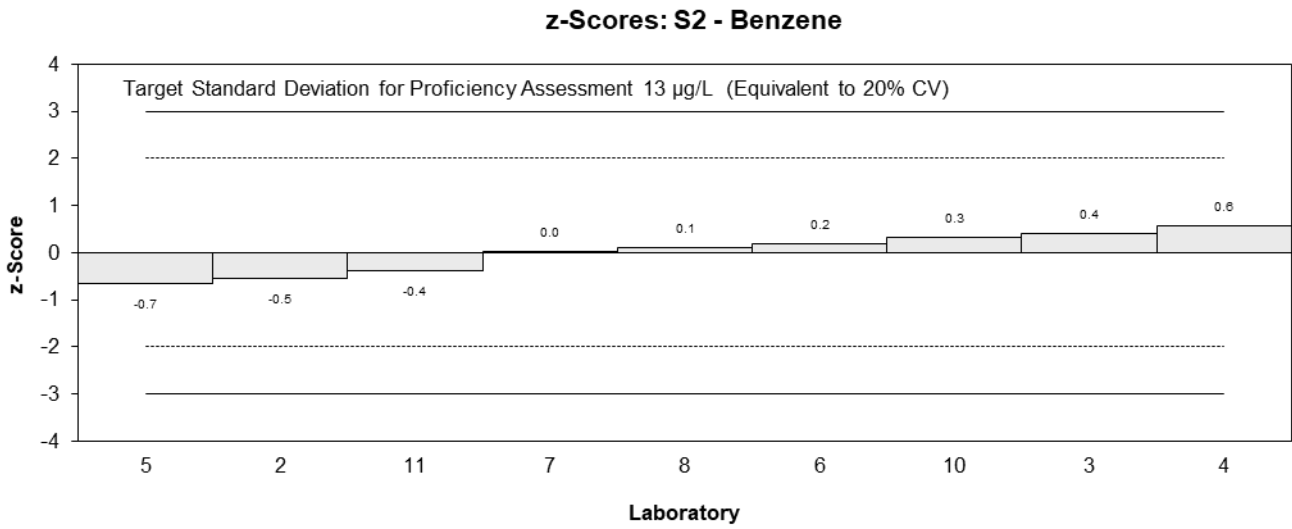
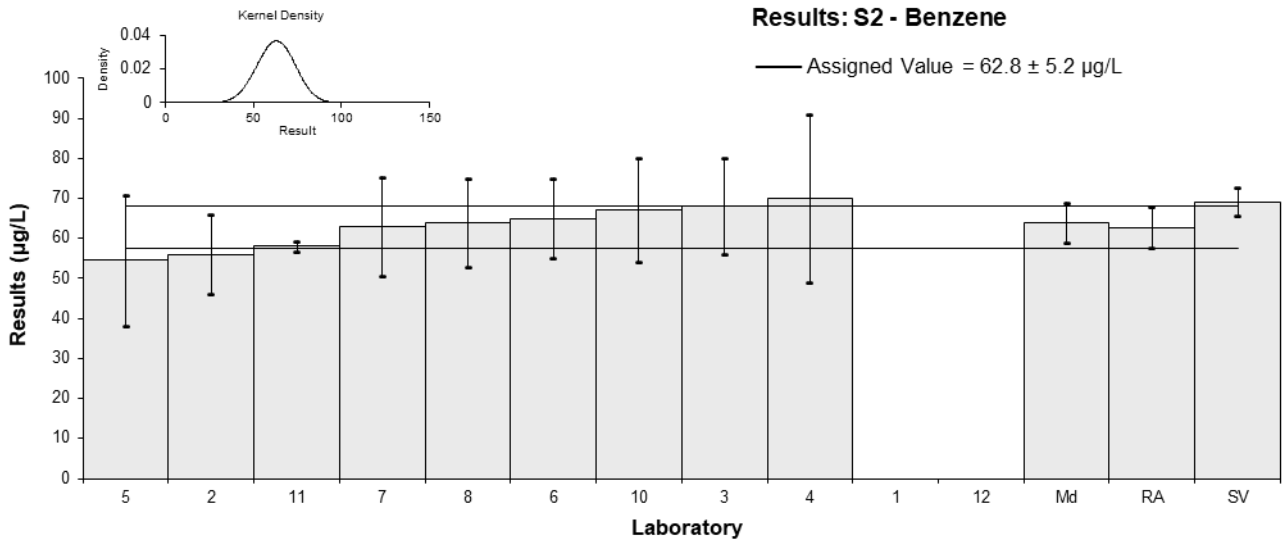


Figure 7

Table 12

Sample Details

Sample No.	S2
Matrix	Wastewater
Analyte	Toluene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	NT	NT		
2	209	24.662	0.17	0.26
3	215	38.4	0.32	0.33
4	183.9	55	-0.45	-0.32
5	185	55.5	-0.42	-0.30
6	206	31	0.10	0.12
7	208	42	0.15	0.14
8	209	35	0.17	0.19
10	200	40	-0.05	-0.05
11	204	47	0.05	0.04
12	NT	NT		

Statistics

Assigned Value	202	10
Spike Value	261	13
Robust Average	202	10
Median	206	4
Mean	202	
N	9	
Max	215	
Min	183.9	
Robust SD	12	
Robust CV	6.1%	

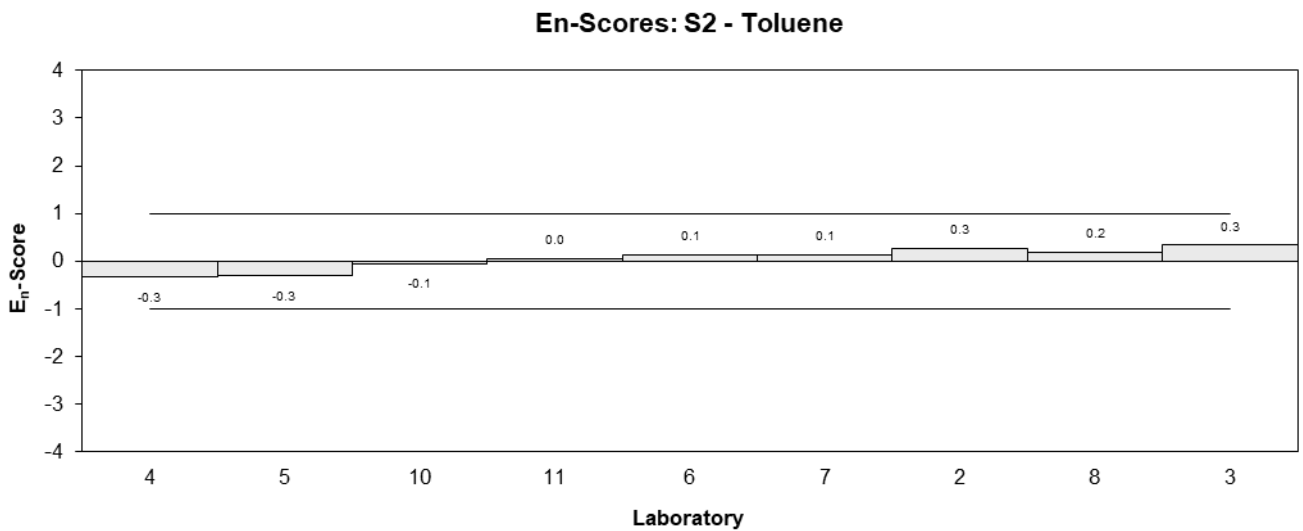
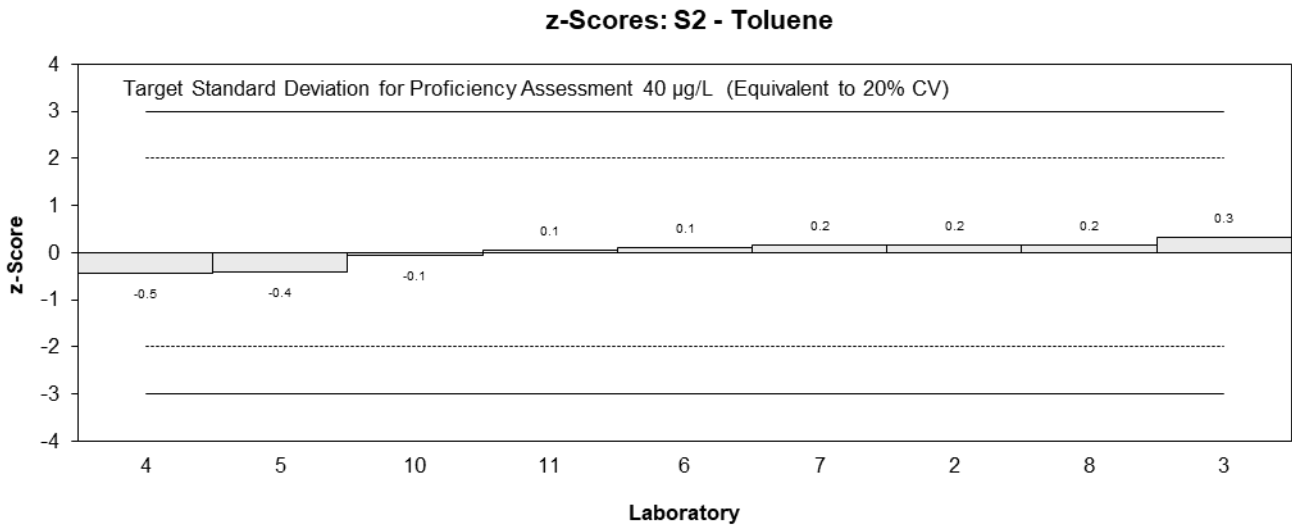
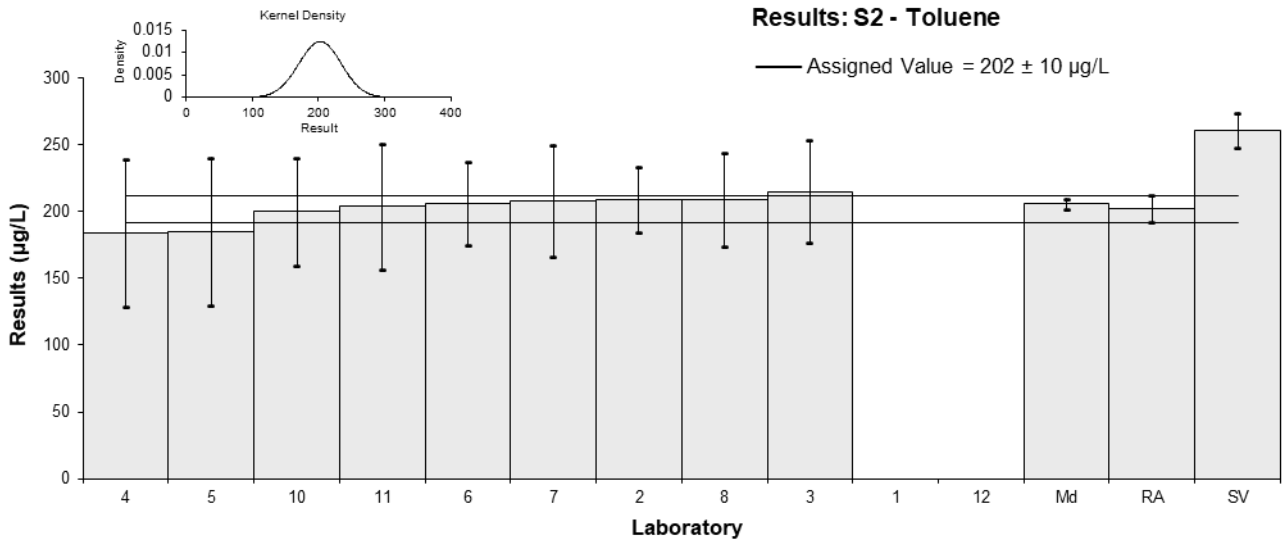


Figure 8

Table 13

Sample Details

Sample No.	S2
Matrix	Wastewater
Analyte	Ethylbenzene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty
1	NT	NT
2	<20	NR
3	<20	NR
4	22.48	7
5	15.5	4.65
6	19	3
7	21	3.7
8**	2	0.6
10	17	3.4
11	11.2	3.6
12	NT	NT

** Extreme Outlier, see Section 4.2

Statistics

Assigned Value	Not Set	
Spike Value	29.0	1.5
Robust Average	17.7	4.7
Median	18.0	4.2
Mean	17.7	
N	6	
Max	22.48	
Min	11.2	
Robust SD	4.6	
Robust CV	26%	

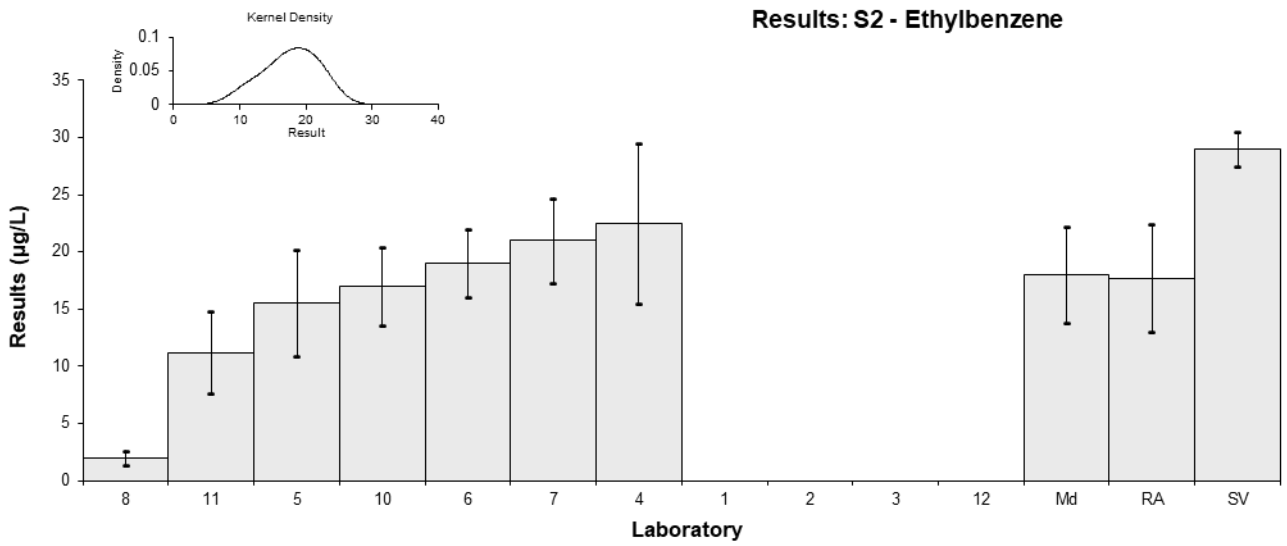


Figure 9

Table 14

Sample Details

Sample No.	S2
Matrix	Wastewater
Analyte	Xylenes
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	NT	NT		
2	98.5	12.805	0.60	0.69
3	94	16.9	0.34	0.32
4	87.99	26	0.00	0.00
5	54	16.2	-1.93	-1.88
6	91	NR	0.17	0.37
7	95	19	0.40	0.34
8	93	18	0.28	0.25
10	80	16	-0.45	-0.45
11	79	2.1	-0.51	-1.08
12	NT	NT		

Statistics

Assigned Value	88.0	8.1
Spike Value	129	6
Robust Average	88.0	8.1
Median	91.0	4.9
Mean	85.8	
N	9	
Max	98.5	
Min	54	
Robust SD	9.7	
Robust CV	11%	

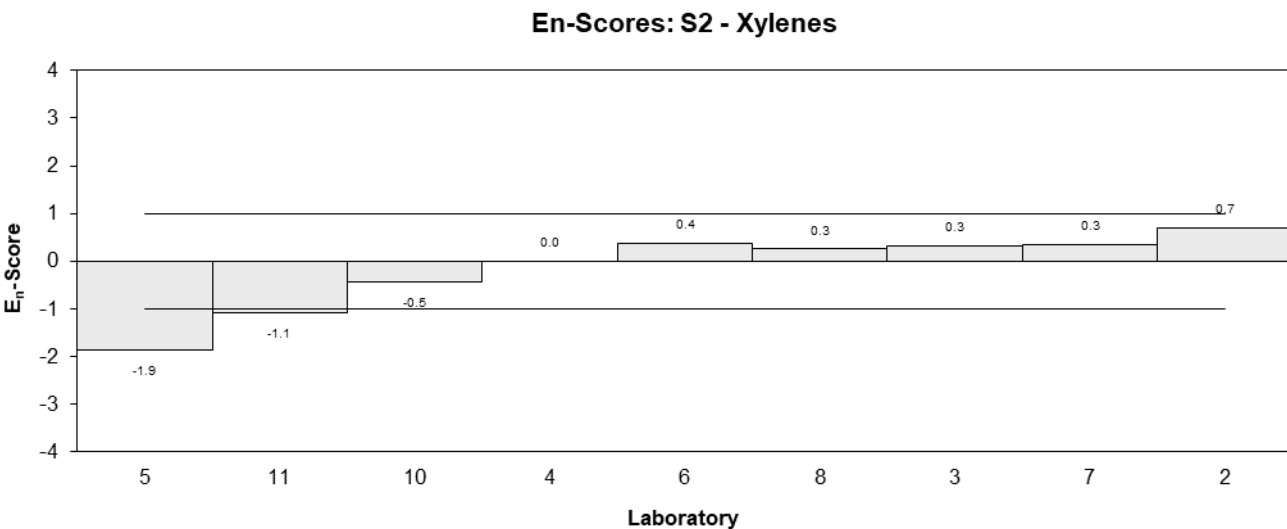
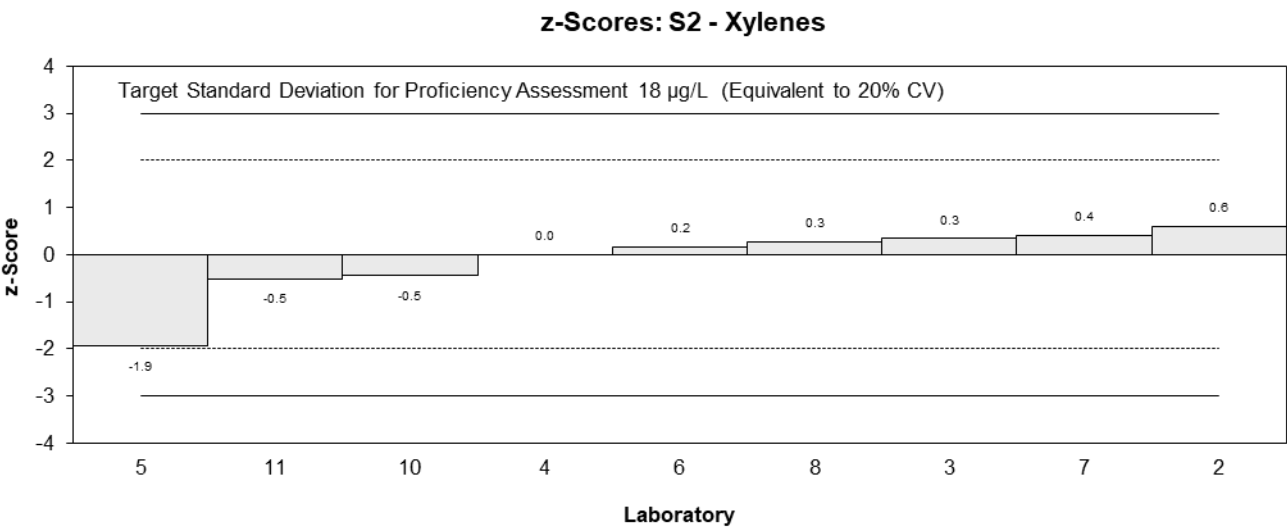
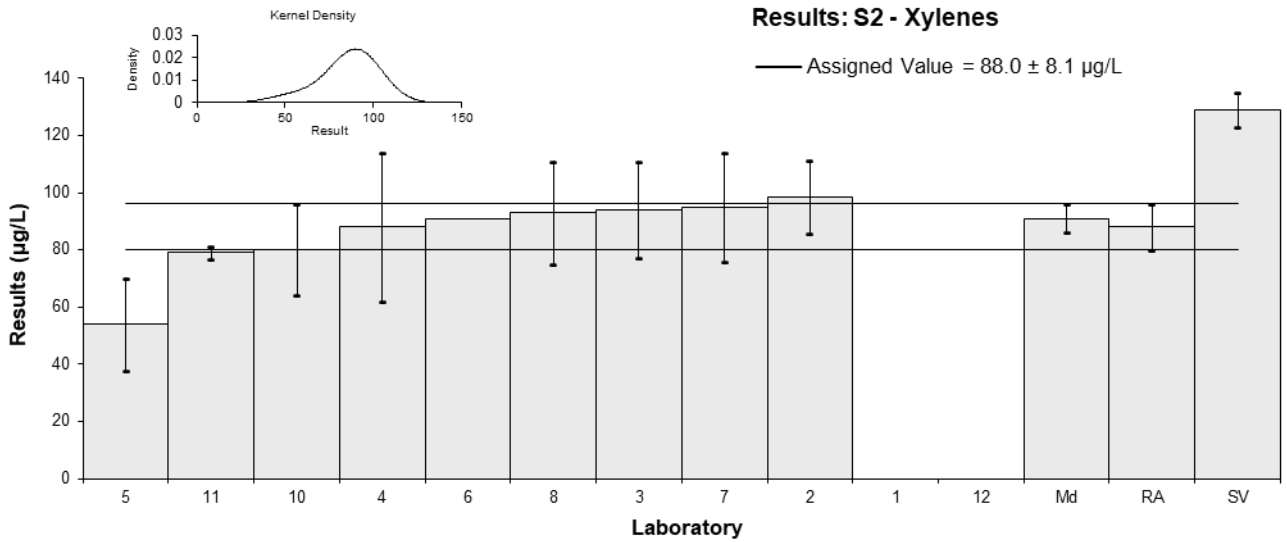


Figure 10

Table 15

Sample Details

Sample No.	S2
Matrix	Wastewater
Analyte	Total BTEX
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	NT	NT		
2	364	72.8	-0.08	-0.08
3	380	71.9	0.14	0.14
4	369.63	111	-0.01	0.00
5**	21.5	6.45	-4.71	-25.58
6	381	NR	0.15	0.92
7	387	77.4	0.23	0.22
8	368	70	-0.03	-0.03
10	360	72	-0.14	-0.14
11	352	100	-0.24	-0.18
12	NT	NT		

** Extreme Outlier, see Section 4.2

Statistics

Assigned Value	370	12
Spike Value	489	24
Robust Average	370	12
Median	369	13
Mean	370	
N	8	
Max	387	
Min	352	
Robust SD	13	
Robust CV	3.6%	

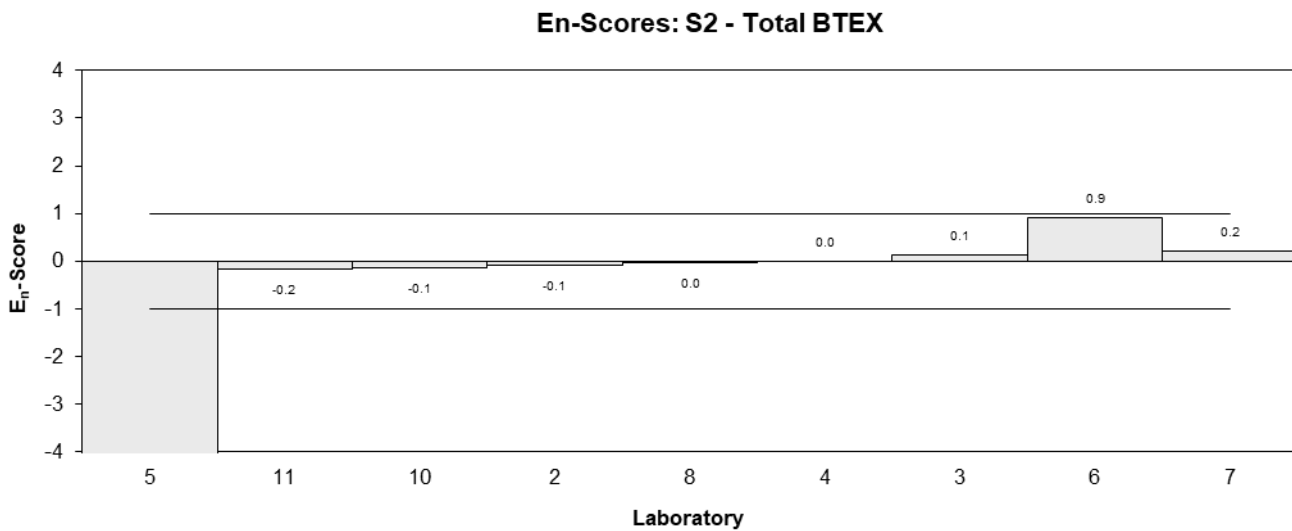
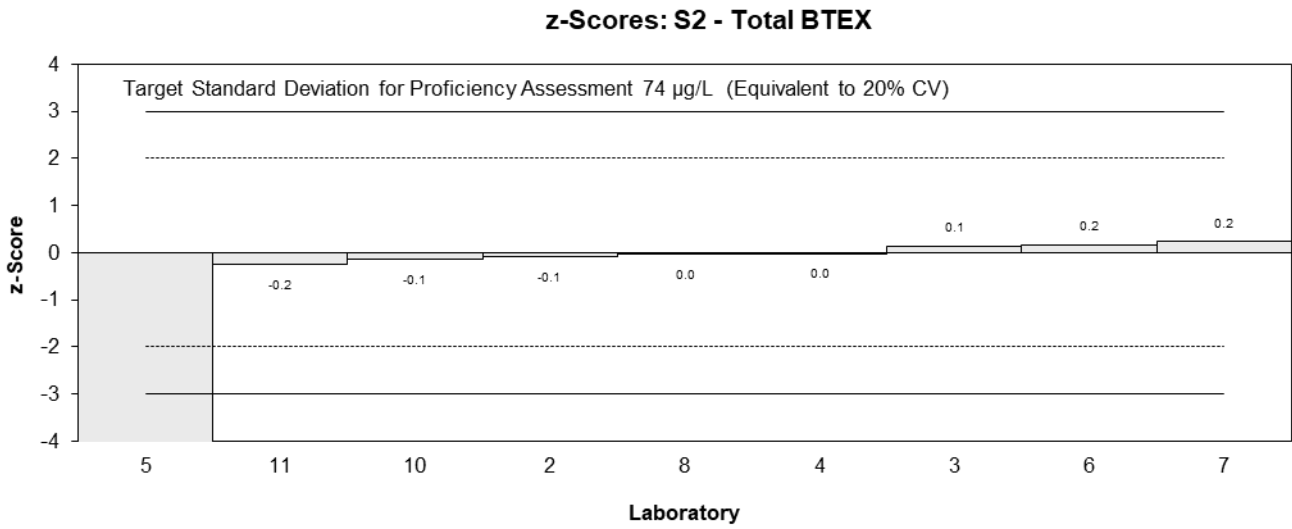
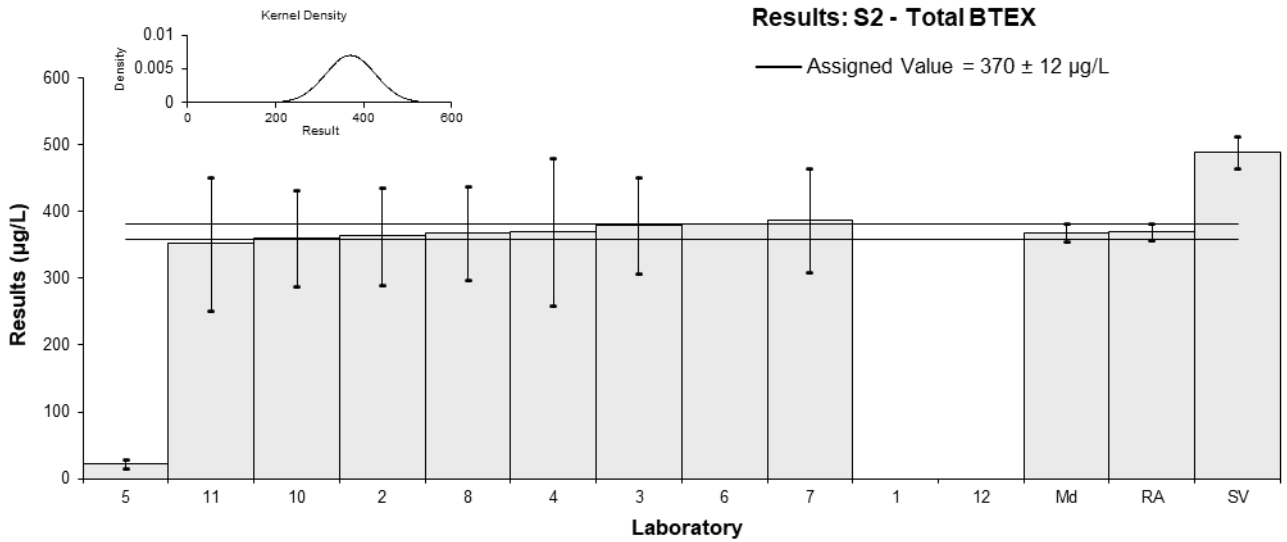


Figure 11

Table 16

Sample Details

Sample No.	S3
Matrix	Wastewater
Analyte	Acenaphthene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	4	1	-0.24	-0.17
2	3.31	0.8606	-1.06	-0.86
3	3.6	0.94	-0.71	-0.55
4	5.07	2.03	1.04	0.41
5	5.1	1.53	1.07	0.55
6	3.6	0.7	-0.71	-0.66
7	4.3	1.6	0.12	0.06
8	3.9	0.6	-0.36	-0.36
10	4.8	1.9	0.71	0.30
11	4.3	1.3	0.12	0.07
12*	6.67	0.1	2.00▼	

* Outlier, see Section 4.2; ▼ Adjusted Score, see Section 6.3

Statistics

Assigned Value	4.20	0.57
Spike Value	8.08	0.40
Robust Average	4.32	0.62
Max Acceptable Result	11.3	
Median	4.30	0.78
Mean	4.42	
N	11	
Max	6.67	
Min	3.31	
Robust SD	0.82	
Robust CV	19%	

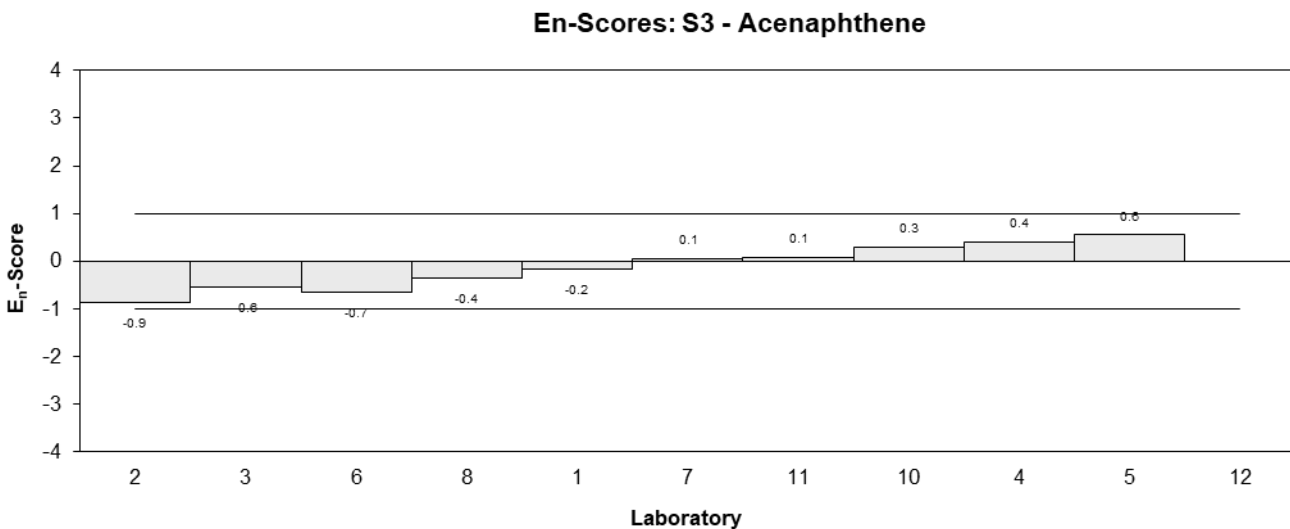
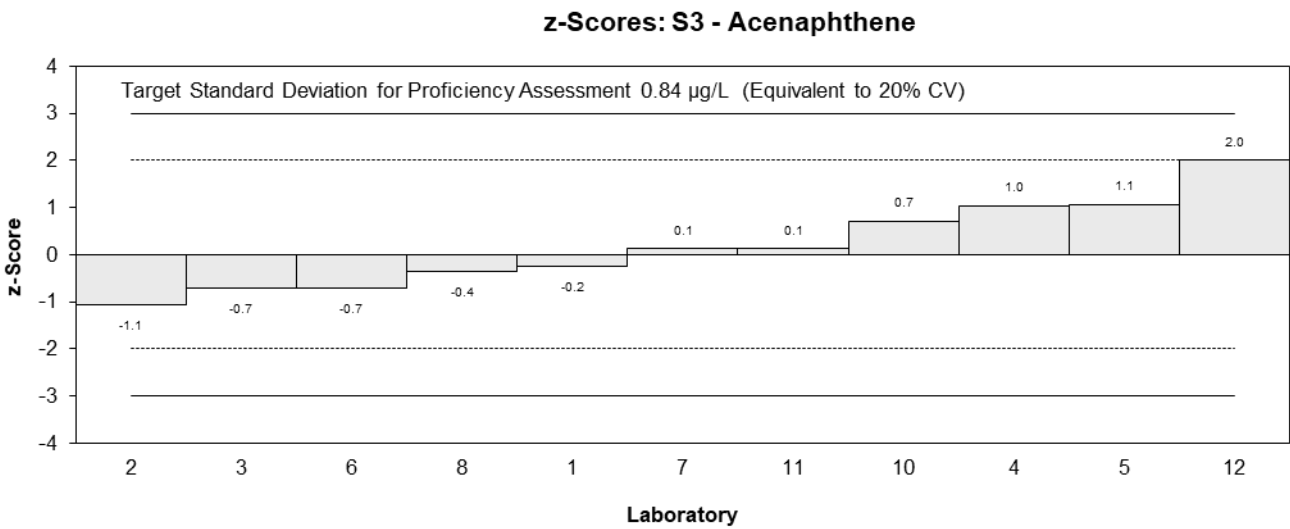
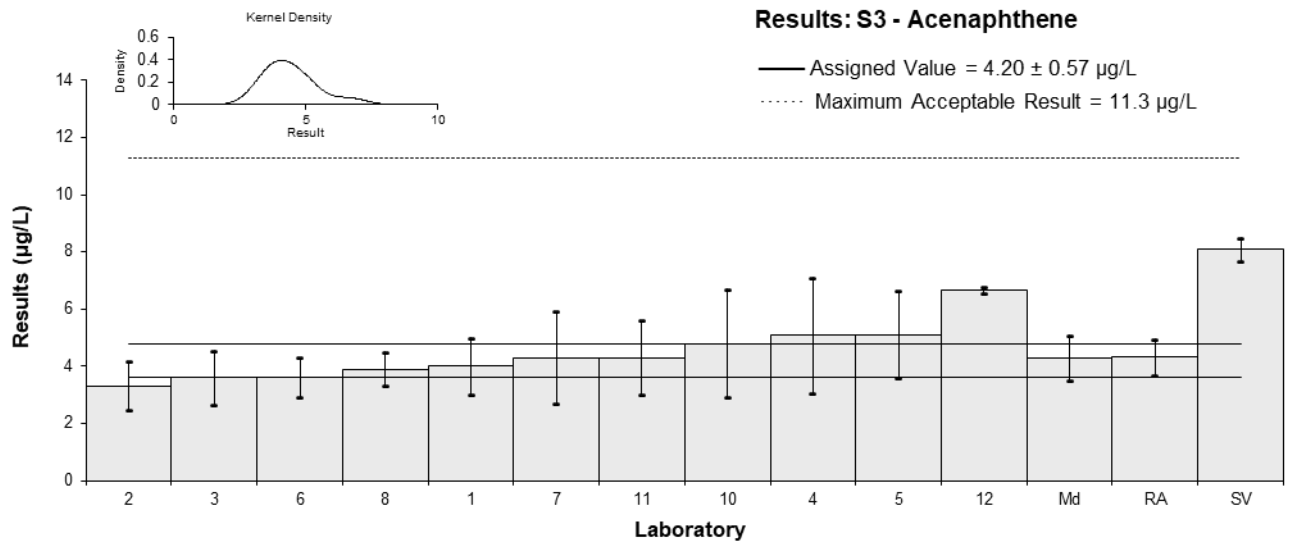


Figure 12

Table 17

Sample Details

Sample No.	S3
Matrix	Wastewater
Analyte	Acenaphthylene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty
1	<1	NR
2	<1	NR
3	<1	NR
4	0.68	0.27
5	0.8	0.24
6	<1.0	NR
7	<1	0.4
8	<1.0	NR
10	0.73	0.29
11	1.66	0.43
12	0.92	0.05

Statistics

Assigned Value	Not Set	
Spike Value	1.09	0.05
Robust Average	NA (N<6)	
Median	0.80	0.20
Mean	0.96	
N	5	
Max	1.66	
Min	0.68	
Robust SD	NA (N<6)	
Robust CV	NA (N<6)	

Results: S3 - Acenaphthylene

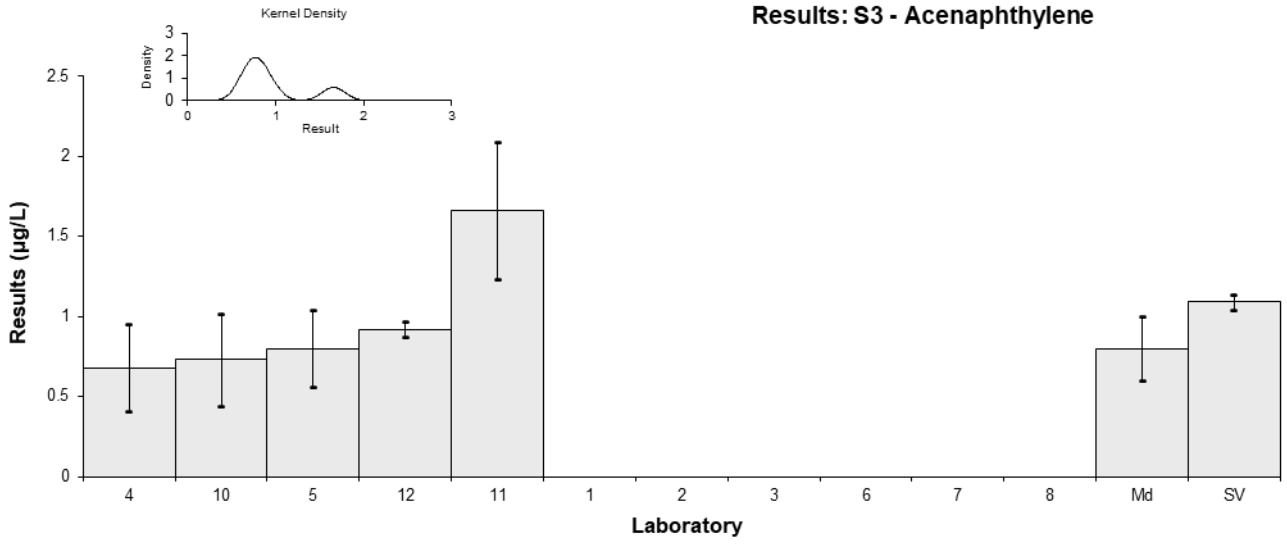


Figure 13

Table 18

Sample Details

Sample No.	S3
Matrix	Wastewater
Analyte	Benz[a]anthracene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	5	2	0.79	0.32
2	5.22	1.284	1.04	0.60
3	4.5	1.1	0.21	0.13
4	3.25	1.30	-1.24	-0.71
5	3.6	1.08	-0.83	-0.54
6	4.5	0.9	0.21	0.15
7	3.9	1.45	-0.49	-0.26
8	4.3	1.2	-0.02	-0.01
10	6.8	2.7	2.00▼	
11	3.14	0.9	-1.37	-0.99
12*	7.67	0.1	2.00▼	

* Outlier, see Section 4.2; ▼ Adjusted Score, see Section 6.3

Statistics

Assigned Value	4.32	0.78
Spike Value	6.50	0.32
Robust Average	4.6	1.0
Max Acceptable Result	9.09	
Median	4.50	0.80
Mean	4.72	
N	11	
Max	7.67	
Min	3.14	
Robust SD	1.4	
Robust CV	30%	

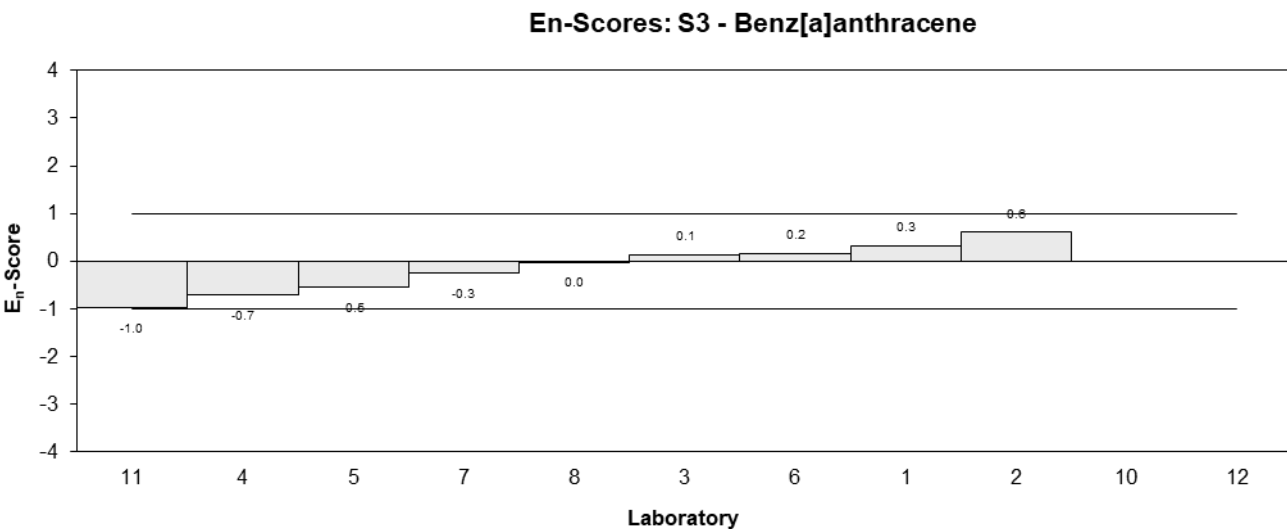
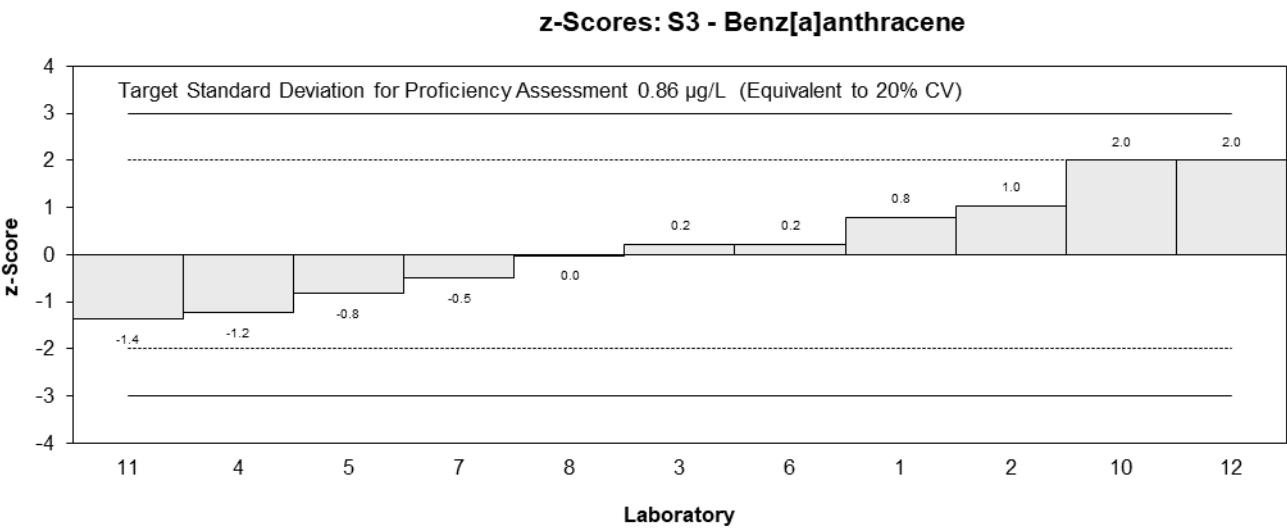
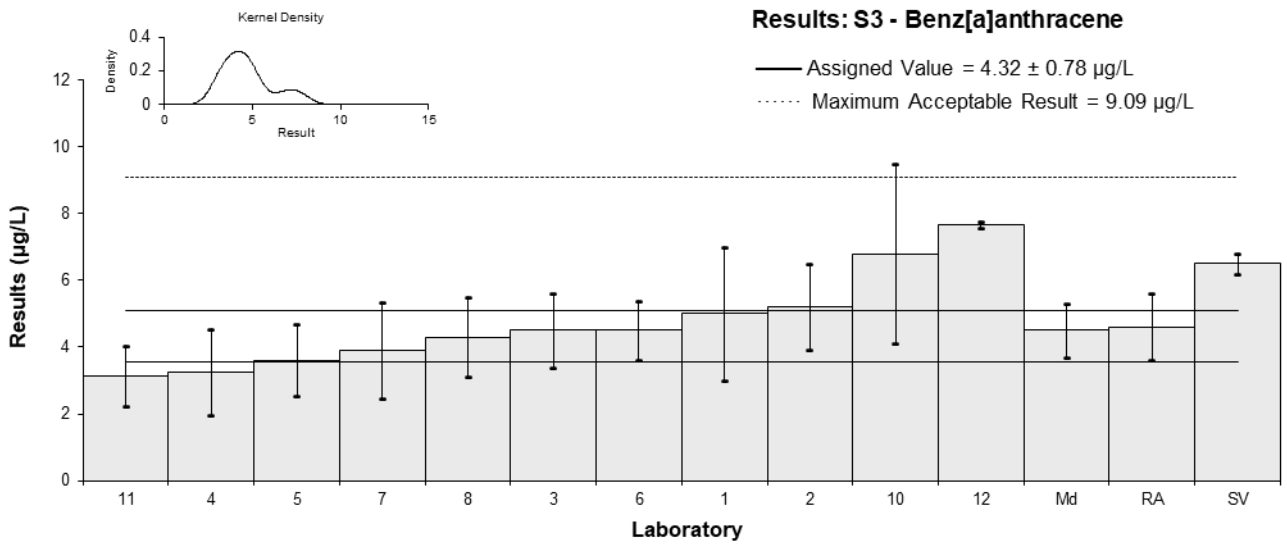


Figure 14

Table 19

Sample Details

Sample No.	S3
Matrix	Wastewater
Analyte	Benzo[a]pyrene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E _n
1*	4	1	2.00 ▼	
2	2.22	0.5328	0.58	0.30
3	1.8	0.43	-0.48	-0.28
4	1.88	0.75	-0.28	-0.12
5	1.2	0.36	-1.98	-1.22
6	2.6	0.5	1.53	0.83
7	2.8	1.1	2.00 ▼	
8	1.9	0.4	-0.23	-0.13
10	1.5	0.60	-1.23	-0.61
11*	0.76	0.24	-3.09	-2.08
12*	4.00	0.1	2.00 ▼	

* Outlier, see Section 4.2; ▼ Adjusted Score, see Section 6.3

Statistics

Assigned Value	1.99	0.54
Spike Value	3.62	0.18
Robust Average	2.24	0.89
Max Acceptable Result	5.06	
Median	1.90	0.78
Mean	2.24	
N	11	
Max	4	
Min	0.76	
Robust SD	1.2	
Robust CV	53%	

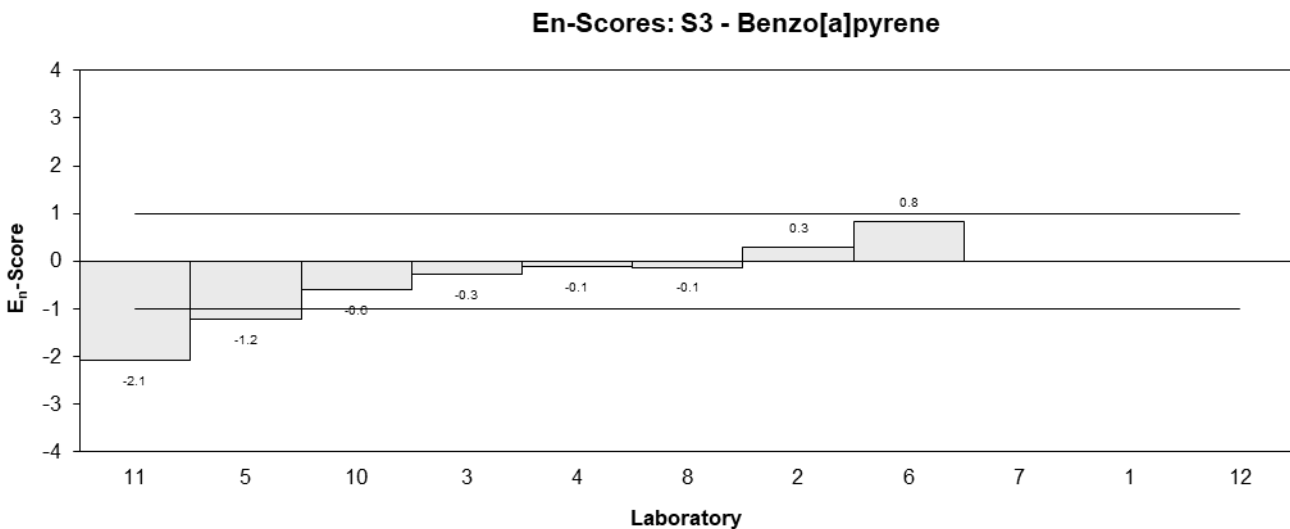
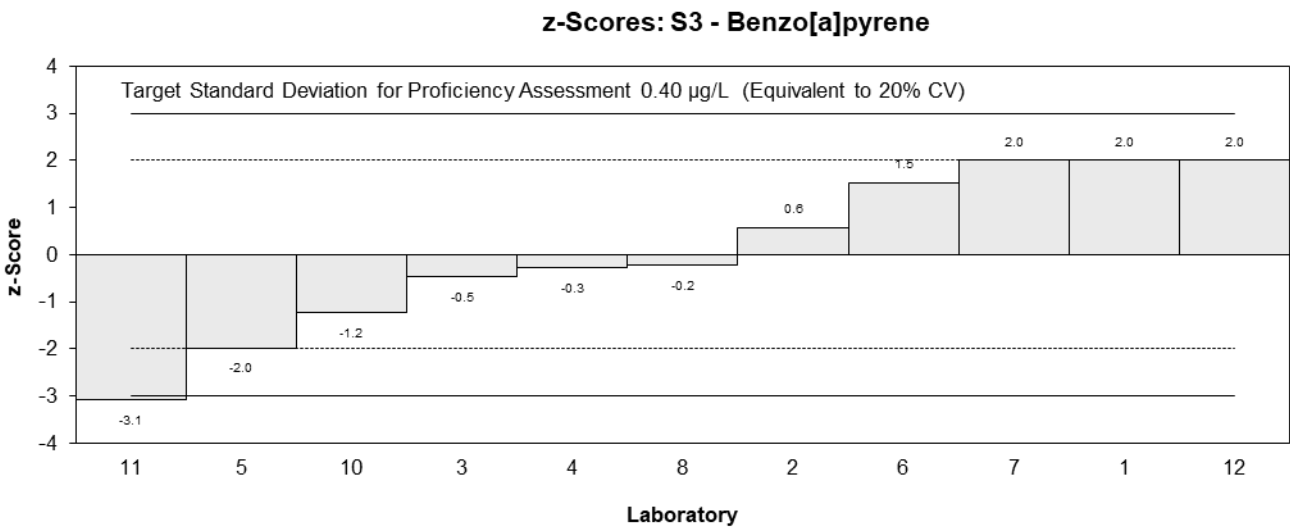
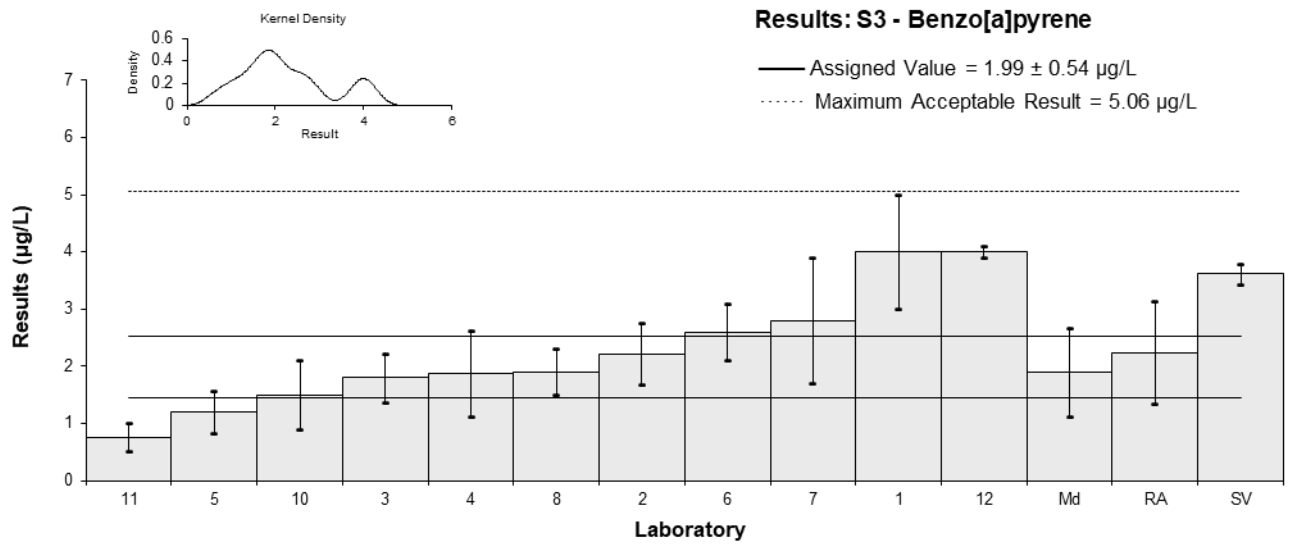


Figure 15

Table 20

Sample Details

Sample No.	S3
Matrix	Wastewater
Analyte	Fluorene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	2	1	-0.41	-0.17
2	1.8	0.45	-0.87	-0.71
3	2.0	0.49	-0.41	-0.32
4	2.32	0.93	0.32	0.14
5	<0.02	NR		
6	1.8	0.3	-0.87	-0.91
7	2.1	0.81	-0.18	-0.09
8	2.0	0.3	-0.41	-0.43
10	2.6	1.0	0.96	0.40
11	2.42	0.56	0.55	0.38
12	3.19	0.1	2.00▼	

▼ Adjusted Score, see Section 6.3

Statistics

Assigned Value	2.18	0.29
Spike Value	3.10	0.15
Robust Average	2.18	0.29
Max Acceptable Result	4.34	
Median	2.05	0.29
Mean	2.22	
N	10	
Max	3.19	
Min	1.8	
Robust SD	0.37	
Robust CV	17%	

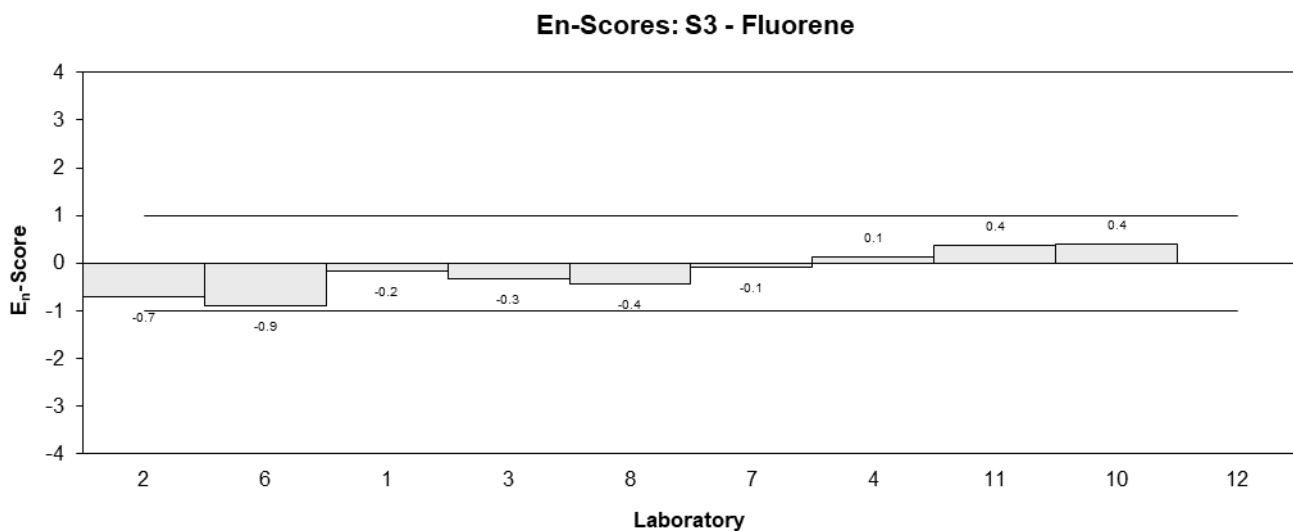
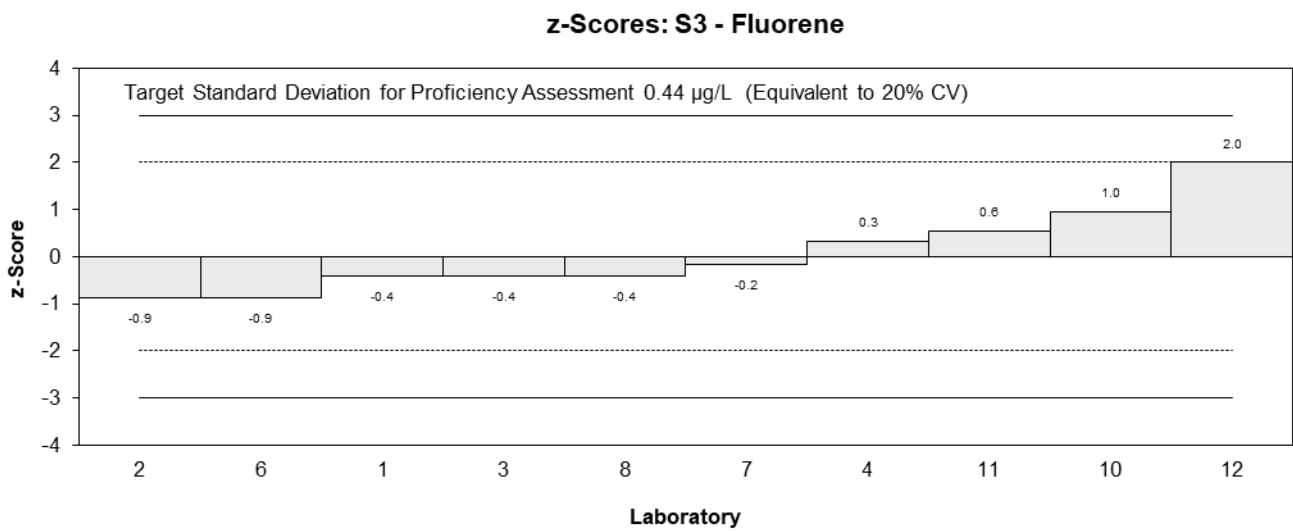
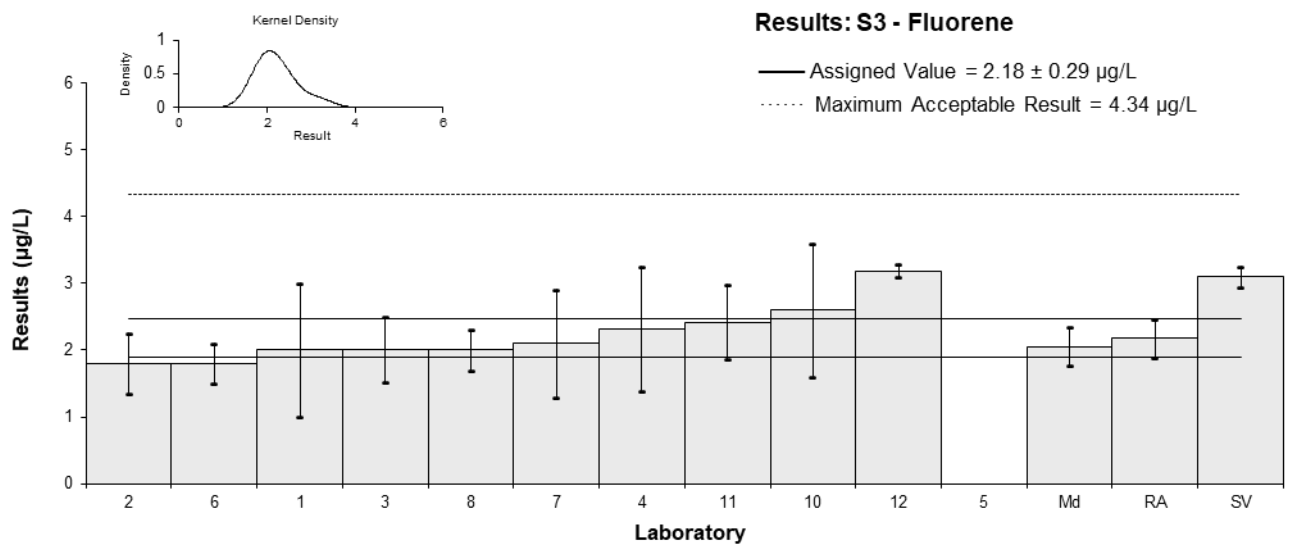


Figure 16

Table 21

Sample Details

Sample No.	S3
Matrix	Wastewater
Analyte	Phenanthrene
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	2	1	0.62	0.21
2	1.8	0.432	0.06	0.04
3	1.4	0.33	-1.07	-0.77
4	1.75	0.70	-0.08	-0.04
5	2.3	0.69	1.46	0.66
6	1.3	0.3	-1.35	-1.01
7	1.2	0.41	-1.63	-1.05
8	1.3	0.2	-1.35	-1.14
10	1.9	0.76	0.34	0.14
11	2.41	0.48	1.77	1.04
12	2.23	0.1	1.26	1.17

Statistics

Assigned Value	1.78	0.37
Spike Value	1.95	0.10
Robust Average	1.78	0.37
Median	1.80	0.48
Mean	1.78	
N	11	
Max	2.41	
Min	1.2	
Robust SD	0.49	
Robust CV	28%	

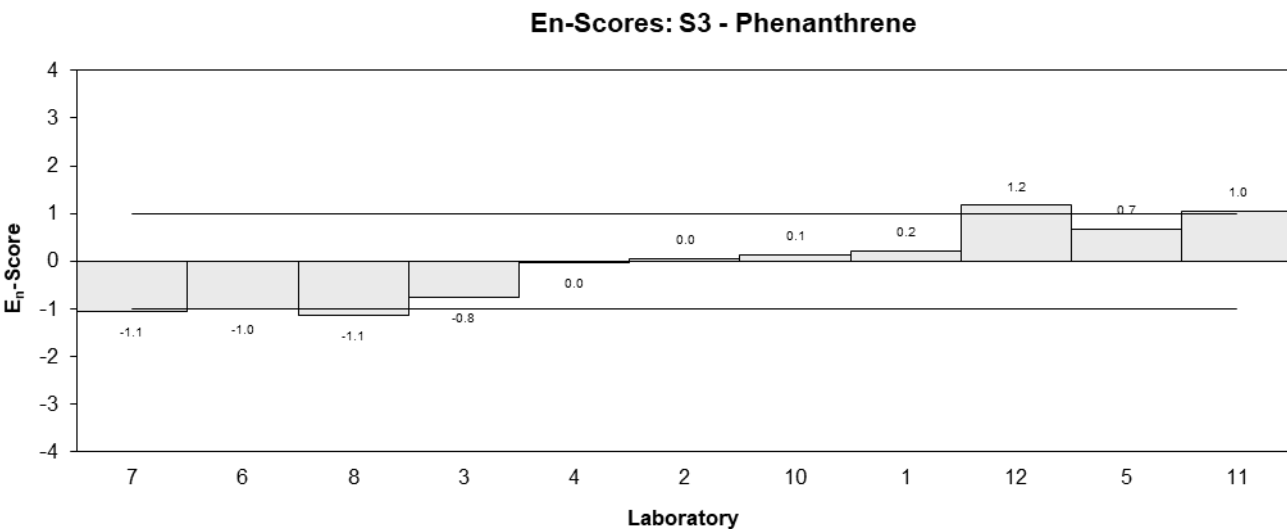
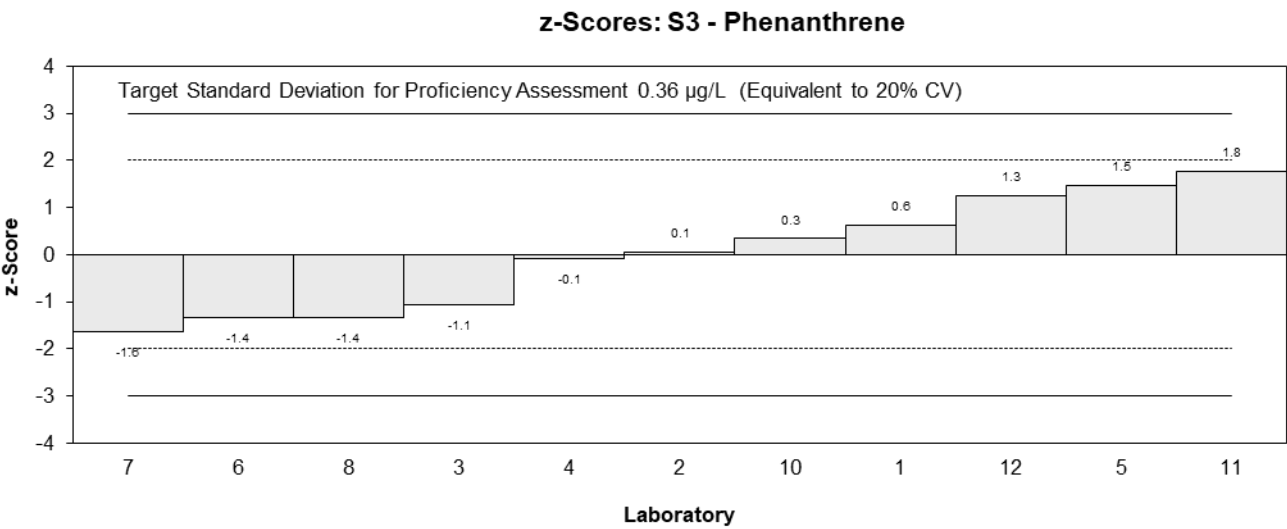
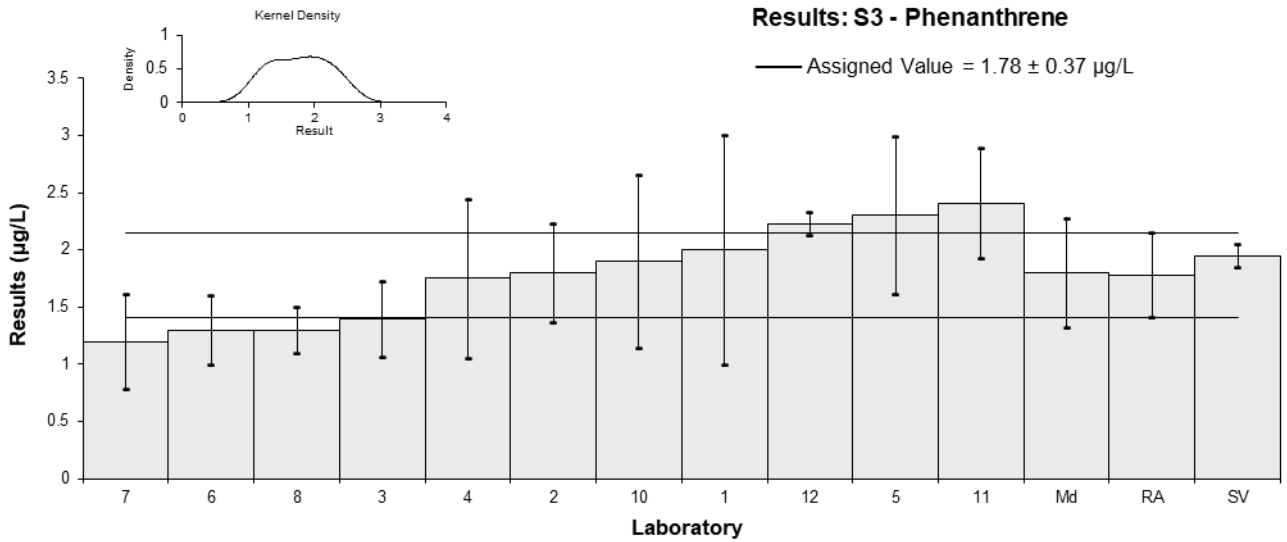


Figure 17

Table 22

Sample Details

Sample No.	S4
Matrix	Wastewater
Analyte	Atrazine
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	8.6	2.6	-0.74	-0.52
2	14.1	2.88	1.98	1.27
3	9.4	2.1	-0.35	-0.28
4	9.49	3.80	-0.30	-0.15
5	9.6	2.88	-0.25	-0.16
6	8	1.3	-1.04	-1.14
7	10	3.6	-0.05	-0.03
8	NT	NT		
10	10	4.0	-0.05	-0.02
11	10.45	2.3	0.17	0.13
12	12.55	0.6	1.21	1.71

Statistics

Assigned Value	10.1	1.3
Spike Value	11.6	0.6
Robust Average	10.1	1.3
Median	9.80	0.62
Mean	10.2	
N	10	
Max	14.1	
Min	8	
Robust SD	1.7	
Robust CV	17%	

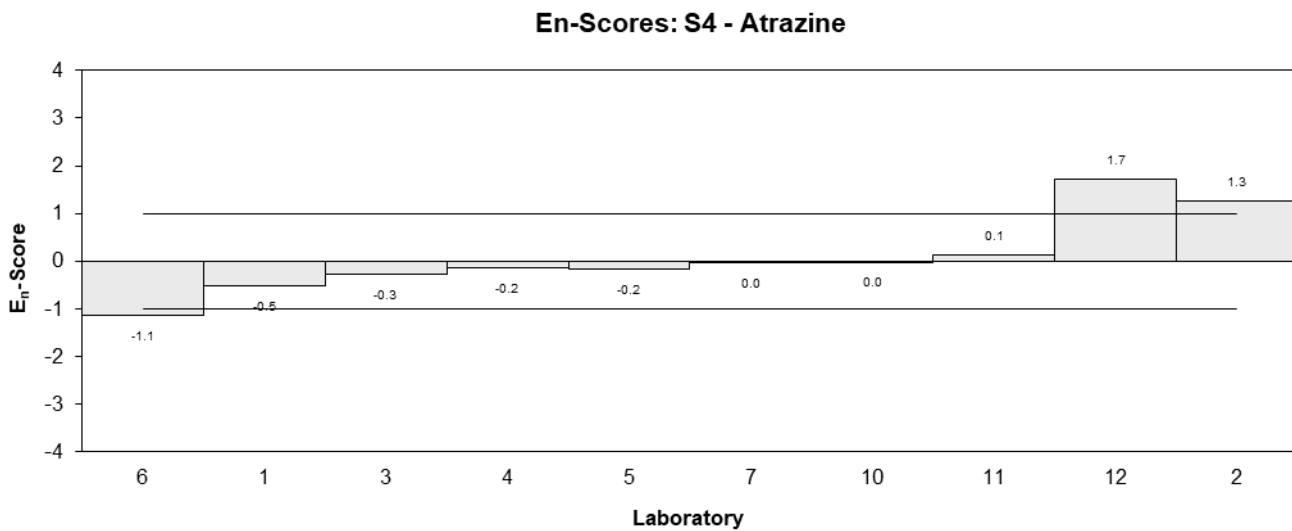
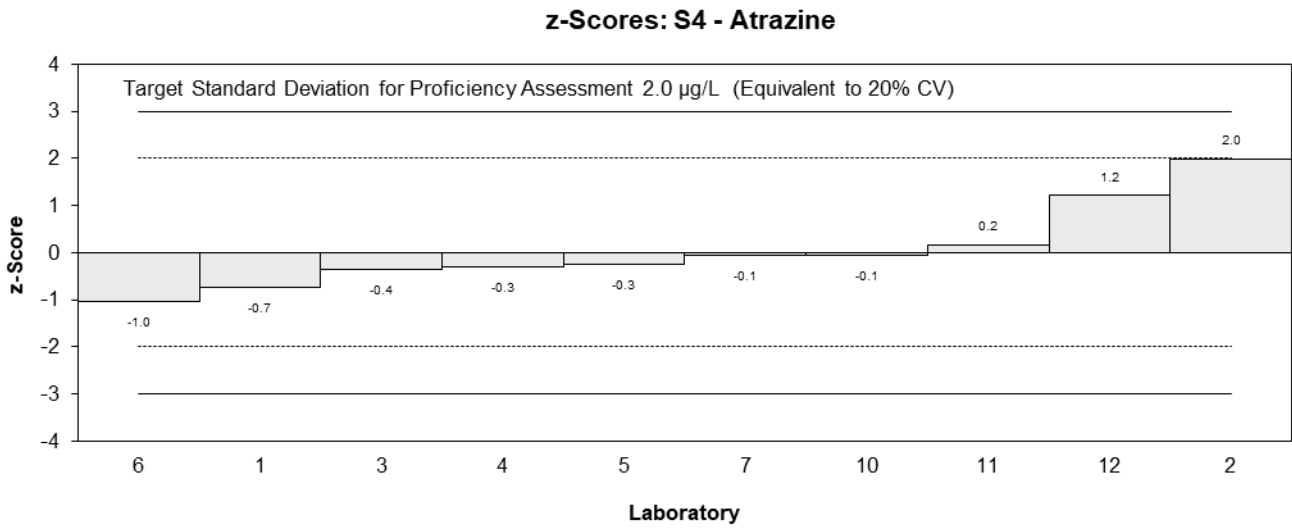
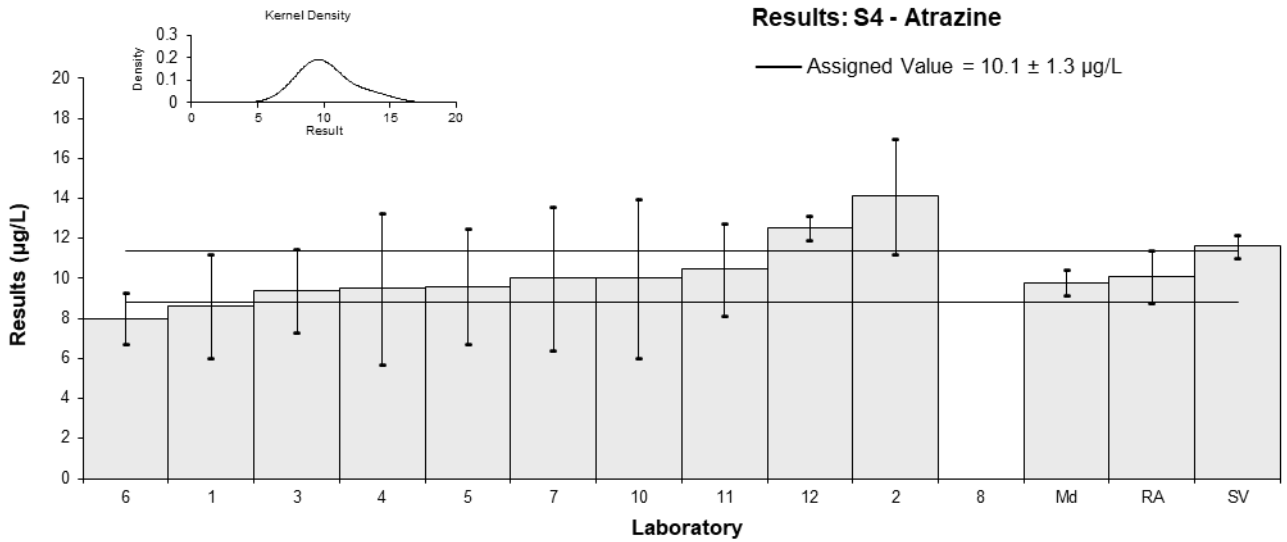


Figure 18

Table 23

Sample Details

Sample No.	S4
Matrix	Wastewater
Analyte	Chlorpyrifos
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	14	4	-0.27	-0.19
2	14.8	1.5836	0.00	0.00
3	12.5	3.54	-0.78	-0.59
4	15.34	6.13	0.18	0.09
5	17	5.1	0.74	0.41
6	11.9	1.3	-0.98	-1.41
7	12.6	4.5	-0.74	-0.46
8	15.4	3.8	0.20	0.15
10	15	6.0	0.07	0.03
11	15.8	3.8	0.34	0.24
12	18.00	0.9	1.08	1.74

Statistics

Assigned Value	14.8	1.6
Spike Value	20.1	1.0
Robust Average	14.8	1.6
Median	15.0	1.1
Mean	14.8	
N	11	
Max	18	
Min	11.9	
Robust SD	2.1	
Robust CV	15%	

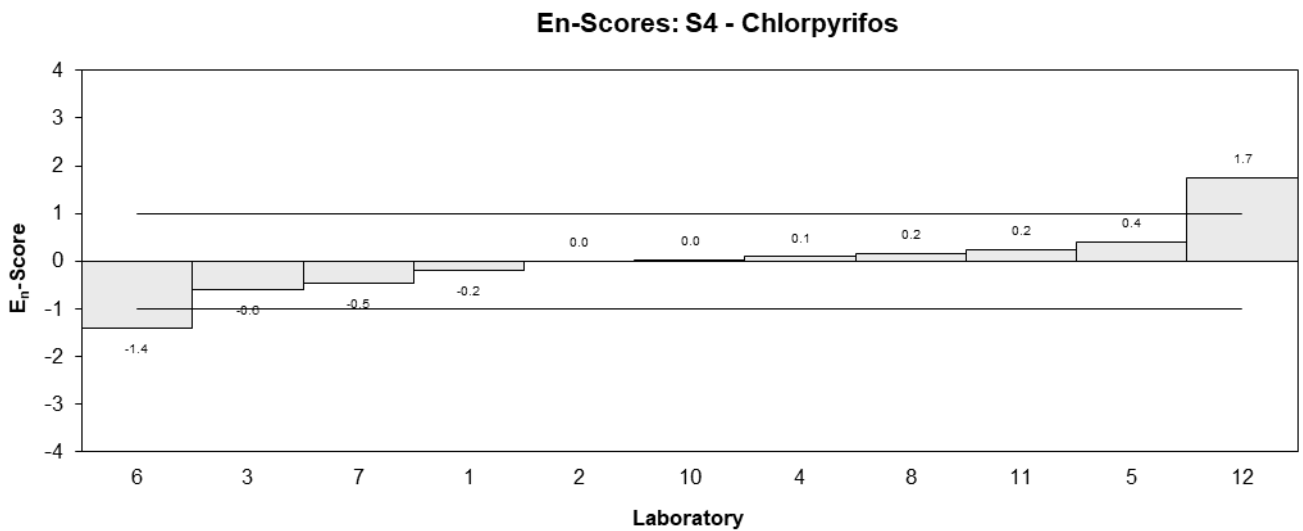
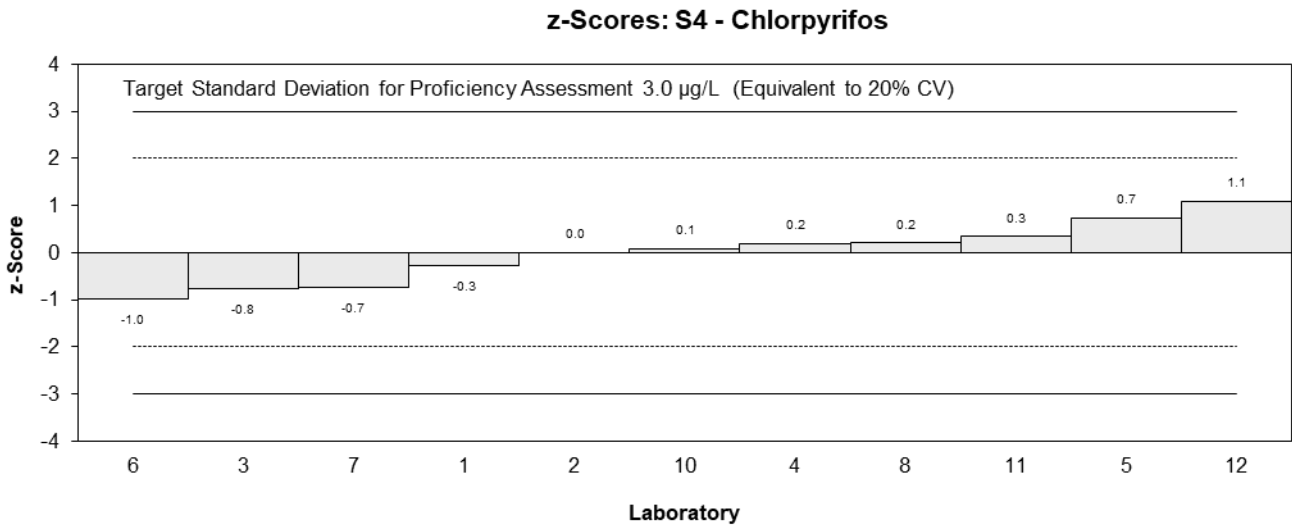
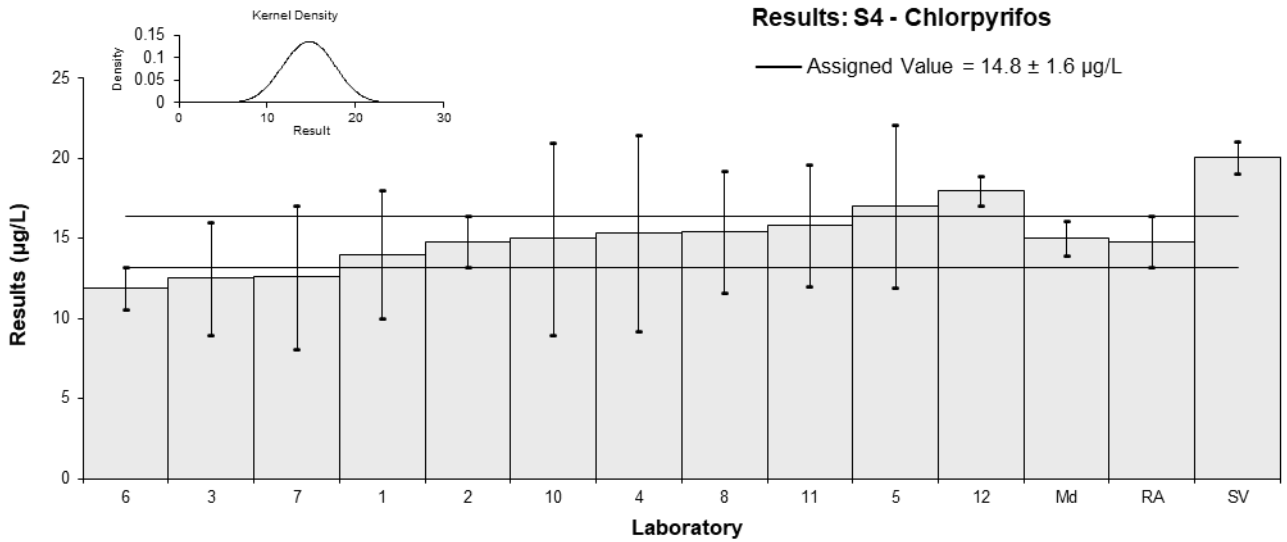


Figure 19

Table 24

Sample Details

Sample No.	S4
Matrix	Wastewater
Analyte	Dicamba
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty
1	NT	NT
2	7.31	0.365
3	NT	NT
4	NT	NT
5	NT	NT
6	<10	NR
7	NT	NT
8	NT	NT
10	6.5	1.6
11	4.66	1.4
12	<0.05	0.05

Statistics

Assigned Value	Not Set	
Spike Value	7.49	0.37
Robust Average	NA (N<6)	
Median	6.5	1.7
Mean	6.2	
N	3	
Max	7.31	
Min	4.66	
Robust SD	NA (N<6)	
Robust CV	NA (N<6)	

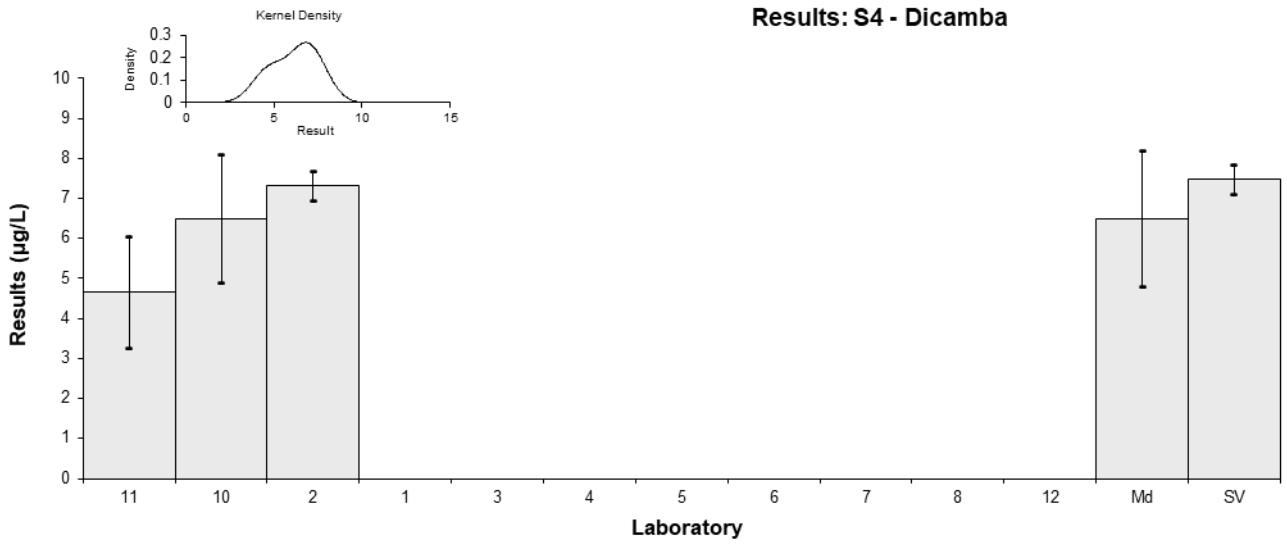


Figure 20

Table 25

Sample Details

Sample No.	S4
Matrix	Wastewater
Analyte	Dieldrin
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	3.9	1.2	0.03	0.02
2	3.82	1.23	-0.08	-0.05
3	3.0	0.71	-1.13	-1.02
4	4.49	1.79	0.79	0.33
5	3	0.9	-1.13	-0.86
6	3.6	0.45	-0.36	-0.42
7	3.6	1.4	-0.36	-0.19
8	4.3	1.0	0.54	0.38
10	4.1	1.6	0.28	0.13
11	5.73	1.2	2.00 ▼	
12	4.04	0.2	0.21	0.30

▼ Adjusted Score, see Section 6.3

Statistics

Assigned Value	3.88	0.49
Spike Value	5.01	0.25
Robust Average	3.88	0.49
Max Acceptable Result	7.01	
Median	3.90	0.34
Mean	3.96	
N	11	
Max	5.73	
Min	3	
Robust SD	0.65	
Robust CV	17%	

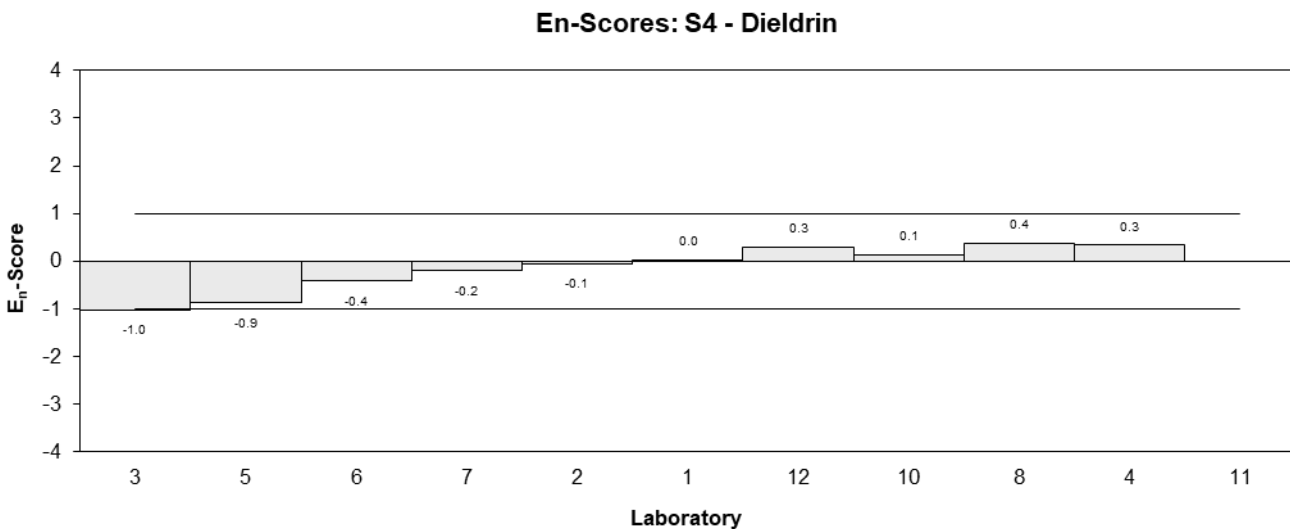
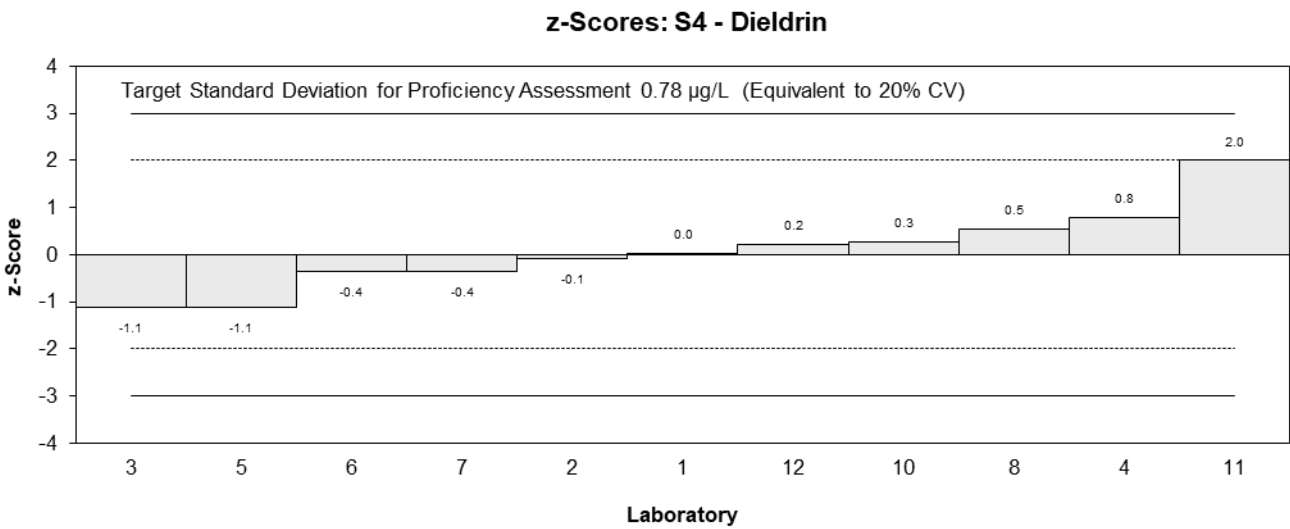
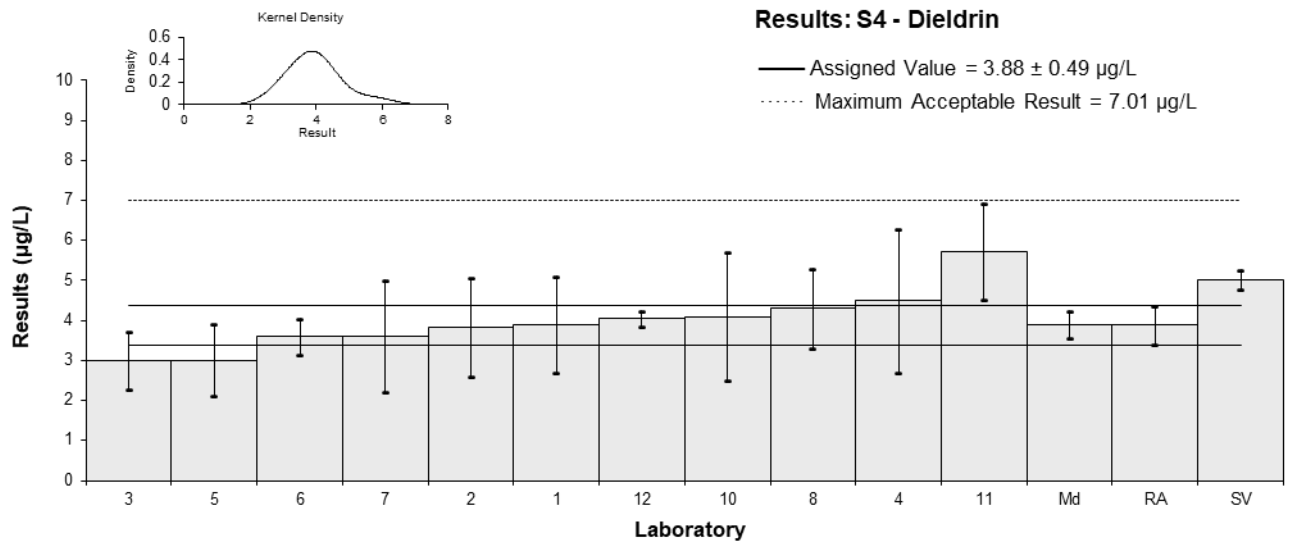


Figure 21

Table 26

Sample Details

Sample No.	S4
Matrix	Wastewater
Analyte	Ethion
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	2.6	0.8	0.37	0.21
2	2.78	0.6116	0.74	0.53
3	2.4	0.69	-0.04	-0.03
4	1.7183	0.68	-1.45	-0.94
5	NT	NT		
6	2.3	0.3	-0.25	-0.28
7	2.2	0.91	-0.45	-0.23
8	2.4	0.8	-0.04	-0.02
10	2.8	1.1	0.79	0.33
11	NT	NT		
12*	3.75	0.2	2.00▼	

* Outlier, see Section 4.2; ▼ Adjusted Score, see Section 6.3

Statistics

Assigned Value	2.42	0.30
Spike Value	3.99	0.20
Robust Average	2.50	0.36
Max Acceptable Result	5.58	
Median	2.40	0.25
Mean	2.55	
N	9	
Max	3.75	
Min	1.7183	
Robust SD	0.44	
Robust CV	17%	

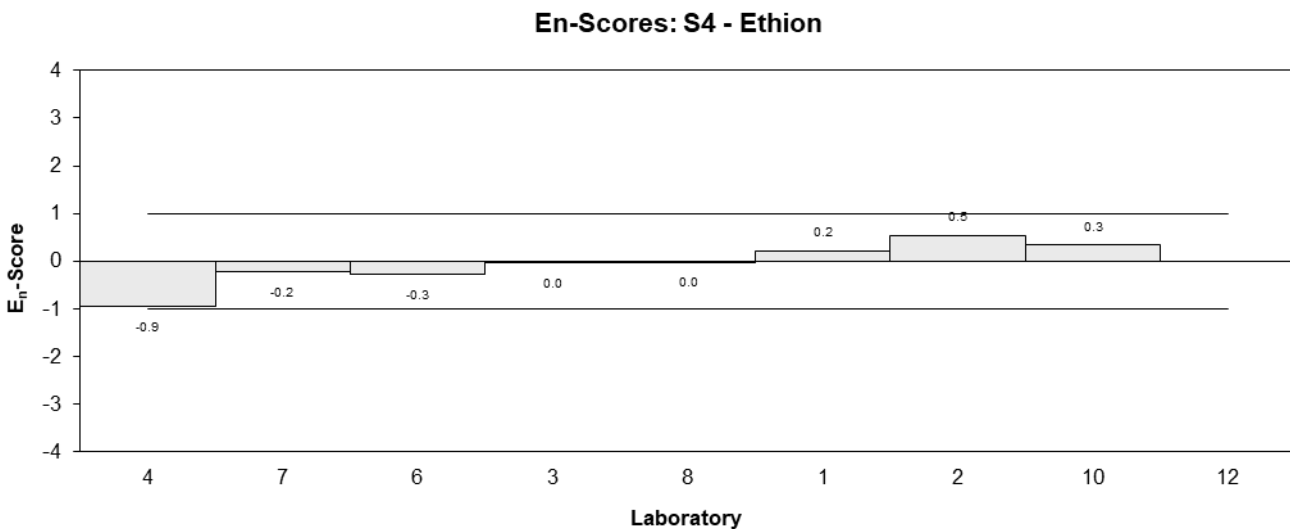
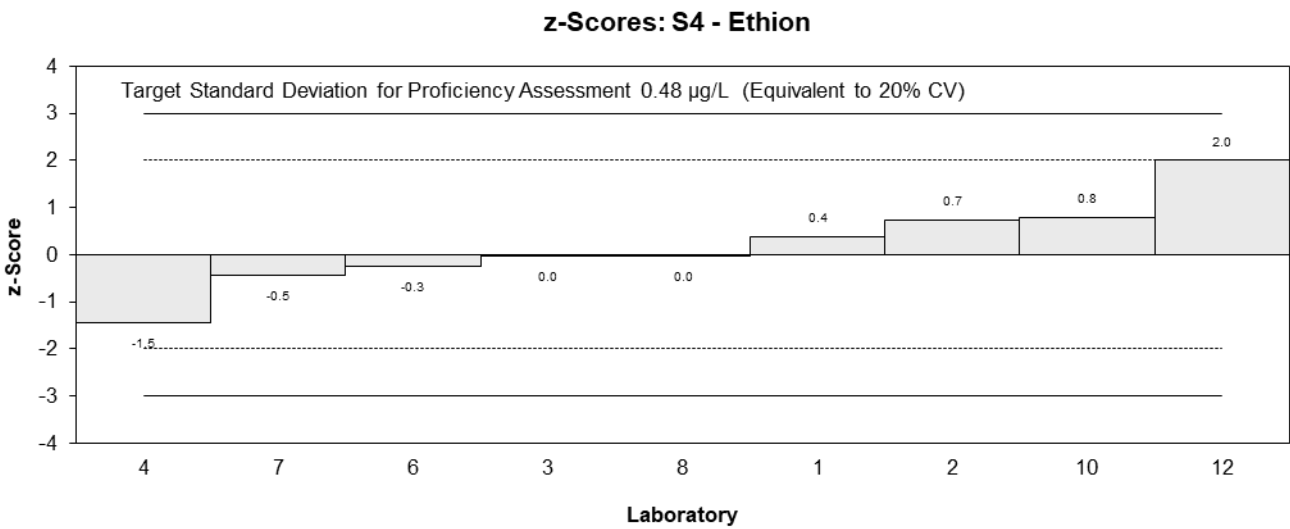
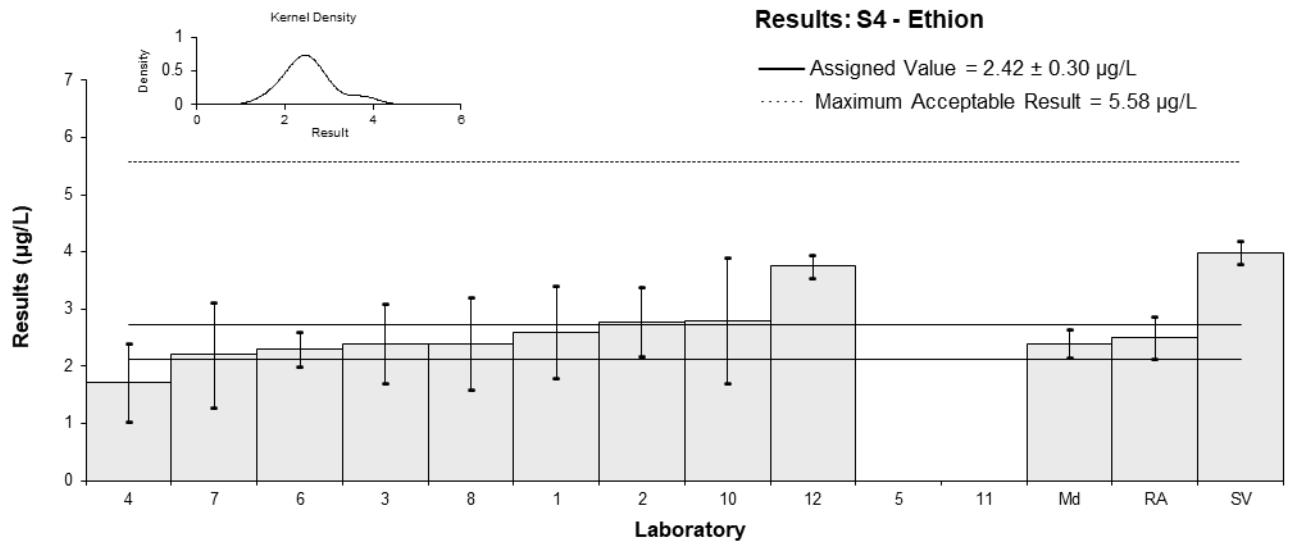


Figure 22

Table 27

Sample Details

Sample No.	S4
Matrix	Wastewater
Analyte	Lindane
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	7.3	2.2	-0.60	-0.41
2	7.48	2.44	-0.49	-0.31
3	7.6	2	-0.42	-0.31
4	8.96	3.59	0.40	0.18
5	7	2.1	-0.78	-0.55
6	7.5	NR	-0.48	-0.73
7	7.7	3.5	-0.36	-0.16
8	9.4	2.3	0.66	0.43
10*	3.7	1.5	-2.77	-2.47
11	10.02	2.6	1.04	0.61
12	10.57	0.5	1.37	1.88

* Outlier, see Section 4.2

Statistics

Assigned Value	8.3	1.1
Spike Value	10.0	0.5
Robust Average	8.1	1.2
Median	7.60	0.67
Mean	7.9	
N	11	
Max	10.57	
Min	3.7	
Robust SD	1.7	
Robust CV	20%	

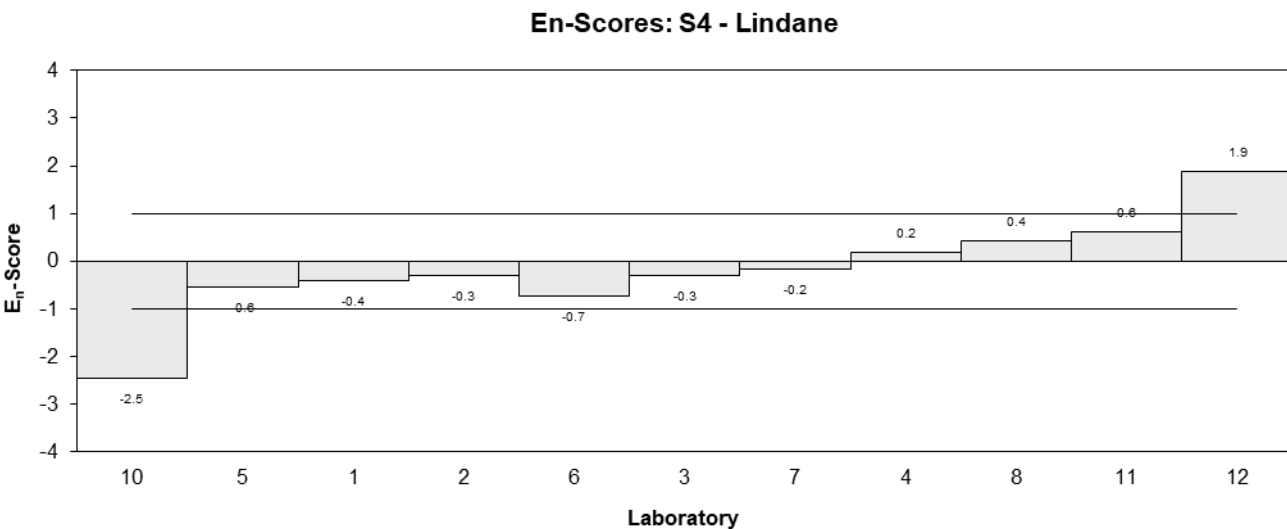
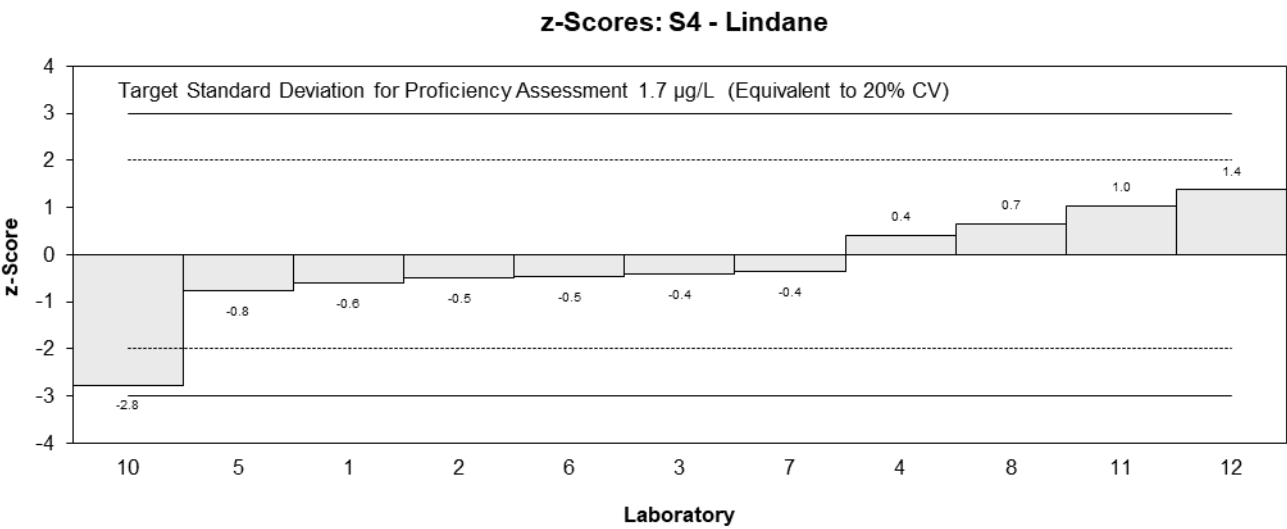
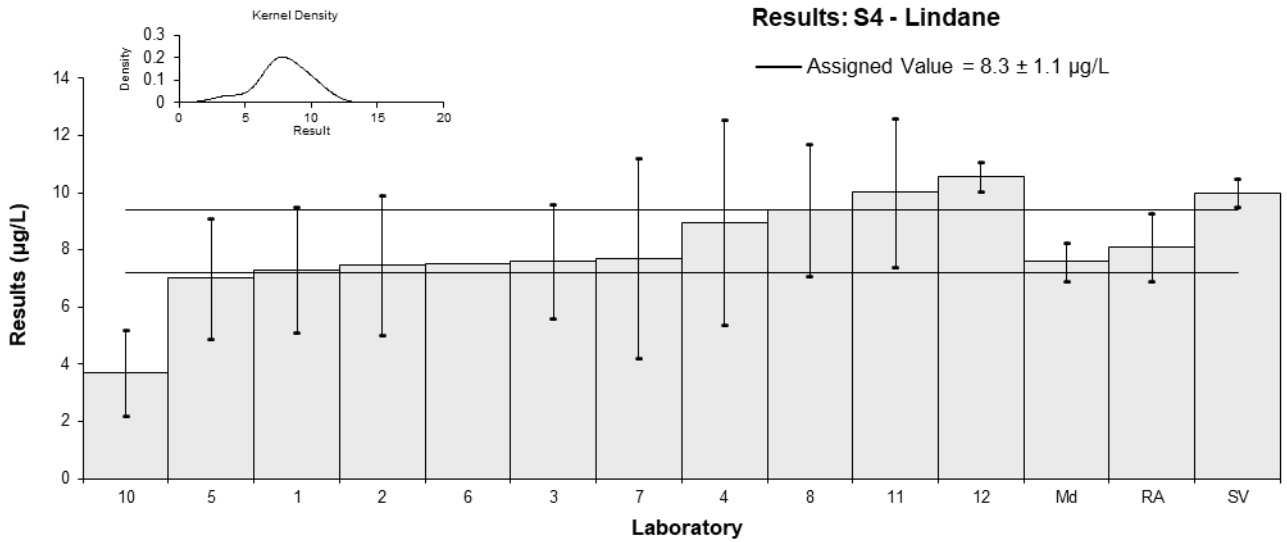


Figure 23

Table 28

Sample Details

Sample No.	S4
Matrix	Wastewater
Analyte	Simazine
Unit	µg/L

Participant Results

Lab. Code	Result	Uncertainty	z	E_n
1	3.7	1.1	-0.60	-0.40
2	NT	NT		
3	4.2	0.98	0.00	0.00
4	3.64	1.46	-0.67	-0.36
5	4.5	1.35	0.36	0.20
6	3.2	0.51	-1.19	-1.28
7	4.2	1.65	0.00	0.00
8	NT	NT		
10	4.7	1.9	0.60	0.25
11	4.43	1.2	0.27	0.17
12	5.38	0.2	1.40	1.89

Statistics

Assigned Value	4.20	0.59
Spike Value	5.05	0.25
Robust Average	4.20	0.59
Median	4.20	0.62
Mean	4.22	
N	9	
Max	5.38	
Min	3.2	
Robust SD	0.70	
Robust CV	17%	

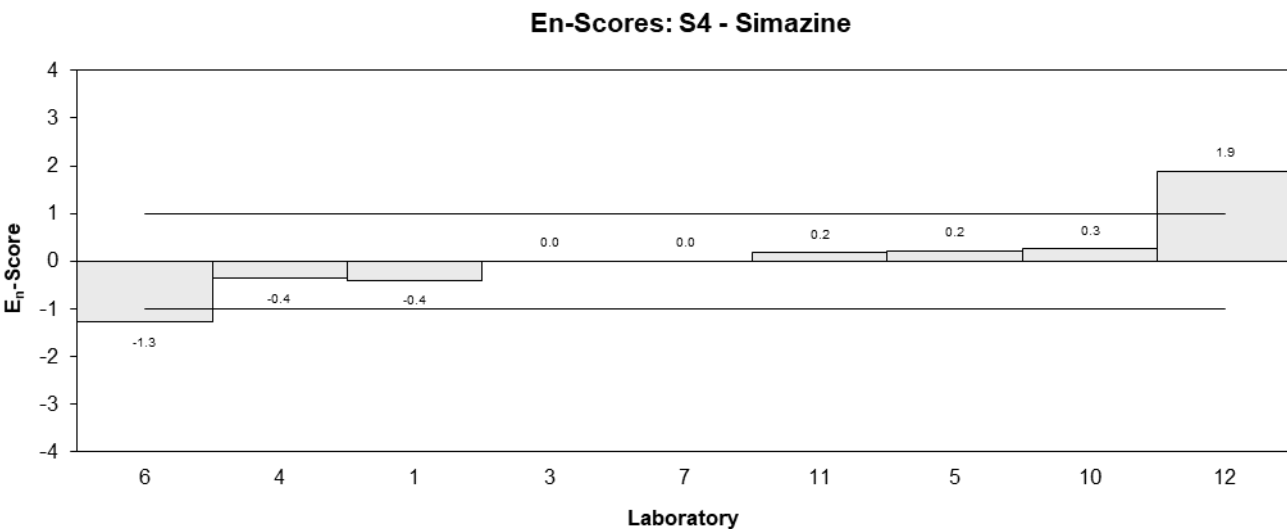
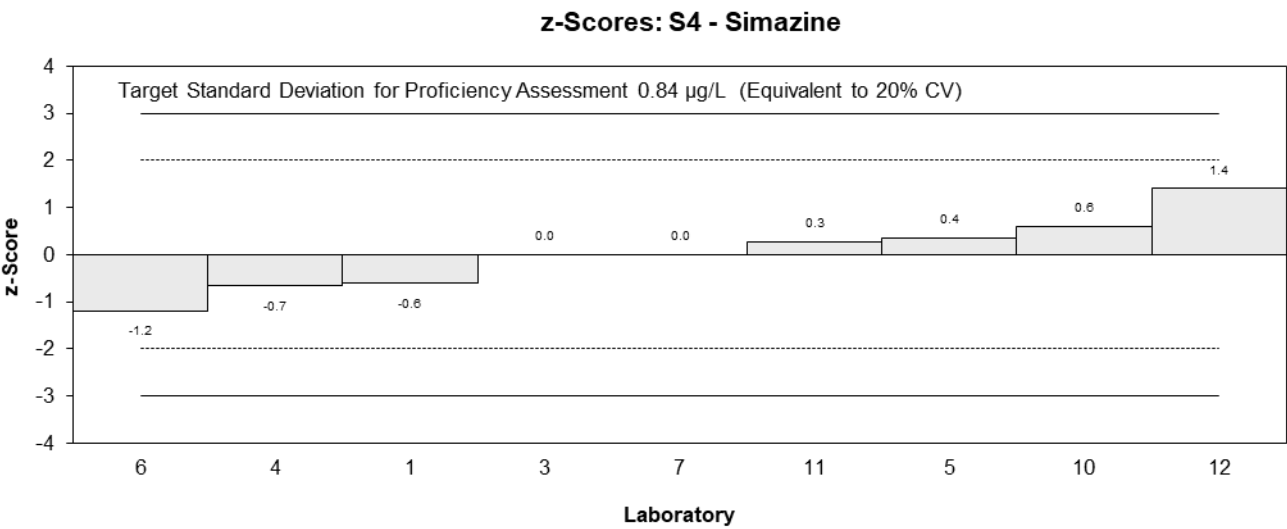
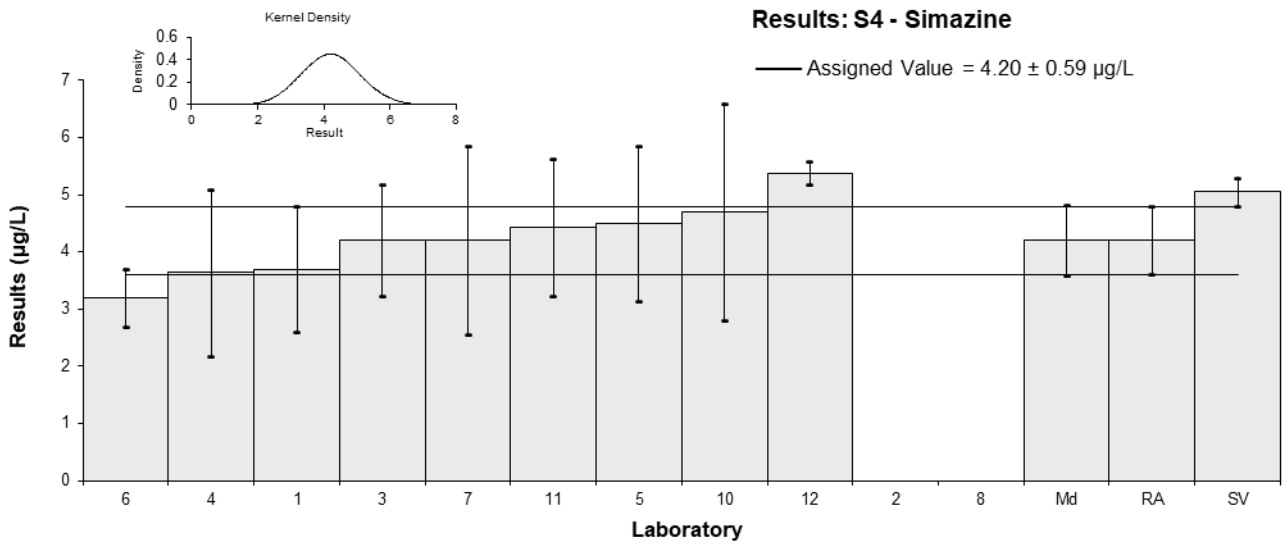


Figure 24

6 DISCUSSION OF RESULTS

6.1 Assigned Value

The robust averages and associated expanded uncertainties were calculated using the procedure described in ISO 13528.⁵ The assigned values for all scored analytes were the robust averages of participants' results, after results less than 50% and greater than 150% of the robust average had been removed.^{3,4} The calculation of the expanded uncertainty for robust averages is presented in Appendix 3, using Sample S2 benzene as an example.

Traceability: The consensus of participants' results is not traceable to any external reference, so although expressed in SI units, metrological traceability has not been established.

For Sample S1, an assigned value was set only for TRH; no assigned values were set for the individual hydrocarbon ranges. No assigned value was set for Sample S2 ethylbenzene as there was greater variability of results between participants as compared to the other BTEX analytes, and this variability was also observed in a subsequent internal investigation. No assigned values were set for Sample S3 acenaphthylene and Sample S4 dicamba as there were too few reported numeric results. Sample S2 C6-C10 was also not scored; this was due to its volatile nature and therefore data has been provided for information only, though there was reasonable consensus between participants' results for this analyte in this study. For these analytes without assigned values, participants may still compare their results with the descriptive statistics and spiked values as presented in Section 5.

A comparison of the assigned values (or robust average if no assigned value was set) and spiked values is presented in Table 29.

In general, recoveries in this study were lower than previously observed for similar analytes in river water matrix, however, this may be due to the more complex wastewater matrix. Analytes have been scored if there was a reasonable consensus between participants' results.

Table 29 Comparison of Assigned Value (*Robust Average*) and Spiked Value

Sample	Analyte	Assigned Value (<i>Robust Average</i>) (µg/L)	Spiked Value (µg/L)	Assigned Value (<i>Robust Average</i>) / Spiked Value (%)
S1	TRH	1390	2810	49
S2	Benzene	62.8	69.2	91
	Toluene	202	261	77
	Ethylbenzene	(17.7)	29.0	(61)
	Xylenes	88.0	129	68
	Total BTEX	370	489	76
S3	Acenaphthene	4.20	8.08	52
	Acenaphthylene	(0.90)	1.09	(83)
	Benz[a]anthracene	4.32	6.50	66
	Benzo[a]pyrene	1.99	3.62	55
	Fluorene	2.18	3.10	70
	Phenanthrene	1.78	1.95	91
S4	Atrazine	10.1	11.6	87
	Chlorpyrifos	14.8	20.1	74
	Dicamba	(6.2)	7.49	(83)

Sample	Analyte	Assigned Value (<i>Robust Average</i>) ($\mu\text{g/L}$)	Spiked Value ($\mu\text{g/L}$)	Assigned Value (<i>Robust Average</i>) / Spiked Value (%)
	Dieldrin	3.88	5.01	77
	Ethion	2.42	3.99	61
	Lindane	8.3	10.0	83
	Simazine	4.20	5.05	83

6.2 Measurement Uncertainty Reported by Participants

Participants were asked to report an estimate of the expanded uncertainty associated with their results and the basis of this uncertainty estimate. It is a requirement of ISO/IEC 17025 that laboratories have procedures to estimate the uncertainty of chemical measurements, and to report this uncertainty in specific circumstances, including when the client's instruction so requires.⁷

Of 206 numeric results submitted for the analytes of interest in this study, 201 (98%) were reported with an expanded MU. Participants used a wide variety of procedures to estimate their uncertainty (Table 4).

Laboratory 2 did not report a TRH result for Sample S1; the study coordinator summed the individual hydrocarbon ranges and no estimate of the uncertainty of the TRH result was made. Laboratory 6 did not report uncertainties for some of their numeric results; this participant reported that they were accredited to ISO/IEC 17025.

The magnitude of reported uncertainties was within the range of 1.3% to 50% relative to the result. In general, an expanded uncertainty of less than 15% relative is likely to be unrealistically small for routine analysis, while an uncertainty of greater than 50% relative is likely to be too large and not fit-for-purpose. Of 201 MUs reported for this study, 24 were less than 15% relative.

Uncertainties associated with results returning a satisfactory z -score but an unsatisfactory E_n -score may have been underestimated.

Laboratories 7 and 12 attached estimates of MU to non-value results reported. An estimate of uncertainty expressed as a value should not be attached to a non-value result.⁸

In some cases, the results and/or uncertainties were reported with an inappropriate number of significant figures. Including too many significant figures may inaccurately reflect the precision of measurements. The recommended format is to write the uncertainty to no more than two significant figures, and then write the result with the corresponding number of decimal places. For example, instead of $3.31 \pm 0.8606 \mu\text{g/L}$, it is better to report this as $3.31 \pm 0.86 \mu\text{g/L}$.⁸

6.3 z-Score

Wastewater was a new water matrix for NMI PT studies, and so target SDs equivalent to 20% PCV were used to calculate z -scores. CVs predicted by the Thompson-Horwitz equation,⁶ the between-laboratory CVs obtained in this study, and target SDs (as PCV) are presented for comparison in Table 30.

Table 30 Comparison of Thompson-Horwitz CV, Between-Laboratory CV and Target SD

Sample	Analyte	Assigned Value (Robust Average) (µg/L)	Thompson-Horwitz CV ^a (%)	Between-Laboratory CV ^b (%)	Target SD (as PCV) (%)
S1	>C10-C16	(690)	17	43	Not Set
	>C16-C34	(740)	17	49	Not Set
	>C34-C40	N/A	N/A	N/A	Not Set
	TRH	1390	15	38	20
S2	C6-C10	(1020)	16	14	Not Set
	Benzene	62.8	22	9.9	20
	Toluene	202	20	6.1	20
	Ethylbenzene	(17.7)	22	26	Not Set
	Xylenes	88.0	22	11	20
	Total BTEX	370	19	3.6	20
S3	Acenaphthene	4.20	22	17	20
	Acenaphthylene	(0.90)	22	36	Not Set
	Benz[<i>a</i>]anthracene	4.32	22	23	20
	Benzo[<i>a</i>]pyrene	1.99	22	31	20
	Fluorene	2.18	22	17	20
	Phenanthrene	1.78	22	28	20
S4	Atrazine	10.1	22	17	20
	Chlorpyrifos	14.8	22	15	20
	Dicamba	(6.2)	22	25	Not Set
	Dieldrin	3.88	22	17	20
	Ethion	2.42	22	14	20
	Lindane	8.3	22	17	20
	Simazine	4.20	22	17	20

^a Calculated from the assigned value (robust average).

^b Robust between-laboratory CV with outliers removed, if applicable.

To account for possible low bias in the consensus value due to participants using inefficient extraction or analytical techniques, 11 *z*-scores were adjusted across the following analytes: Sample S1 TRH, Sample S3 acenaphthene, benz[*a*]anthracene, benzo[*a*]pyrene and fluorene, and Sample S4 dieldrin and ethion. A maximum acceptable result was set as the spiked value plus two target SDs of the spiked value. Results lower than the maximum acceptable result but with a *z*-score greater than 2.0 had their *z*-score adjusted to 2.0. This ensured that any participants reporting results close to the spiked value were not penalised. *z*-Scores for results greater than the maximum acceptable result, and *z*-scores less than 2.0, were left unaltered.

Of 162 results for which *z*-scores were calculated, 157 (97%) returned a score of $|z| \leq 2.0$, indicating a satisfactory performance.

Laboratories **3, 4, 6, 7** and **10** reported numeric results for all 16 scored analytes. Of these participants, Laboratories **3, 4** and **6** returned satisfactory *z*-scores for all analytes.

Four participants received satisfactory z -scores for all reported results that were scored: Laboratories **2** (15), **8** (14), **1** (12) and **12** (12).

The dispersal of z -scores is presented by laboratory in Figure 25, and by analyte in Figure 26.

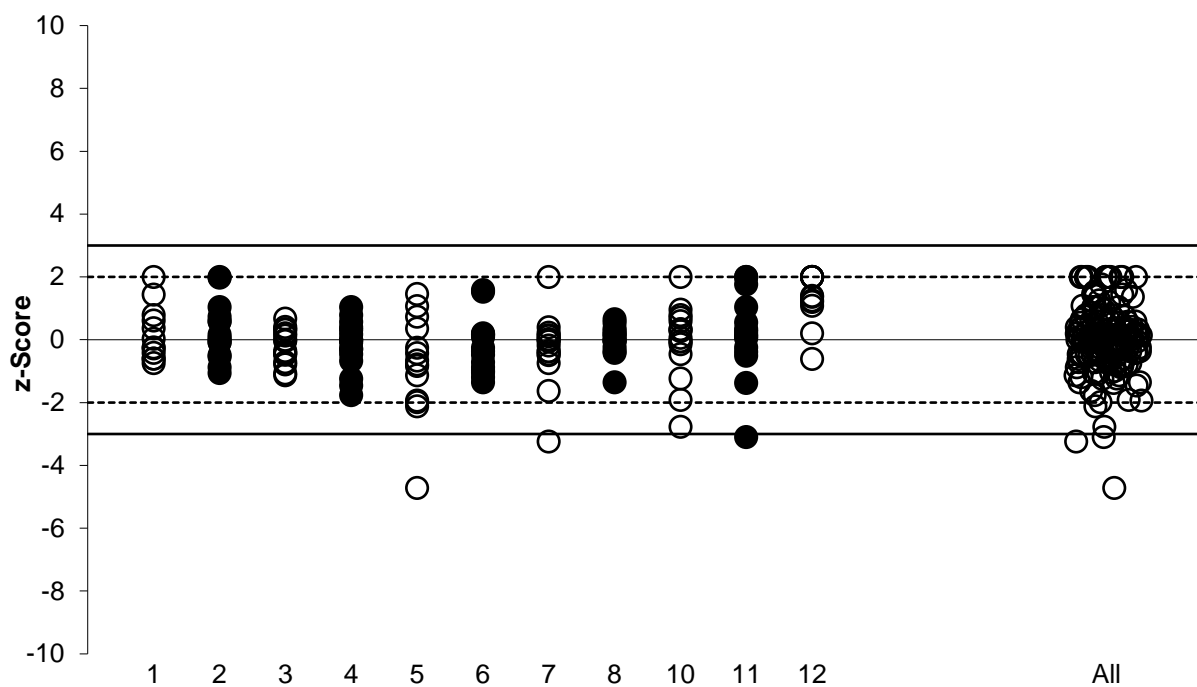


Figure 25 z -Score Dispersal by Laboratory

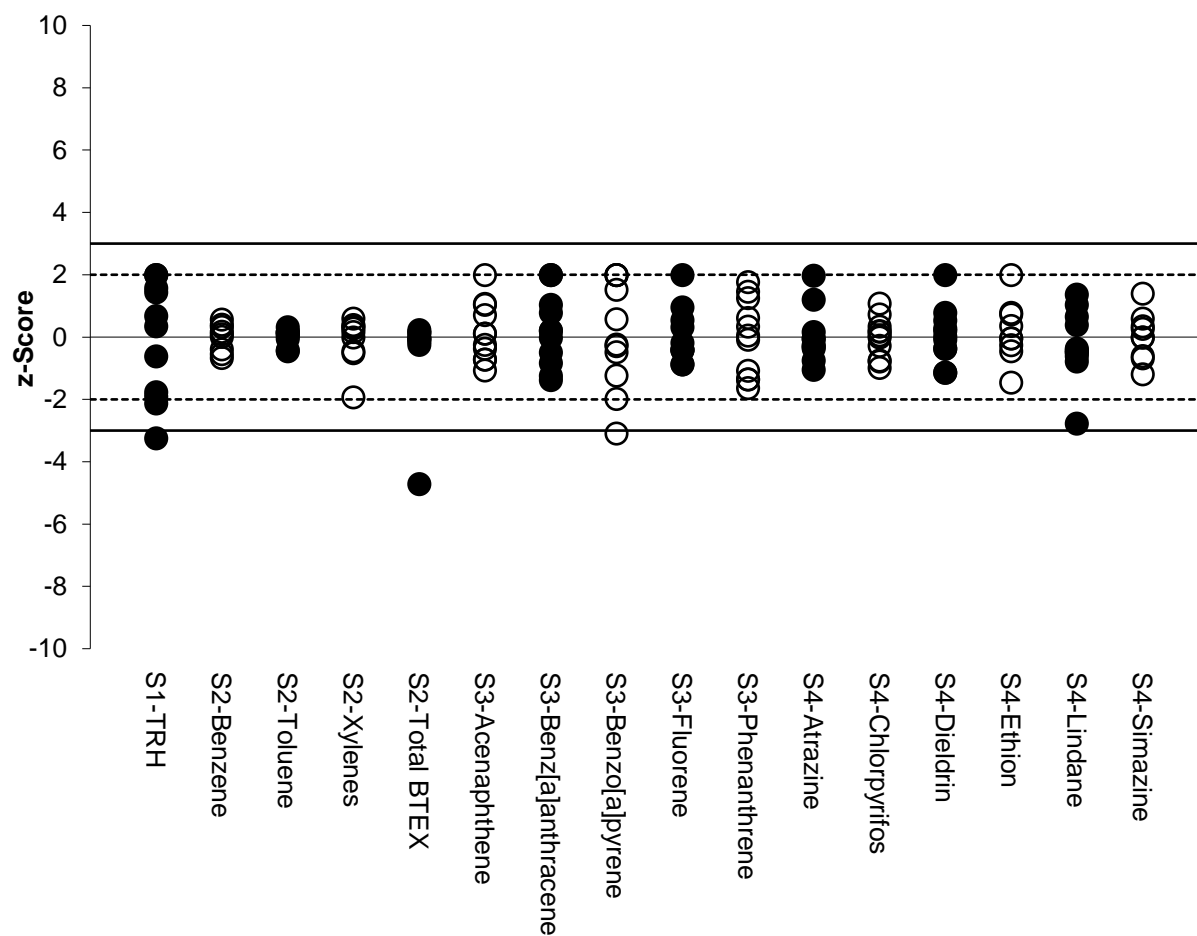


Figure 26 z -Score Dispersal by Analyte

6.4 E_n -Score

E_n -Scores can be interpreted in conjunction with z -scores, as an unsatisfactory E_n -score can either be caused by an inappropriate measurement or uncertainty, or both.

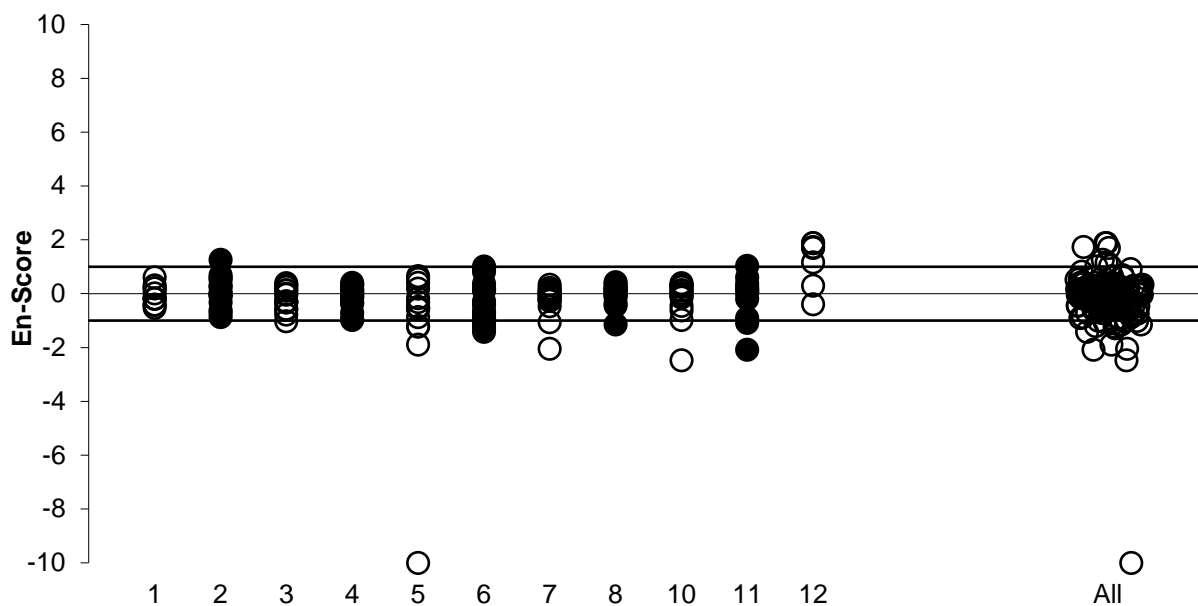
If a participant did not report an expanded MU with a result, an expanded uncertainty of zero (0) was used to calculate the E_n -score. For results whose z -scores were adjusted as discussed in Section 6.3 z -Score, no E_n -score has been calculated.

Of 151 results for which E_n -scores were calculated, 128 (85%) returned a score of $|E_n| \leq 1.0$, indicating agreement of the participant's result with the assigned value within their respective expanded uncertainties.

Laboratory 4 returned satisfactory E_n -scores for all 16 scored analytes.

One participant received satisfactory E_n -scores for all reported results that were scored: Laboratory 1 (11).

The dispersal of E_n -scores by laboratory is presented in Figure 27.



E_n -scores less than -10.0 have plotted as -10.0.

Figure 27 E_n -Score Dispersal by Laboratory

6.5 False Negatives

Table 31 presents false negative results. These are analytes present in the samples which a participant tested for, but did not report a numeric result; for example, participants reporting a 'less than' result ($< x$) when the assigned value was higher than their limit of reporting (LOR), or participants that did not report anything. For analytes where no assigned value was set, results have only been considered to be false negatives where the robust average and spiked value were significantly higher than the participants' LOR (i.e. the robust average minus the expanded uncertainty, and the spiked value minus the expanded uncertainty, were both greater than the LOR), or if no value was reported.

Table 31 False Negatives

Lab. Code	Sample	Analyte	Assigned Value (Robust Average) ($\mu\text{g/L}$)	Spiked Value ($\mu\text{g/L}$)	Result ($\mu\text{g/L}$)
5	S3	Fluorene	2.18	3.10	<0.02
12	S4	Dicamba	(6.2)	7.49	<0.05

6.6 Reporting of Additional Analytes

Analytes reported by participants which were not spiked into the test samples are presented in Table 32. In general, participants should take care to avoid any potential cross-contamination when analysing their samples.

Laboratory **11** commented that they detected propazine at a low level (0.05 µg/L) in Sample S4; this may have been a trace impurity in the atrazine spiking standard used.

Table 32 Analytes Reported by Participants Not Spiked into the Test Samples

Lab. Code	Sample	Analyte	Result (µg/L)	Uncertainty (µg/L)
4	S3	Anthracene	1.87	0.75
		Chrysene	2.51	1.00
	S4	Malathion	2.99	1.20
5	S3	Anthracene	1.5	0.45
12	S4	Piperonyl butoxide	0.06	0.02

6.7 Range of PAHs and Pesticides Analysed by Participants

Participants were provided with a list of potential PAHs (for Sample S3) and pesticides (for Sample S4) that could have been spiked into the samples, given in Tables 1 and 2 respectively. Of these, six PAHs and seven pesticides were spiked into the samples (Table 3). Participants were not required to test for all analytes, and were requested to report 'NT' (for 'Not Tested') for any that they did not analyse the samples for. A summary of participants' testing of the spiked analytes is presented in Table 33.

Laboratories **6**, **10** and **12** tested for all spiked analytes. The proportion of analytes each participant tested for ranged from 77% to 100%.

All participants tested for all the spiked PAHs. Of the spiked pesticides, all participants tested for chlorpyrifos, dieldrin and lindane. Only 4 participants (45%) tested for dicamba.

Table 33 Summary of Participants' Analyses

Lab. Code Analyte	1	2	3	4	5	6	7	8	10	11	12	Proportion of Participants (%)
Acenaphthene	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Acenaphthylene	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Benz[<i>a</i>]anthracene	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Benzo[<i>a</i>]pyrene	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Fluorene	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Phenanthrene	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Atrazine	✓	✓	✓	✓	✓	✓	✓	NT	✓	✓	✓	91
Chlorpyrifos	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Dicamba	NT	✓	NT	NT	NT	✓	NT	NT	✓	✓	✓	45
Dieldrin	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Ethion	✓	✓	✓	✓	NT	✓	✓	✓	✓	NT	✓	82
Lindane	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	100
Simazine	✓	NT	✓	✓	✓	✓	✓	NT	✓	✓	✓	82
Proportion of Analytes (%)	92	92	92	92	85	100	92	77	100	92	100	

6.8 Participants' Analytical Methods

Participants used a variety of analytical methods for the test samples (Appendix 4).

Sample S1 TRH

For Sample S1, participants were provided with 500 mL, and participants reported using test portions ranging from 40 mL to the whole bottle. A comparison of results and the sample volume used is presented in Figure 28. For this study, some participants using smaller sample volumes reported lower TRH results. Participants subsampling the bottle should ensure that the subsample taken is a suitable representation of the whole sample.

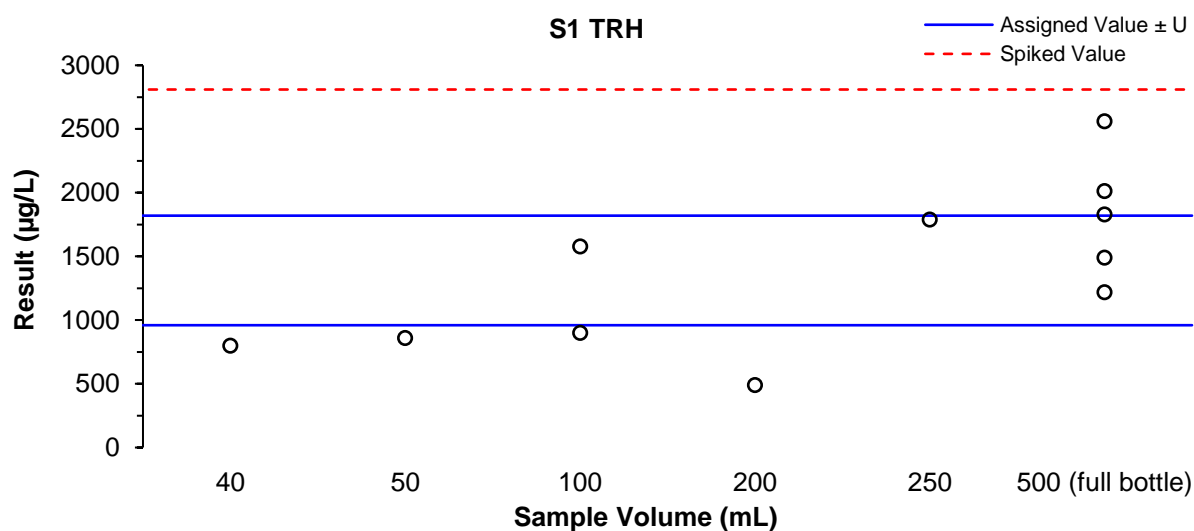


Figure 28 Sample S1 TRH Results vs Sample Volume

Most participants used liquid-liquid extraction (LLE), with one participant using solid phase extraction (SPE). For extraction solvents, participants used either dichloromethane (DCM) or hexane (HEX). Two participants reported a silica clean-up step, and one participant reported a filtration step. Most participants used gas chromatography (GC) coupled with flame ionisation detection (FID), with one participant using GC coupled with mass chromatography (MS) instead.

A plot of results against the methodology employed (extraction technique, extraction solvent and measurement instrument) is presented in Figure 29. In this study, the participants using hexane as the extraction solvent in general returned lower recoveries of TRH.

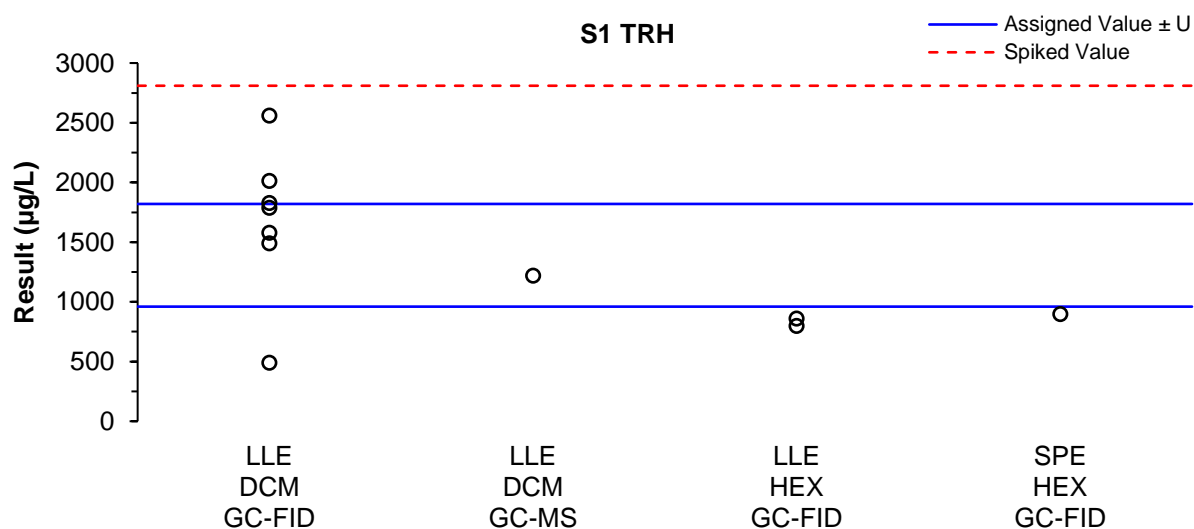


Figure 29 Sample S1 TRH Results vs Methodology

Sample S2 BTEX

For Sample S2 Total BTEX, Laboratory 5 reported a result of 21.5 $\mu\text{g/L}$, which was significantly lower than many of their individual BTEX analyte results. This result was therefore excluded from all statistical calculations as an extreme outlier, and has also been excluded from the following discussion.

For Sample S2, participants were provided with 2 x 42 mL (two vials were provided so that participants could repeat the measurement if required). Participants reported test portions of either 5 mL or the whole vial. A comparison of the results and sample volume used is presented in Figure 30; there was no evident correlation observed.

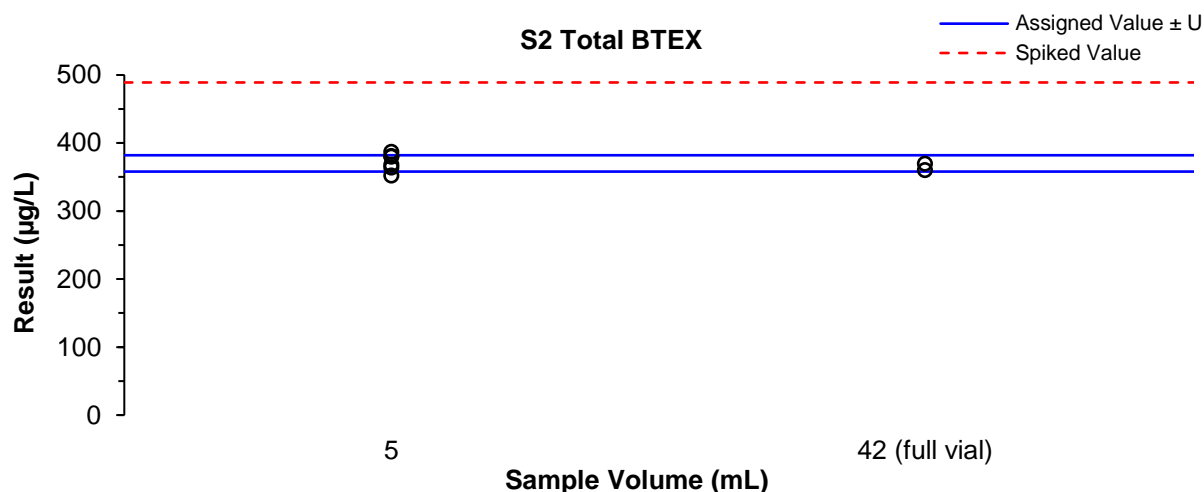


Figure 30 Sample S2 Total BTEX Results vs Sample Volume

All participants used purge-and-trap (P&T) GC-MS(/MS). One participant reported LLE using methanol (MeOH) as part of their preparation. A plot of results against methodology employed is presented in Figure 31.

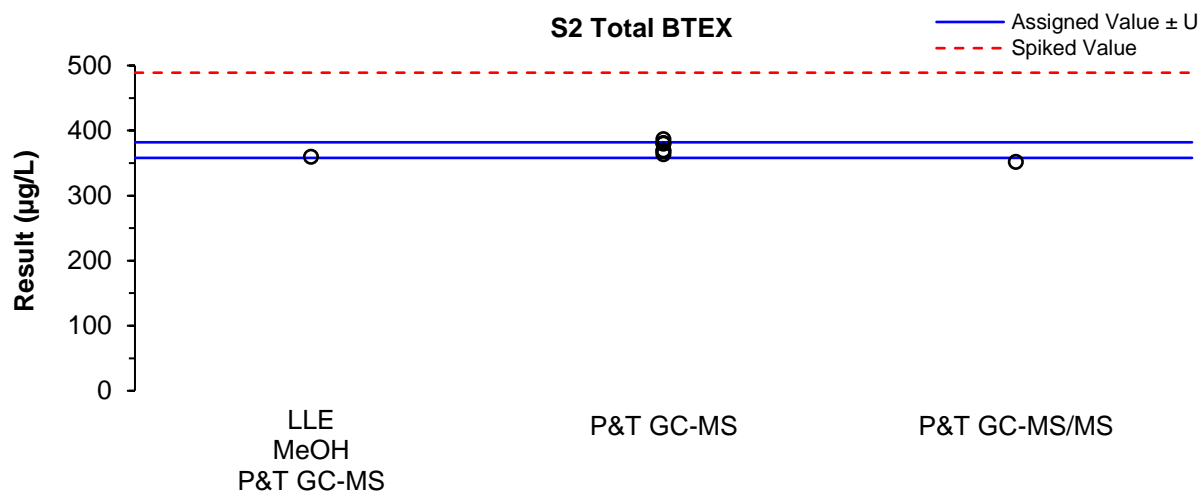


Figure 31 Sample S2 Total BTEX Results vs Methodology

Sample S3 PAHs

For Sample S3, participants were given the option of samples as 1 x 500 mL (3 participants) or as 3 x 100 mL (8 participants), depending on what suited their laboratory's method. Participants reported test portions ranging from 40 mL to 500 mL. A comparison of z-scores for scored analytes and sample volume used is presented in Figure 32; there was no evident correlation observed.

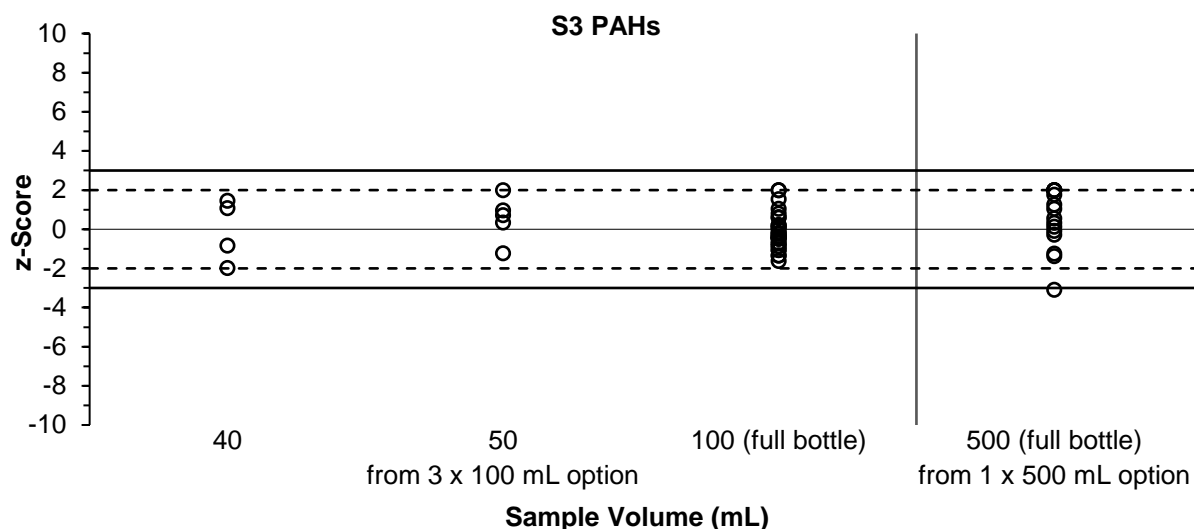


Figure 32 Sample S3 PAHs z-Scores vs Sample Volume

Participants reported using either LLE or SPE, with DCM, HEX or DCM/ethyl acetate (EtOAc) as the extraction solvent. One participant reported using a filtration step. All participants used GC-MS(/MS) for analysis. A plot of z-scores against methodology employed is presented in Figure 33; there was no evident correlation observed.

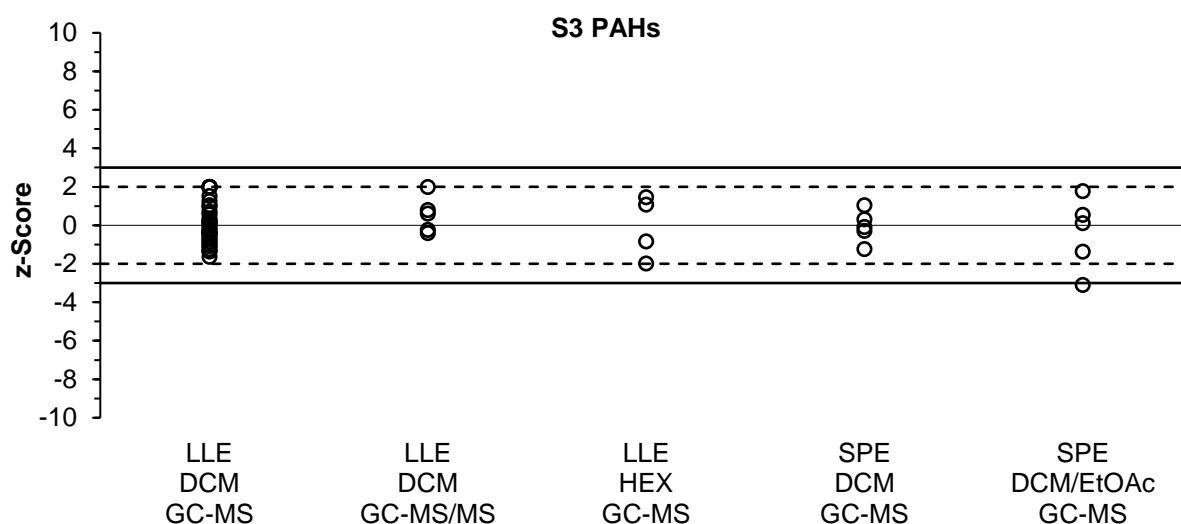


Figure 33 Sample S3 PAHs z-Scores vs Methodology

Sample S4 Pesticides

For Sample S4, participants were given the option of samples as 1 x 500 mL (3 participants) or as 3 x 100 mL (8 participants), depending on what suited their laboratory's method. Participants reported test portions ranging from 1 mL to 500 mL. A comparison of z-scores for scored analytes and sample volume used is presented in Figure 34 (if a participant did not report a volume, this has been marked as NR).

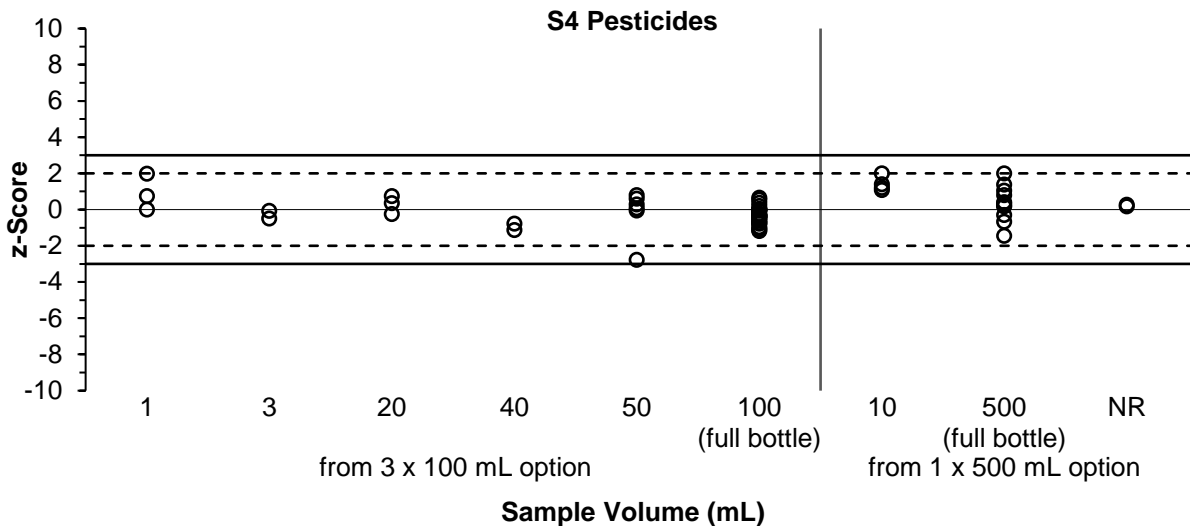


Figure 34 Sample S4 Pesticides z-Scores vs Sample Volume

Participants reported using LLE, QuEChERS, SPE, or direct injection (DI). Extraction solvents used included acetonitrile (ACN), DCM, DCM/EtOAc, HEX and toluene. Most participants did not use a clean-up step for their analysis, though a few participants reported a centrifugation, dilution and/or filtration step. Samples were analysed using GC coupled with electron capture detection (ECD), GC-MS(/MS) or liquid chromatography (LC) coupled with MS(/MS).

Figures 35 to 40 show comparisons of results and methodology employed for scored analytes. If a participant did not report part of their methodology, this has been marked as NR. A wide range of methodologies were employed, and there was no evident correlation observed.

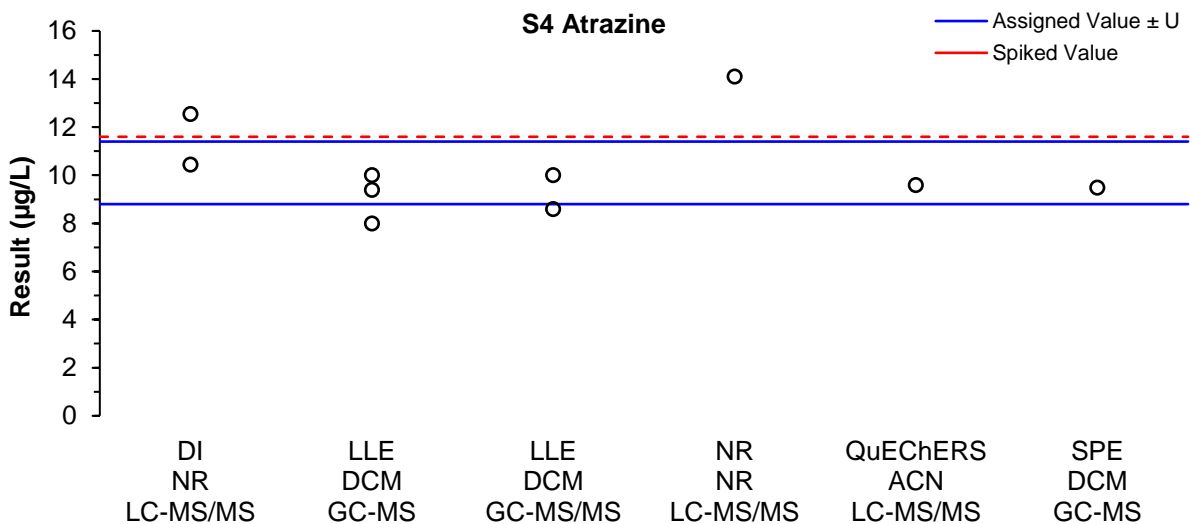


Figure 35 Sample S4 Atrazine Results vs Methodology

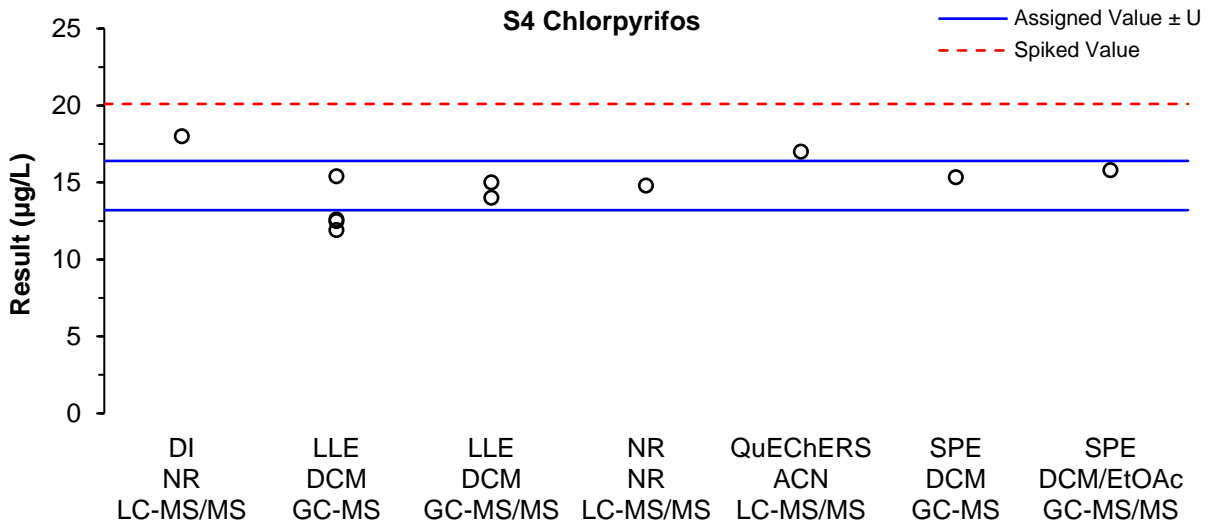


Figure 36 Sample S4 Chlorpyrifos Results vs Methodology

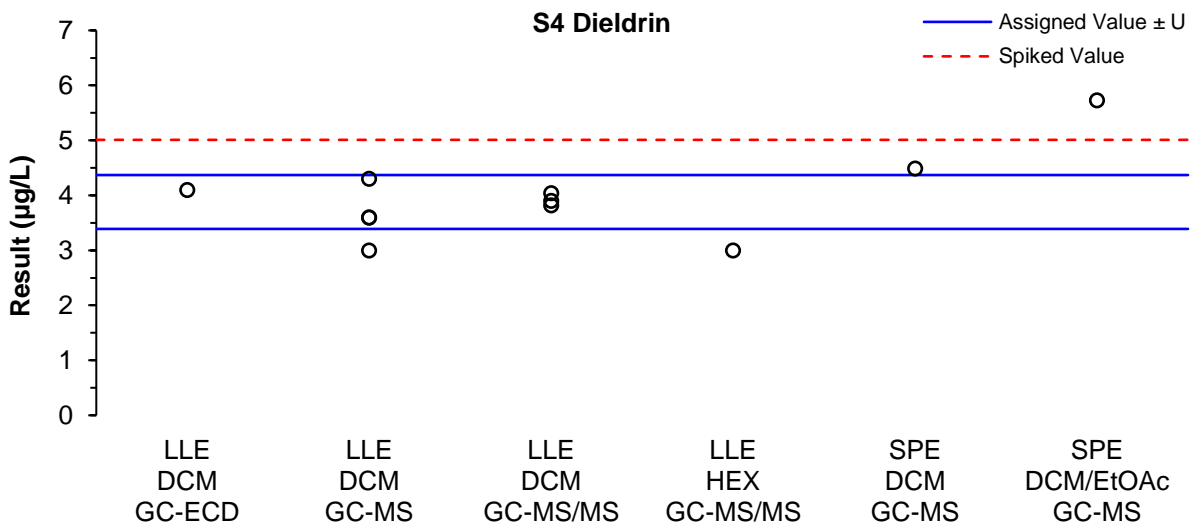


Figure 37 Sample S4 Dieldrin Results vs Methodology

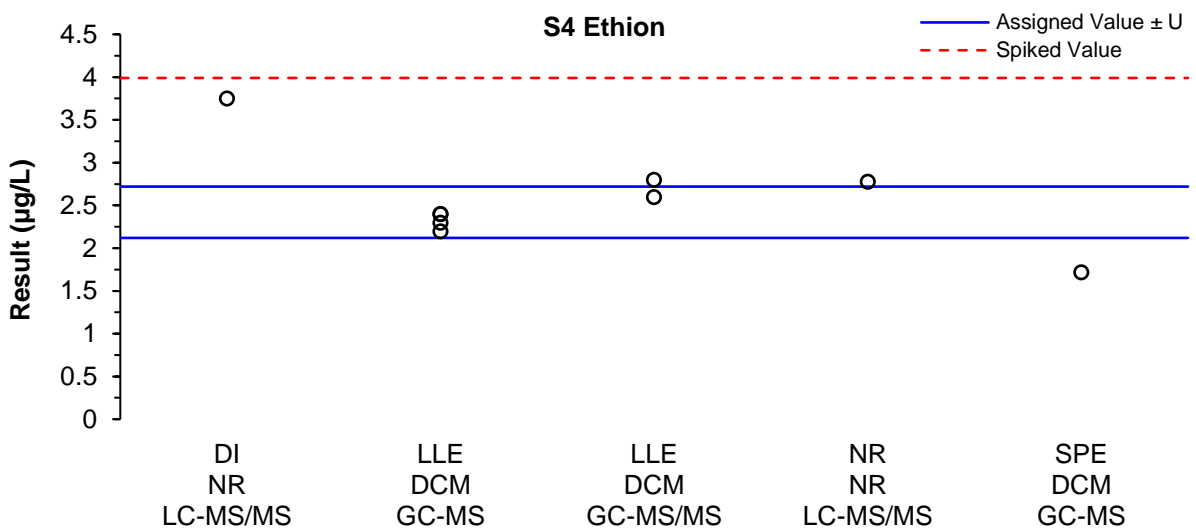


Figure 38 Sample S4 Ethion Results vs Methodology

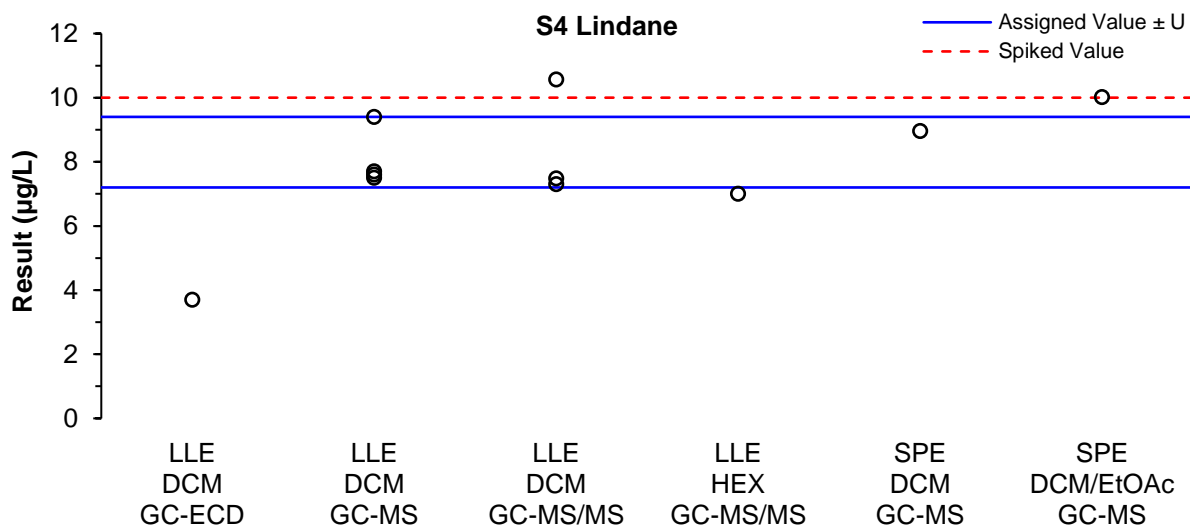


Figure 39 Sample S4 Lindane Results vs Methodology

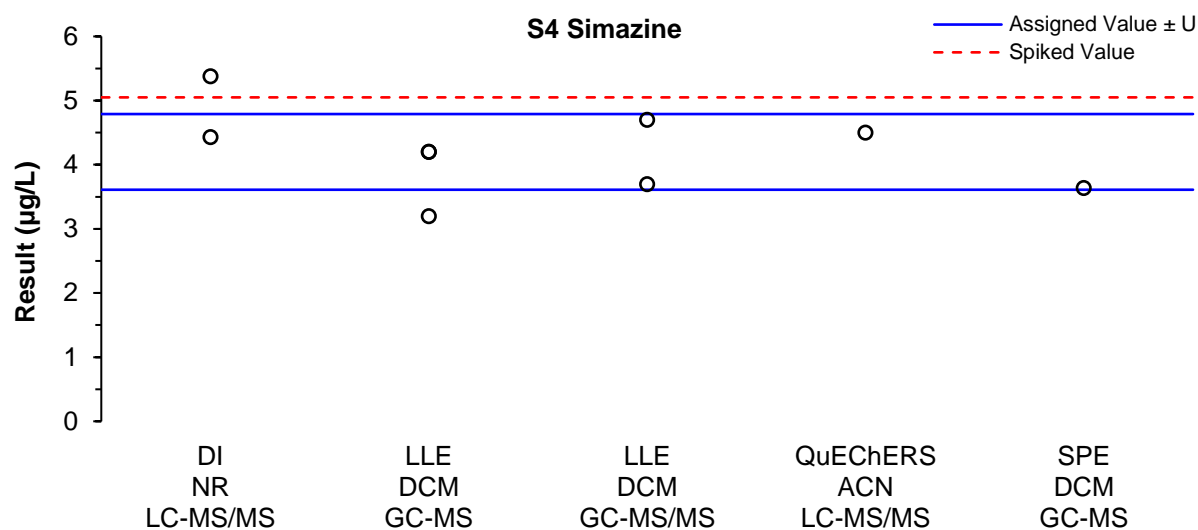


Figure 40 Sample S4 Simazine Results vs Methodology

6.9 Certified Reference Materials

Participants were requested to indicate whether certified standards or matrix reference materials had been used as part of the quality assurance for their analysis.

Nine participants reported using certified standards. The following were listed:

- AccuStandard
- o2si
- ISO 17034 compliant standards

These materials may or may not meet the internationally recognised definition of a certified reference material:

‘reference material, accompanied by documentation issued by an authoritative body and providing one or more specified property values with associated uncertainties and traceabilities, using valid procedures’⁹

6.10 Comparison with TRH and Total BTEX in River Water

This PT study was NMI's first study for organic compounds and pesticides in wastewater, however NMI currently runs annual studies for similar compounds in river water.

A comparison of TRH in wastewater, with TRH in river water for the last five studies (2018–2022), is shown in Figure 41. In river water, the recovery has remained relatively similar across the different studies and across different spiked levels, with an average recovery of 62%. For this study, the recovery in wastewater was lower, at 49%.

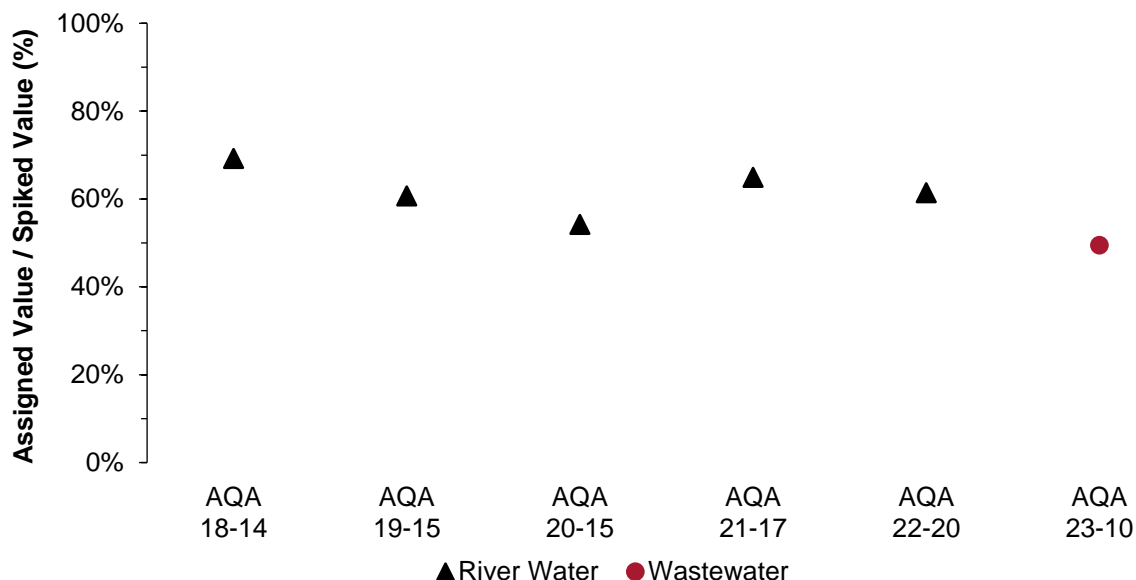


Figure 41 TRH Results in River Water and Wastewater

A comparison of total BTEX in wastewater, with total BTEX in river water for the last five studies (2018–2022), is shown in Figure 42. In river water, the recovery has remained relatively high across the different studies and across different spiked levels, with an average recovery of 89%. For this study, the recovery in wastewater was lower, at 76%.

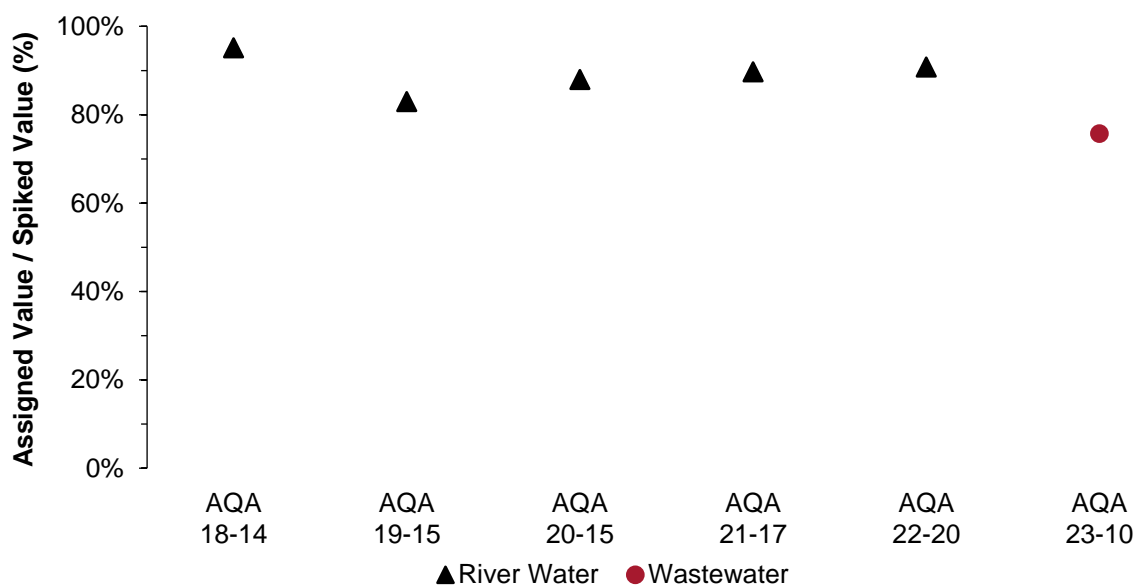


Figure 42 Total BTEX Results in River Water and Wastewater

The lower recoveries observed in the wastewater samples may be due to an increased complexity of the matrix.

6.11 Summary of Participants' Results and Performances

Summaries of participants' results and performances in this PT study for scored analytes are presented in Tables 34 to 36, and Figure 43.

Table 34 Summary of Participants' Results for Scored Analytes in Samples S1 and S2*

Lab. Code	Sample S1	Sample S2			
	Total TRH	Benzene	Toluene	Xylenes	Total BTEX
AV	1390	62.8	202	88.0	370
SV	2810	69.2	261	129	489
1	1790	NT	NT	NT	NT
2	2014	56	209	98.5	364
3	1580	68	215	94	380
4	900	70.02	183.9	87.99	369.63
5	800	54.5	185	54	21.5
6	1830	65	206	91	381
7	490	63	208	95	387
8	1490	64	209	93	368
10	860	67	200	80	360
11	2560	58	204	79	352
12	1220	NT	NT	NT	NT

* All values are in $\mu\text{g/L}$. Shaded cells are results which returned a questionable or unsatisfactory z -score. AV = Assigned Value, SV = Spiked Value.

Table 35 Summary of Participants' Results for Scored Analytes in Sample S3*

Lab. Code	Sample S3				
	Acenaphthene	Benz[a]anthracene	Benzo[a]pyrene	Fluorene	Phenanthrene
AV	4.20	4.32	1.99	2.18	1.78
SV	8.08	6.50	3.62	3.10	1.95
1	4	5	4	2	2
2	3.31	5.22	2.22	1.8	1.8
3	3.6	4.5	1.8	2.0	1.4
4	5.07	3.25	1.88	2.32	1.75
5	5.1	3.6	1.2	<0.02	2.3
6	3.6	4.5	2.6	1.8	1.3
7	4.3	3.9	2.8	2.1	1.2
8	3.9	4.3	1.9	2.0	1.3
10	4.8	6.8	1.5	2.6	1.9
11	4.3	3.14	0.76	2.42	2.41
12	6.67	7.67	4.00	3.19	2.23

* All values are in $\mu\text{g/L}$. Shaded cells are results which returned a questionable or unsatisfactory z -score. AV = Assigned Value, SV = Spiked Value.

Table 36 Summary of Participants' Results for Scored Analytes in Sample S4*

Lab. Code	Sample S4					
	Atrazine	Chlorpyrifos	Dieldrin	Ethion	Lindane	Simazine
AV	10.1	14.8	3.88	2.42	8.3	4.20
SV	11.6	20.1	5.01	3.99	10.0	5.05
1	8.6	14	3.9	2.6	7.3	3.7
2	14.1	14.8	3.82	2.78	7.48	NT
3	9.4	12.5	3.0	2.4	7.6	4.2
4	9.49	15.34	4.49	1.7183	8.96	3.64
5	9.6	17	3	NT	7	4.5
6	8	11.9	3.6	2.3	7.5	3.2
7	10	12.6	3.6	2.2	7.7	4.2
8	NT	15.4	4.3	2.4	9.4	NT
10	10	15	4.1	2.8	3.7	4.7
11	10.45	15.8	5.73	NT	10.02	4.43
12	12.55	18.00	4.04	3.75	10.57	5.38

* All values are in µg/L. Shaded cells are results which returned a questionable or unsatisfactory z-score. AV = Assigned Value, SV = Spiked Value.

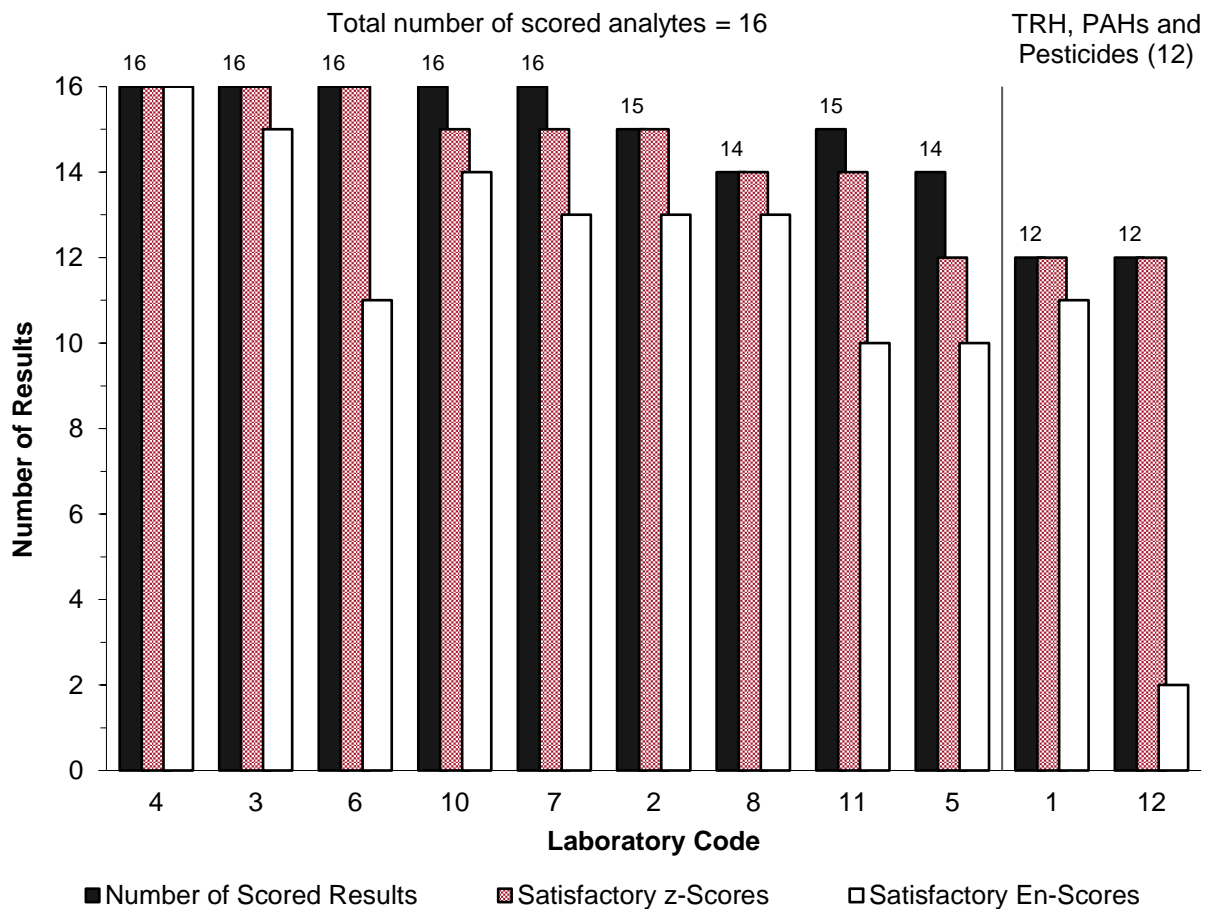


Figure 43 Summary of Participants' Performance

7 REFERENCES

Please note that for all undated references, the latest edition of the referenced document (including any amendments) applies.

- [1] ISO/IEC 17043:2010, *Conformity assessment – General requirements for proficiency testing*.
- [2] NMI, 2023, *Study Protocol for Proficiency Testing*, viewed September 2023, <https://www.industry.gov.au/sites/default/files/2020-10/cpt_study_protocol.pdf>
- [3] NMI, 2023, *Chemical Proficiency Testing Statistical Manual*, viewed September 2023, <https://www.industry.gov.au/sites/default/files/2019-07/cpt_statistical_manual.pdf>
- [4] Thompson, M., Ellison, S.L.R. and Wood, R., 2006, ‘The International Harmonized Protocol for the Proficiency Testing of Analytical Chemistry Laboratories’, *Pure Appl. Chem.*, vol 78, pp 145–196.
- [5] ISO 13528, *Statistical methods for use in proficiency testing by interlaboratory comparison*.
- [6] Thompson, M., 2000, ‘Recent Trends in Inter-laboratory Precision at ppb and sub-ppb Concentrations in Relation to Fitness for Purpose Criteria in Proficiency Testing’, *Analyst*, vol 125, pp 385–386.
- [7] ISO/IEC 17025, *General requirements for the competence of testing and calibration laboratories*.
- [8] Eurachem/CITAC Guide CG 4, QUAM:2012.P1, *Quantifying Uncertainty in Analytical Measurement*, 3rd ed., viewed September 2023, <http://www.eurachem.org/images/stories/Guides/pdf/QUAM2012_P1.pdf>
- [9] BIPM, JCGM 200:2012, *International vocabulary of metrology – Basic and general concepts and associated terms (VIM)*, 3rd ed.

APPENDIX 1 SAMPLE PREPARATION

A1.1 Diesel Fuel and Water Preparation

Diesel fuel was purchased from a local retail outlet and treated to remove volatiles. Approximately 500 mL of diesel fuel was placed in a heated (80 °C) open container and sparged with nitrogen. Treatment continued until the GC-FID chromatogram indicated that essentially all the hydrocarbons eluting before C₁₀ had been removed. This same treated diesel fuel had been used in previous NMI Hydrocarbon PTs.

Wastewater was obtained from Sydney Water at West Ryde. It was filtered through an ADVANTEC GB140 Glass Filter and autoclaved.

A1.2 Test Sample Preparation

Sample S1

A diesel spiking solution was prepared by weighing 0.51017 g of the treated diesel fuel into a 500 mL volumetric flask and making to volume with methanol. Amber glass bottles of approximately 500 mL capacity were rinsed with acetone and dried. The cleaned bottles were placed in an air-conditioned room overnight. Filtered, autoclaved water (538.4 ± 0.2 g, 540 mL at 25 °C) was weighed into each of the bottles. This amount of water was selected to minimise the headspace in the sample bottles after spiking. The methanol/diesel spiking solution (1.490 mL) was added to each bottle using a Hamilton dispenser. The bottles were immediately capped and inverted to mix the solution. Each bottle was then labelled and shrink-wrapped.

Sample S2

Filtered, autoclaved water (41.88 ± 0.05 g, 42 mL) was weighed into headspace vials. A composite spike solution was prepared by adding aliquots of diesel and unleaded petrol to methanol. Two of the BTEX compounds were fortified with additional laboratory solvent. The composite spiking solution was made up to volume with methanol. Composite spiking solution (1.0 mL) was added to each vial using a Hamilton dispenser. Each vial was capped after spiking, labelled and shrink-wrapped.

Sample S3

The PAH spike solutions were prepared by dissolving each standard material in dichloromethane. A 1:1 mix of Milli-Q water and filtered autoclaved wastewater (16045 g) was placed into a stainless steel container. After spiking, the water was stirred using a top-driven impeller stirrer for at least two hours. The samples were then dispensed into 500 mL and 100 mL amber glass bottles which were labelled and shrink-wrapped.

Sample S4

The pesticide spike solutions were prepared by dissolving each standard material in acetone. A 1:1 mix of Milli-Q water and filtered autoclaved wastewater (15341 g) was placed into a stainless steel container. After spiking, the water was stirred using a top-driven impeller stirrer for at least two hours. The samples were then dispensed into 500 mL and 100 mL amber glass bottles which were labelled and shrink-wrapped.

For all samples, the samples were stored at 4 °C between preparation and dispatch.

APPENDIX 2 ASSESSMENT OF HOMOGENEITY AND STABILITY

A2.1 Homogeneity

No homogeneity testing was completed for this study as the samples were prepared using a process previously demonstrated to produce sufficiently homogeneous samples.

Analytes were only set assigned values, and participants' results scored, if there was a reasonable consensus of participants' results.

A2.2 Stability

Samples were stored at 4 °C after preparation and prior to dispatch. For dispatch, the samples were packaged into insulated polystyrene foam boxes with cooler bricks, and all samples were delivered within two days of dispatch for this study. No stability testing was conducted for this study as the samples were prepared, stored and dispatched using a process previously demonstrated to produce sufficiently stable samples.

Comparisons of z-scores to days spent in transit for scored analytes are presented in Figures 44 to 56 (extreme outliers have been removed). No evidence of significant analyte degradation with respect to the amount of time spent in transit was observed.

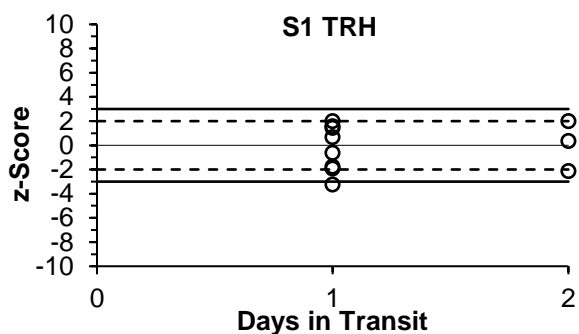


Figure 44 S1 TRH z-Score vs Transit Time

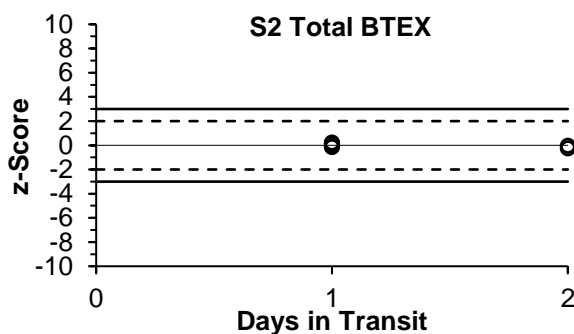


Figure 45 S2 Total BTEX z-Score vs Transit Time

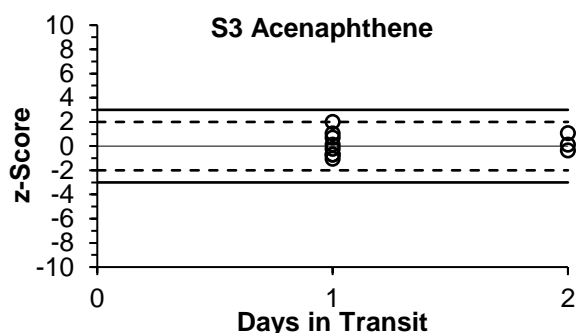


Figure 46 S3 Acenaphthene z-Score vs Transit Time

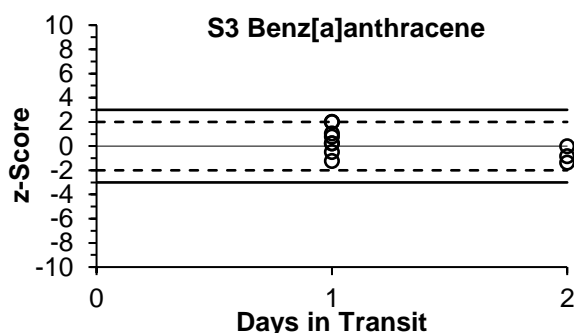


Figure 47 S3 Benz[a]anthracene z-Score vs Transit Time

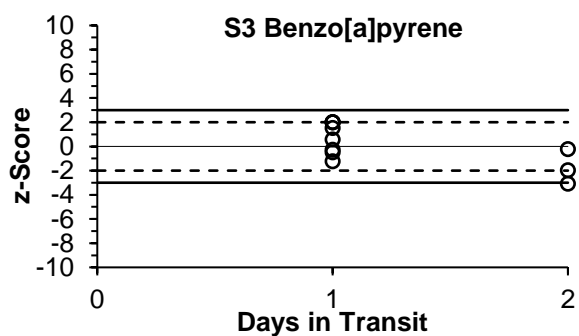


Figure 48 S3 Benzo[a]pyrene z-Score vs Transit Time

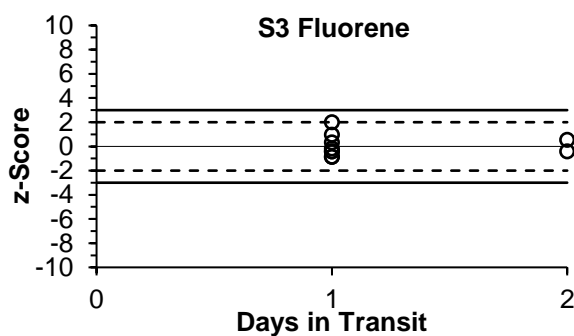


Figure 49 S3 Fluorene z-Score vs Transit Time

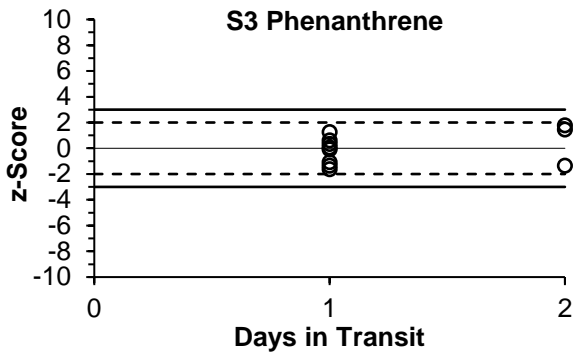


Figure 50 S3 Phenanthrene z -Score vs Transit Time

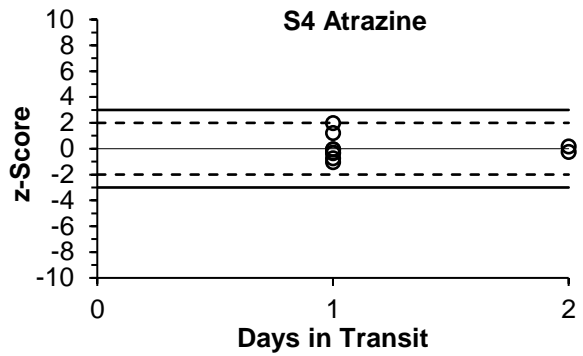


Figure 51 S4 Atrazine z -Score vs Transit Time

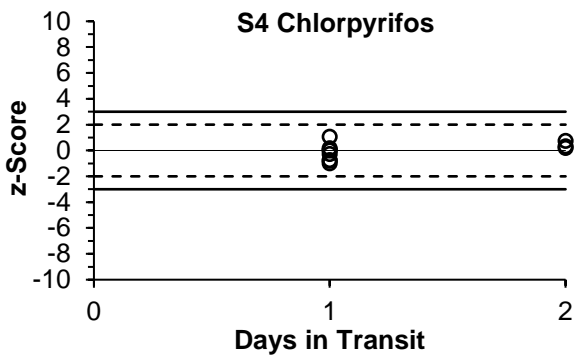


Figure 52 S4 Chlorpyrifos z -Score vs Transit Time

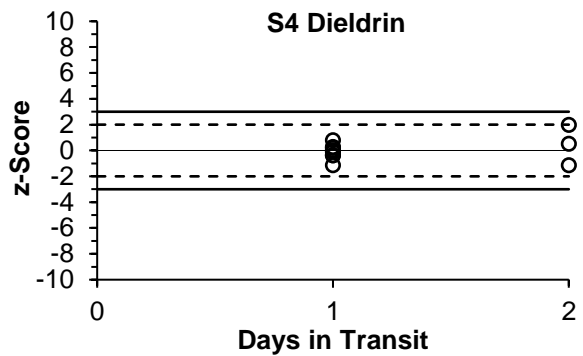


Figure 53 S4 Dieldrin z -Score vs Transit Time

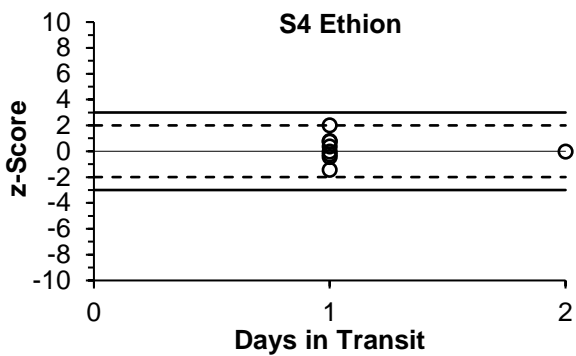


Figure 54 S4 Ethion z -Score vs Transit Time

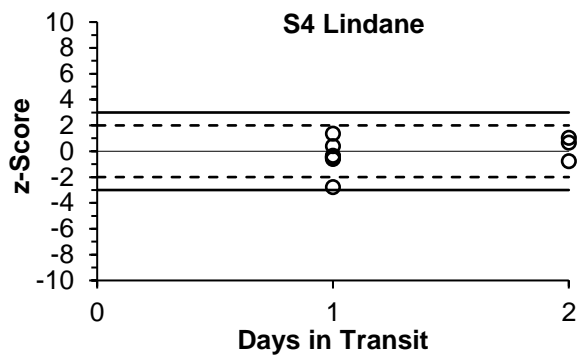


Figure 55 S4 Lindane z -Score vs Transit Time

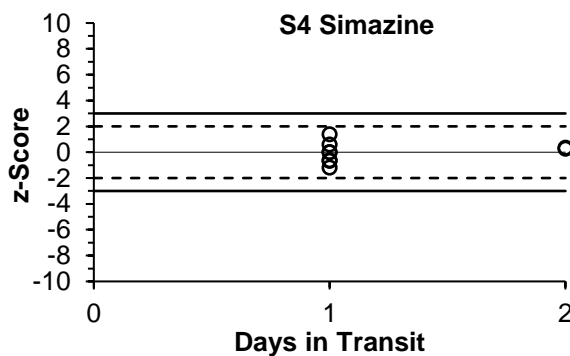


Figure 56 S4 Simazine z -Score vs Transit Time

APPENDIX 3 ROBUST AVERAGE AND ASSOCIATED UNCERTAINTY, z-SCORE AND E_n -SCORE CALCULATIONS

A3.1 Robust Average and Associated Uncertainty

Robust averages were calculated using the procedure described in ISO 13528.⁵ The associated uncertainties were estimated as according to Equation 4.

$$u_{rob\ av} = \frac{1.25 \times S_{rob\ av}}{\sqrt{p}} \quad \text{Equation 4}$$

where:

$u_{rob\ av}$ is the standard uncertainty of the robust average

$S_{rob\ av}$ is the standard deviation of the robust average

p is the number of results

The expanded uncertainty ($U_{rob\ av}$) is the standard uncertainty multiplied by a coverage factor of 2 at approximately 95% confidence level.

A worked example is set out below in Table 37.

Table 37 Uncertainty of Robust Average for Sample S2 Benzene

Number of results (p)	9
Robust Average	62.8 $\mu\text{g/L}$
$S_{rob\ av}$	6.2 $\mu\text{g/L}$
$u_{rob\ av}$	2.6 $\mu\text{g/L}$
k	2
$U_{rob\ av}$	5.2 $\mu\text{g/L}$

Therefore, the robust average for Sample S2 benzene is $62.8 \pm 5.2 \mu\text{g/L}$.

A3.2 z-Score and E_n -Score Calculation

For each participant's result, a z-score and E_n -score are calculated according to Equations 2 and 3 respectively (Section 4).

A worked example is set out below in Table 38, using the result reported by Laboratory 1 for Sample S1 TRH.

Table 38 z-Score and E_n -Score for Sample S1 TRH Result Reported by Laboratory 1

Participant Result ($\mu\text{g/L}$)	Assigned Value ($\mu\text{g/L}$)	Target Standard Deviation	z-Score	E_n -Score
1790 ± 500	1390 ± 430	20% as PCV, or: $0.2 \times 1390 = 278 \mu\text{g/L}$	$z = \frac{1790 - 1390}{278}$ $= 1.44$	$E_n = \frac{1790 - 1390}{\sqrt{500^2 + 430^2}}$ $= 0.61$

APPENDIX 4 PARTICIPANTS' TEST METHODS

Participants were requested to provide information about their test methods. Responses are presented in Tables 39 to 48. Some responses may be modified so that the participant cannot be identified.

Table 39 Methodology – Sample S1 TRH

Lab. Code	Sample Volume (mL)	Extraction Details	Extraction Solvent	Clean-Up	Measurement Instrument
1	250	Liquid-Liquid	DCM	Silica Gel	GC-FID
2	539	Liquid-Liquid	DCM	None	GC-FID
3	100	Liquid-Liquid	DCM	None	GC-FID
4	100	SPE	Hexane	None	GC-FID
5	40	Liquid-Liquid	Hexane	None	GC-FID
6	530	Liquid-Liquid	DCM	None	GC-FID
7	200	Liquid-Liquid	DCM	Filtration	GC-FID
8	500	Liquid-Liquid	DCM	None	GC-FID
10	50	Liquid-Liquid	Hexane	None	GC-FID
11	500	Liquid-Liquid	DCM	Silica	GC-FID
12	500	Liquid-Liquid	DCM	None	GC-MS

Table 40 Methodology – Sample S2 BTEX

Lab. Code	Sample Volume (mL)	Extraction Details	Extraction Solvent	Clean-Up	Measurement Instrument
1	NT				
2	5	Purge & Trap	None	None	P&T GC-MS
3	5	Purging	None	None	P&T GC-MS
4	42	Direct Injection	Water	None	P&T GC-MS
5	10	Direct Injection		None	Headspace GC-MS
6	5	Direct Injection	None	None	P&T GC-MS
7	5	Purge and Trap	NA	NA	P&T GC-MS
8	5	Purge & Trap	n/a	None	GC-MS
10	44	Liquid-Liquid	MeOH	None	P&T GC-MS
11	5	Direct Injection	N/A	None	P&T GC-MS/MS
12	NT				

Table 41 Methodology – Sample S3 PAHs

Lab. Code	Sample Volume (mL)	Extraction Details	Extraction Solvent	Clean-Up	Measurement Instrument
1	100	Liquid-Liquid	DCM	None	GC-MS/MS
2	102	Liquid-Liquid	DCM	None	GC-MS
3	100	Liquid-Liquid	DCM	None	GC-MS

Lab. Code	Sample Volume (mL)	Extraction Details	Extraction Solvent	Clean-Up	Measurement Instrument
4	500	SPE	DCM	None	GC-MS
5	40	Liquid-Liquid	Hexane	None	GC-MS
6	116	Liquid-Liquid	DCM	None	GC-MS
7	100	Liquid-Liquid	DCM	Filtration	GC-MS
8	100	Liquid-Liquid	DCM	None	GC-MS
10	50	Liquid-Liquid	DCM	None	GC-MS
11	500	SPE	DCM/ethyl acetate	None	GC-MS
12	500	Liquid-Liquid	DCM	None	GC-MS

Table 42 Methodology – Sample S4 Atrazine

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
1	100	Liquid-Liquid	DCM	None	GC-MS/MS
2	1			Centrifugation & dilution	LC-MS/MS
3	100	Liquid-Liquid	DCM	None	GC-MS
4	500	SPE	DCM	None	GC-MS
5	20	QuEChERS	acetonitrile	None	LC-MS/MS
6	100	Liquid-Liquid	DCM	None	GC-MS
7	100	Liquid-Liquid	DCM	Filtration	GC-MS
8	NT				
10	50	Liquid-Liquid	DCM	N/A	GC-MS/MS
11		Direct Injection			LC-MS/MS
12	10	Direct Injection		Centrifuge	LC-MS/MS

Table 43 Methodology – Sample S4 Chlorpyrifos

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
1	100	Liquid-Liquid	DCM	None	GC-MS/MS
2	1			Centrifugation & dilution	LC-MS/MS
3	100	Liquid-Liquid	DCM	None	GC-MS
4	500	SPE	DCM	None	GC-MS
5	20	QuEChERS	acetonitrile	None	LC-MS/MS
6	100	Liquid-Liquid	DCM	None	GC-MS
7	100	Liquid-Liquid	DCM	Filtration	GC-MS
8	100	Liquid-Liquid	DCM	None	GC-MS
10	50	Liquid-Liquid	DCM	N/A	GC-MS/MS
11	500	SPE	DCM-EToAC	None	GC-MS/MS
12	10	Direct Injection		Centrifuge	LC-MS/MS

Table 44 Methodology – Sample S4 Dicamba

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
1		NT			
2	1			Filtration & dilution	LC-MS/MS
3		NT			
4		NT			
5		NT			
6	10	Direct Injection	NA	0.2 micron Filter	LC-MS
7		NT			
8		NT			
10	5	Liquid-Liquid	Toluene	N/A	GC-MS
11		Direct Injection			LC-MS/MS
12	10	Direct Injection		Centrifuge	LC-MS/MS

Table 45 Methodology – Sample S4 Dieldrin

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
1	100	Liquid-Liquid	DCM	None	GC-MS/MS
2	3	Liquid-Liquid	DCM	None	GC-MS/MS
3	100	Liquid-Liquid	DCM	None	GC-MS
4	500	SPE	DCM	None	GC-MS
5	40	Liquid-Liquid	Hexane	None	GC-MS/MS
6	100	Liquid-Liquid	DCM	None	GC-MS
7	100	Liquid-Liquid	DCM	Filtration	GC-MS
8	100	Liquid-Liquid	DCM	None	GC-MS
10	50	Liquid-Liquid	DCM	N/A	GC-ECD
11	500	SPE	DCM-EToAC	None	GC-MS
12	500	Liquid-Liquid	DCM	None	GC-MS/MS

Table 46 Methodology – Sample S4 Ethion

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
1	100	Liquid-Liquid	DCM	None	GC-MS/MS
2	1			Centrifugation & dilution	LC-MS/MS
3	100	Liquid-Liquid	DCM	None	GC-MS
4	500	SPE	DCM	None	GC-MS
5		NT			
6	100	Liquid-Liquid	DCM	None	GC-MS
7	100	Liquid-Liquid	DCM	Filtration	GC-MS

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
8	100	Liquid-Liquid	DCM	None	GC-MS
10	50	Liquid-Liquid	DCM	N/A	GC-MS/MS
11	NT				
12	10	Direct Injection		Centrifuge	LC-MS/MS

Table 47 Methodology – Sample S4 Lindane

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
1	100	Liquid-Liquid	DCM	None	GC-MS/MS
2	3	Liquid-Liquid	DCM	None	GC-MS/MS
3	100	Liquid-Liquid	DCM	None	GC-MS
4	500	SPE	DCM	None	GC-MS
5	40	Liquid-Liquid	Hexane	None	GC-MS/MS
6	100	Liquid-Liquid	DCM	None	GC-MS
7	100	Liquid-Liquid	DCM	Filtration	GC-MS
8	100	Liquid-Liquid	DCM	None	GC-MS
10	50	Liquid-Liquid	DCM	N/A	GC-ECD
11	500	SPE	DCM-EToAC	None	GC-MS
12	500	Liquid-Liquid	DCM	None	GC-MS/MS

Table 48 Methodology – Sample S4 Simazine

Lab. Code	Sample Volume (mL)	Extraction	Extraction Solvent	Clean-Up	Measurement Instrument
1	100	Liquid-Liquid	DCM	None	GC-MS/MS
2	NT				
3	100	Liquid-Liquid	DCM	None	GC-MS
4	500	SPE	DCM	None	GC-MS
5	20	QuEChERS	acetonitrile	None	LC-MS/MS
6	100	Liquid-Liquid	DCM	None	GC-MS
7	100	Liquid-Liquid	DCM	Filtration	GC-MS
8	NT				
10	50	Liquid-Liquid	DCM	N/A	GC-MS/MS
11		Direct Injection			LC-MS/MS
12	10	Direct Injection		Centrifuge	LC-MS/MS

APPENDIX 5 ACRONYMS AND ABBREVIATIONS

2,4,5-T	2,4,5-Trichlorophenoxyacetic acid
2,4-D	2,4-Dichlorophenoxyacetic acid
ACN	Acetonitrile
AV	Assigned Value
BTEX	Benzene, Toluene, Ethylbenzene and Xylenes
CITAC	Cooperation on International Traceability in Analytical Chemistry
CRM	Certified Reference Material
CV	Coefficient of Variation
DCM	Dichloromethane
DDT	Dichlorodiphenyltrichloroethane
DI	Direct Injection
ECD	Electron Capture Detection
EtOAc	Ethyl Acetate
FID	Flame Ionisation Detection
GC	Gas Chromatography
GUM	Guide to the Expression of Uncertainty in Measurement
HEX	Hexane
IEC	International Electrotechnical Commission
ISO	International Organization for Standardization
LC	Liquid Chromatography
LLE	Liquid-Liquid Extraction
LOR	Limit of Reporting
Max	Maximum
MCPA	2-methyl-4-chlorophenoxyacetic acid
Md	Median
MeOH	Methanol
Min	Minimum
MS	Mass Spectrometry
MS/MS	Tandem Mass Spectrometry
MU	Measurement Uncertainty
N	Number of numeric results
NA	Not Applicable
NATA	National Association of Testing Authorities, Australia
NMI	National Measurement Institute, Australia

NR	Not Reported
NT	Not Tested
P&T	Purge-and-Trap
PAH	Polycyclic Aromatic Hydrocarbon
PCV	Performance Coefficient of Variation
PT	Proficiency Testing
QuEChERS	Quick, Easy, Cheap, Effective, Rugged and Safe extraction method
RA	Robust Average
RM	Reference Material
SD	Standard Deviation
SI	International System of Units
SPE	Solid Phase Extraction
SS	Spiked Samples
SV	Spiked Value (or formulated concentration of a PT sample)
TRH	Total Recoverable Hydrocarbons
U	Expanded Uncertainty

END OF REPORT